

# D. Excitable Media

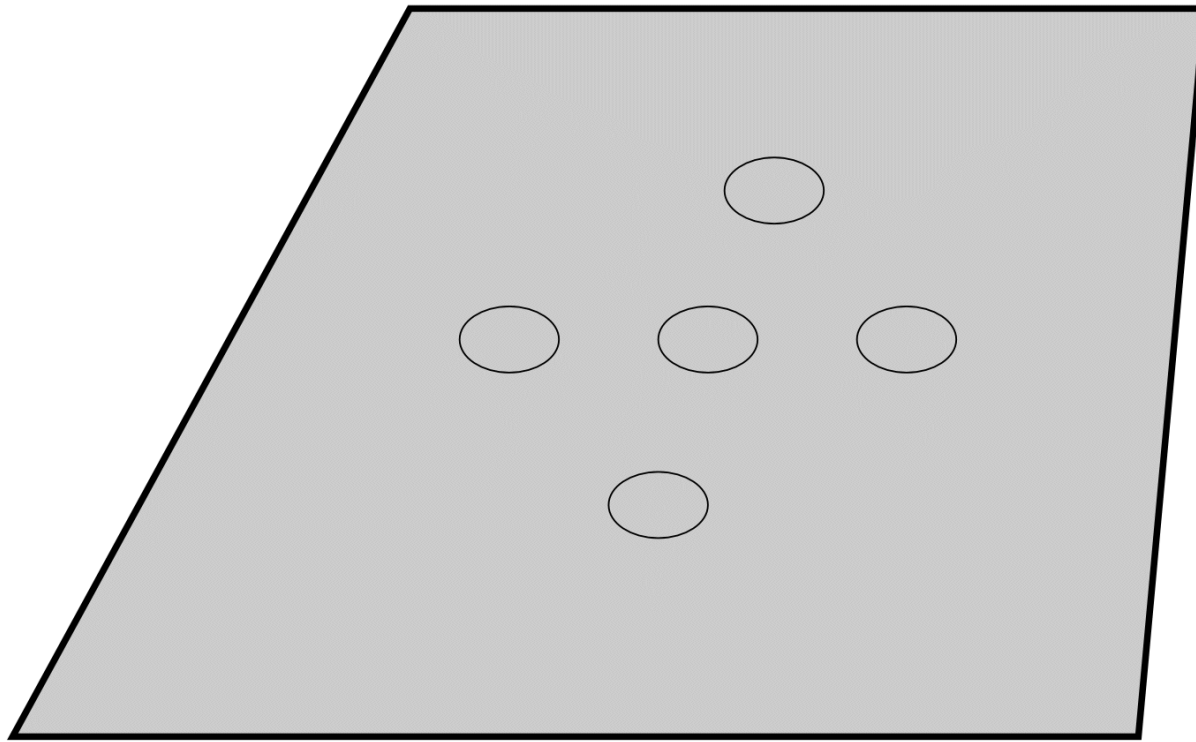
# Examples of Excitable Media

- Slime mold amoebas
- Cardiac tissue (& other muscle tissue)
- Cortical tissue
- Certain chemical systems (e.g., BZ reaction)
- Hodgepodge machine

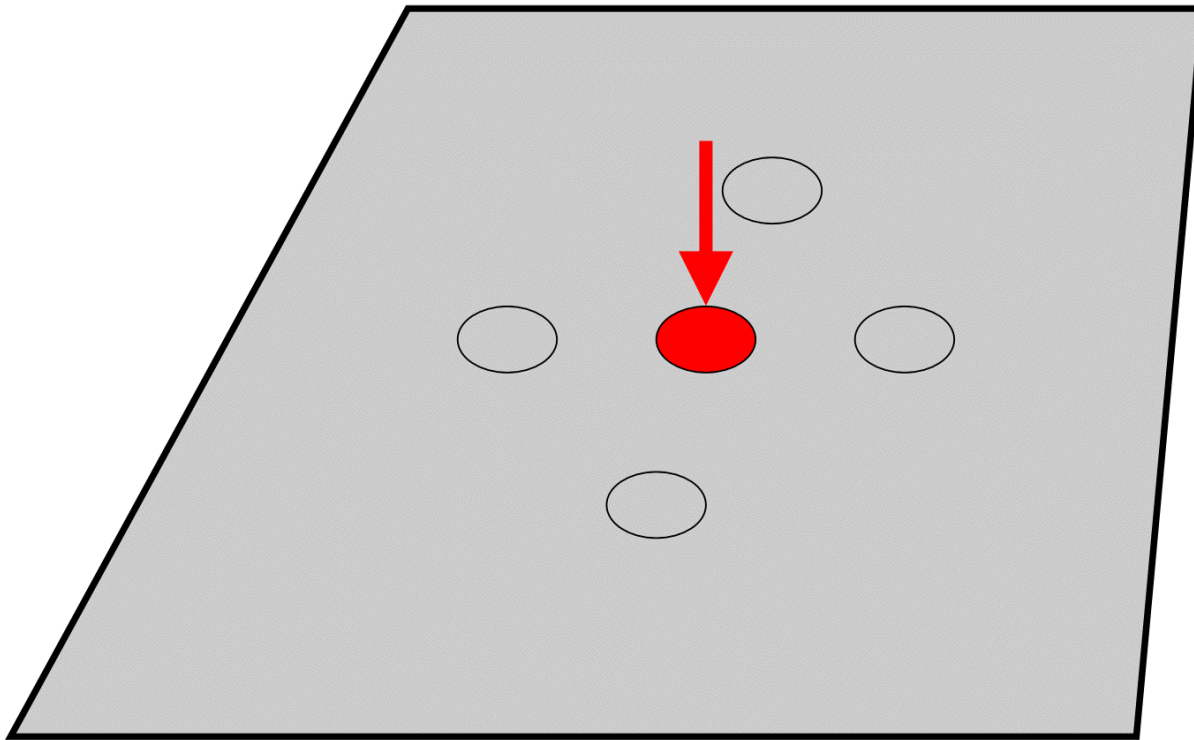
# Characteristics of Excitable Media

- Local spread of excitation
  - for signal propagation
- Refractory period
  - for unidirectional propagation
- Decay of signal
  - avoid saturation of medium

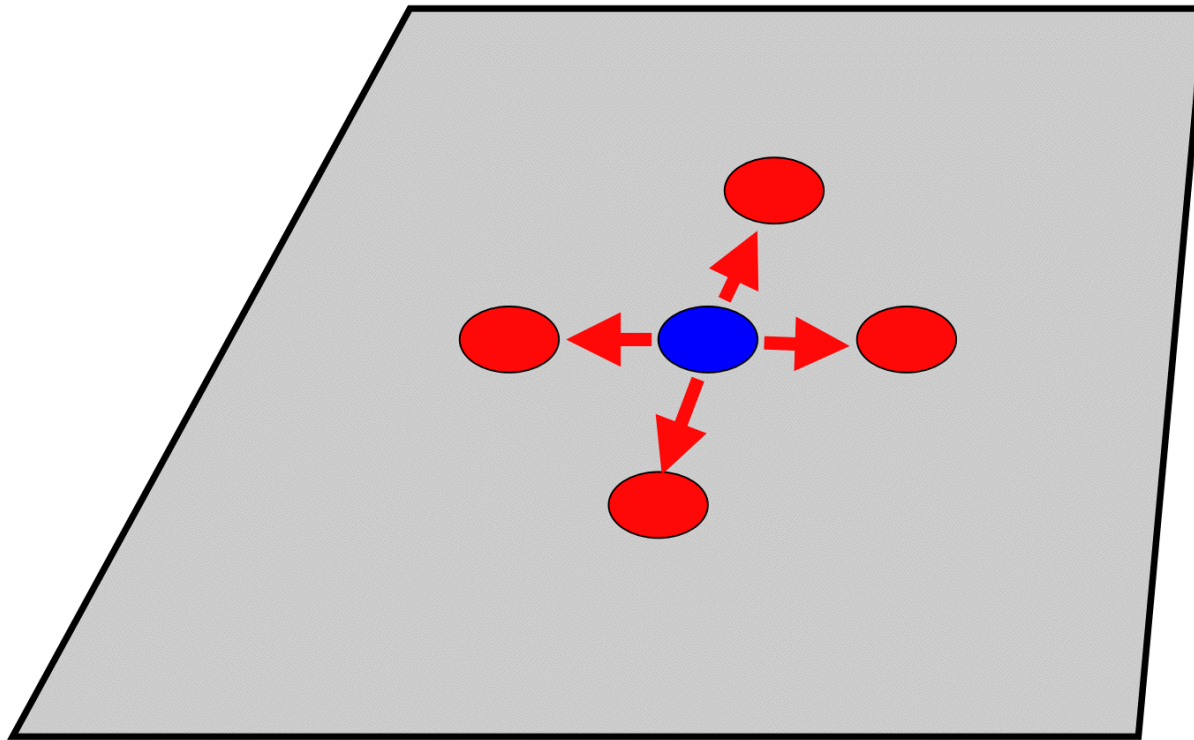
# Behavior of Excitable Media



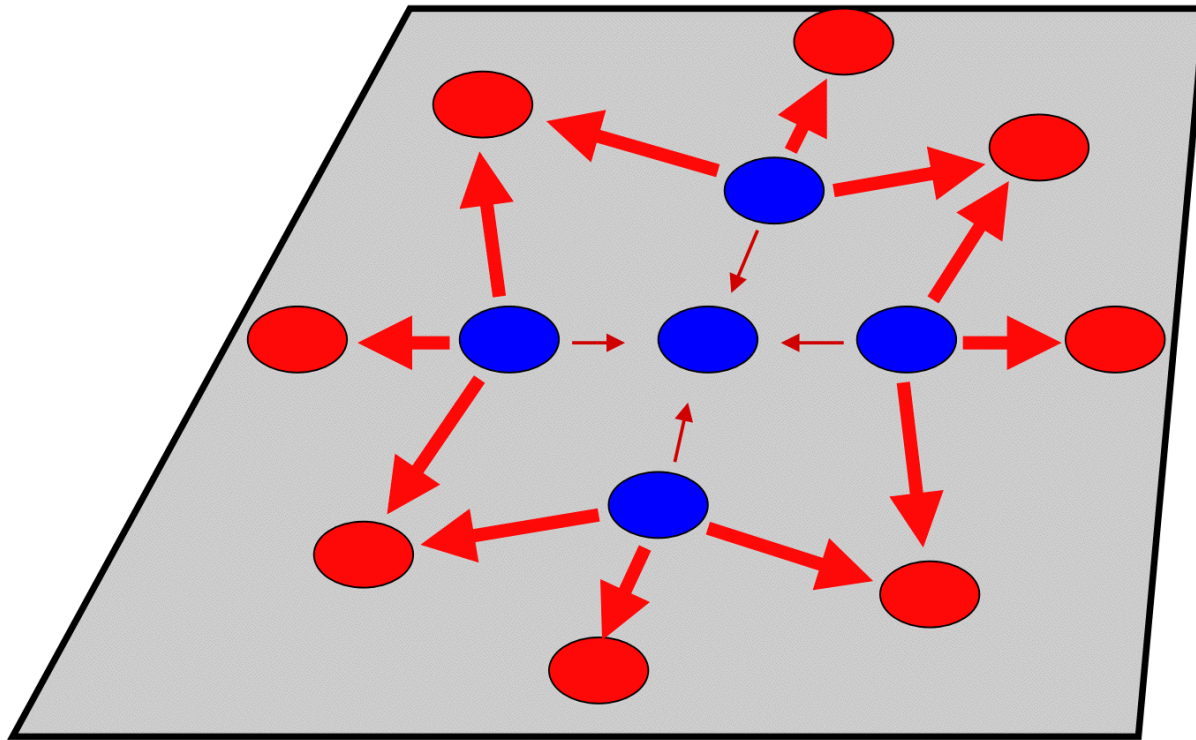
# Stimulation



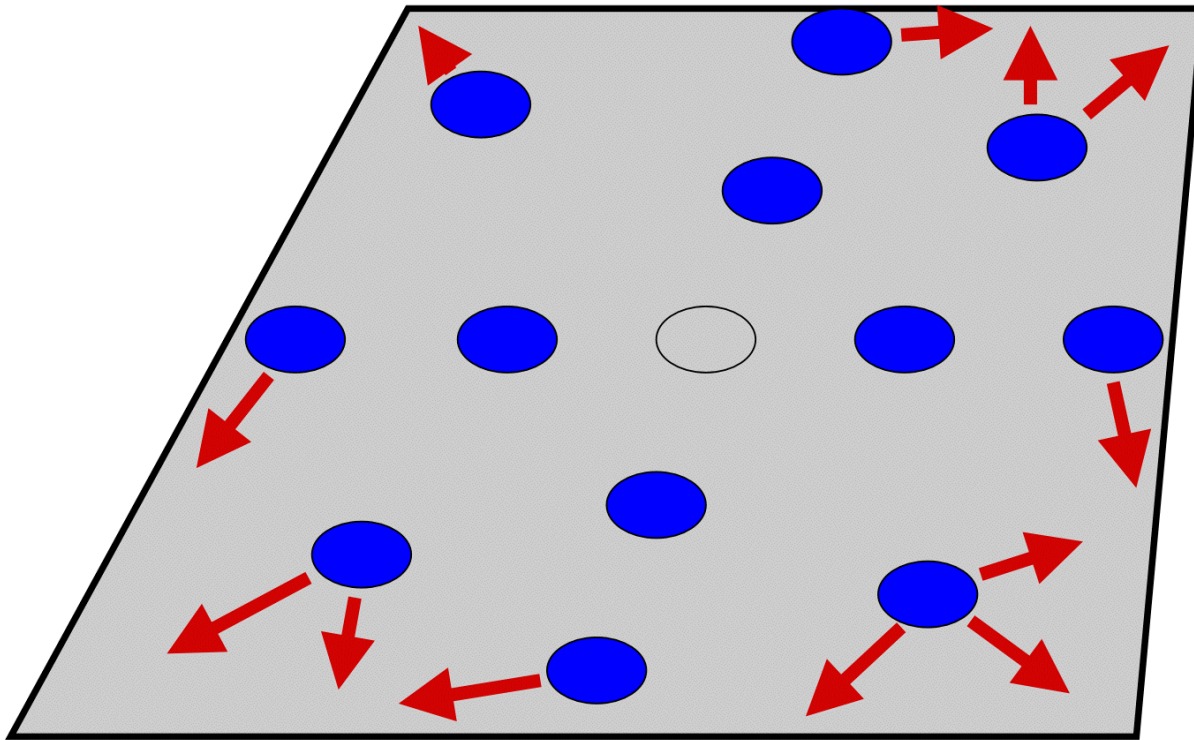
# Relay (Spreading Excitation)



# Continued Spreading

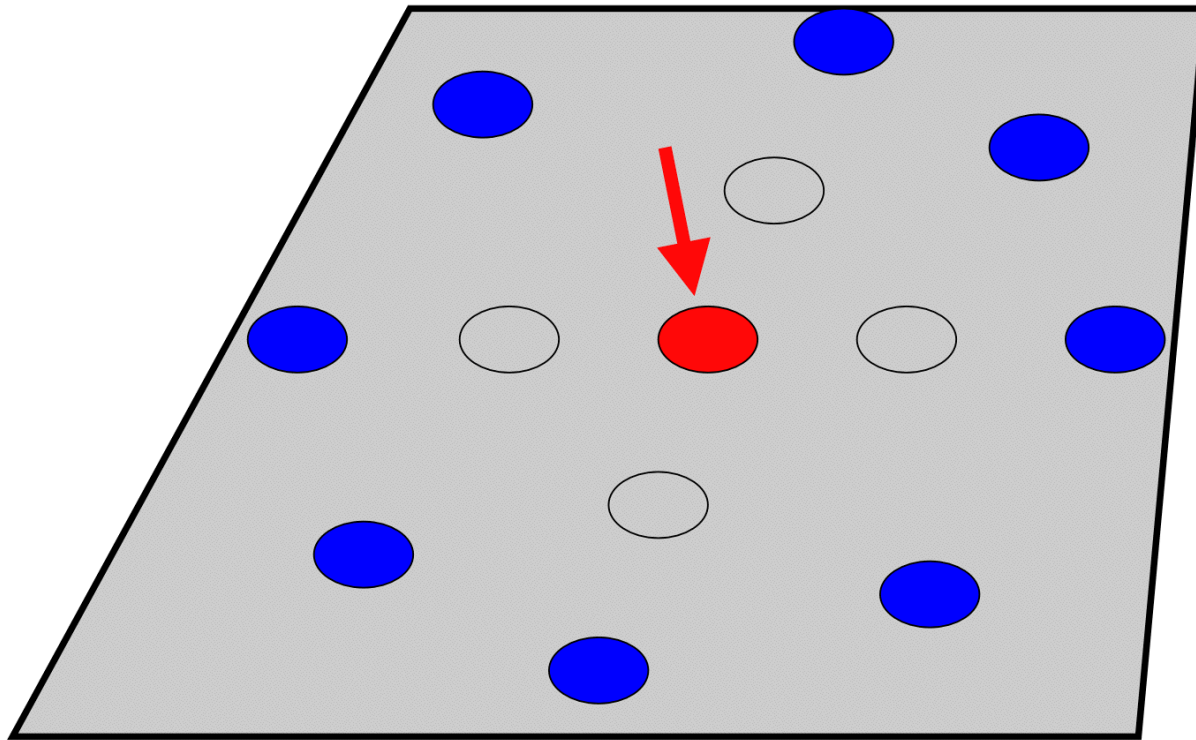


# Recovery





# Restimulation



# Circular & Spiral Waves

## Observed in:

- Slime mold aggregation
- Chemical systems (e.g., BZ reaction)
- Neural tissue
- Retina of the eye
- Heart muscle
- Intracellular calcium flows
- Mitochondrial activity in oocytes

# Cause of Concentric Circular Waves

- Excitability is not enough
- But at certain developmental stages, cells can operate as pacemakers
- When stimulated by cAMP, they begin emitting regular pulses of cAMP

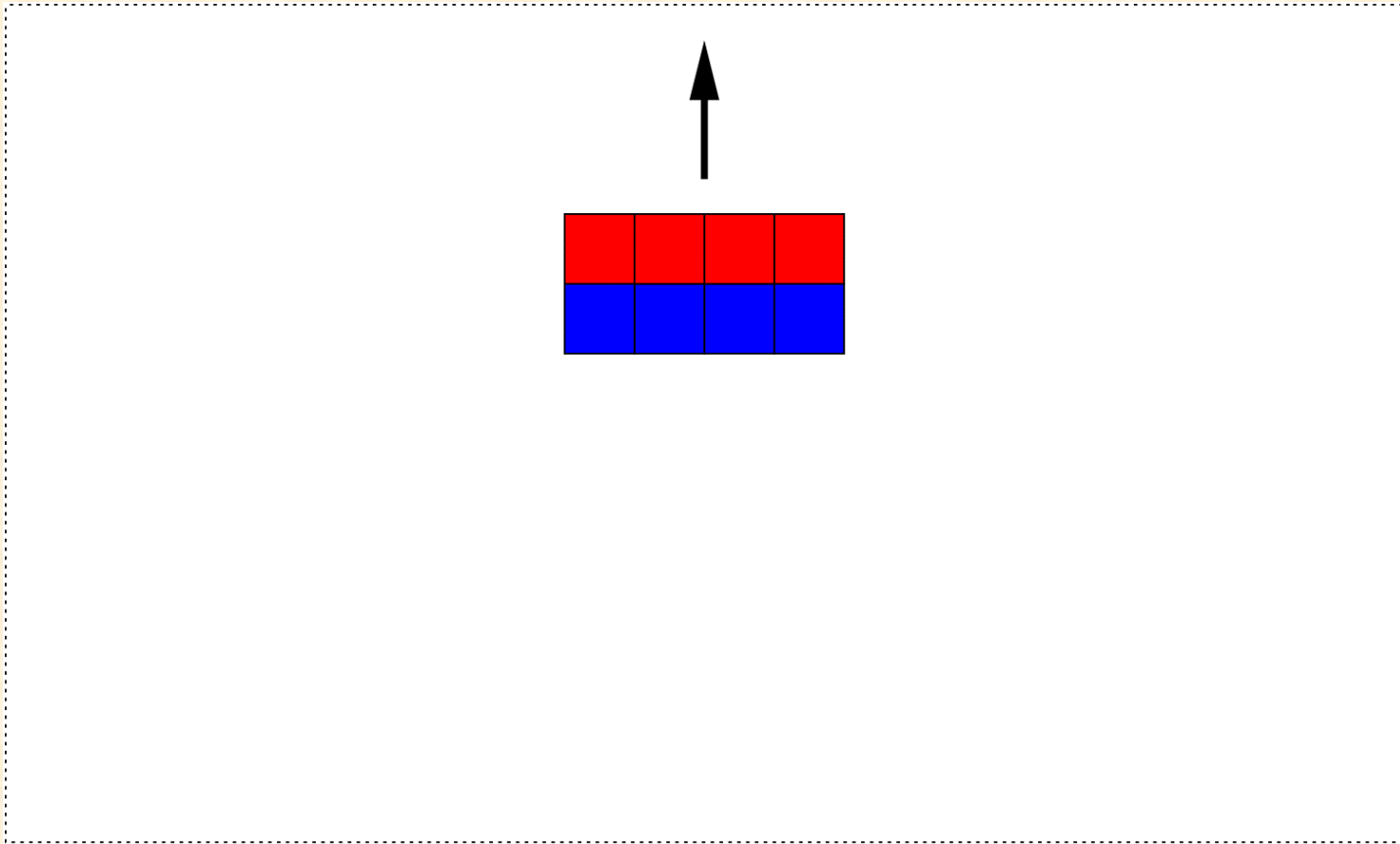
# Spiral Waves

- Persistence & propagation of spiral waves explained analytically (Tyson & Murray, 1989)
- Rotate around a small core of non-excitable cells
- Propagate at higher frequency than circular
- Therefore they dominate circular in collisions
- But how do the spirals form initially?

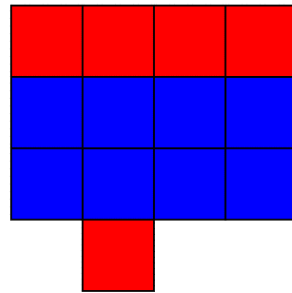
# Some Explanations of Spiral Formation

- “the origin of spiral waves remains obscure” (1997)
- Traveling wave meets obstacle and is broken
- Desynchronization of cells in their developmental path
- Random pulse behind advancing wave front

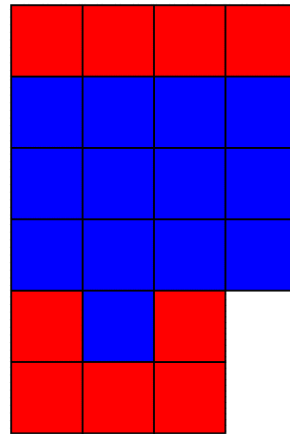
# Step 0: Passing Wave Front



# Step 1: Random Excitation

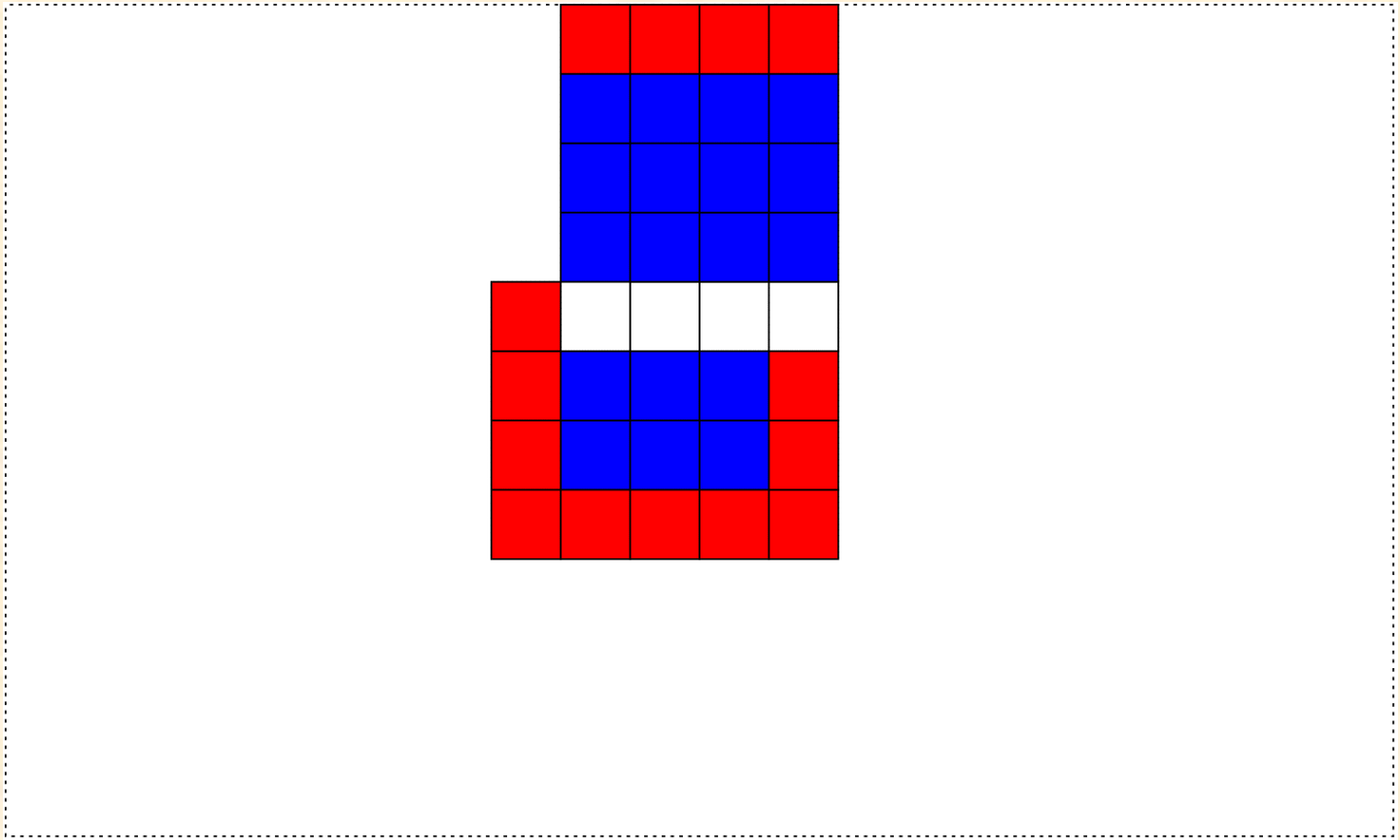


# Step 2: Beginning of Spiral

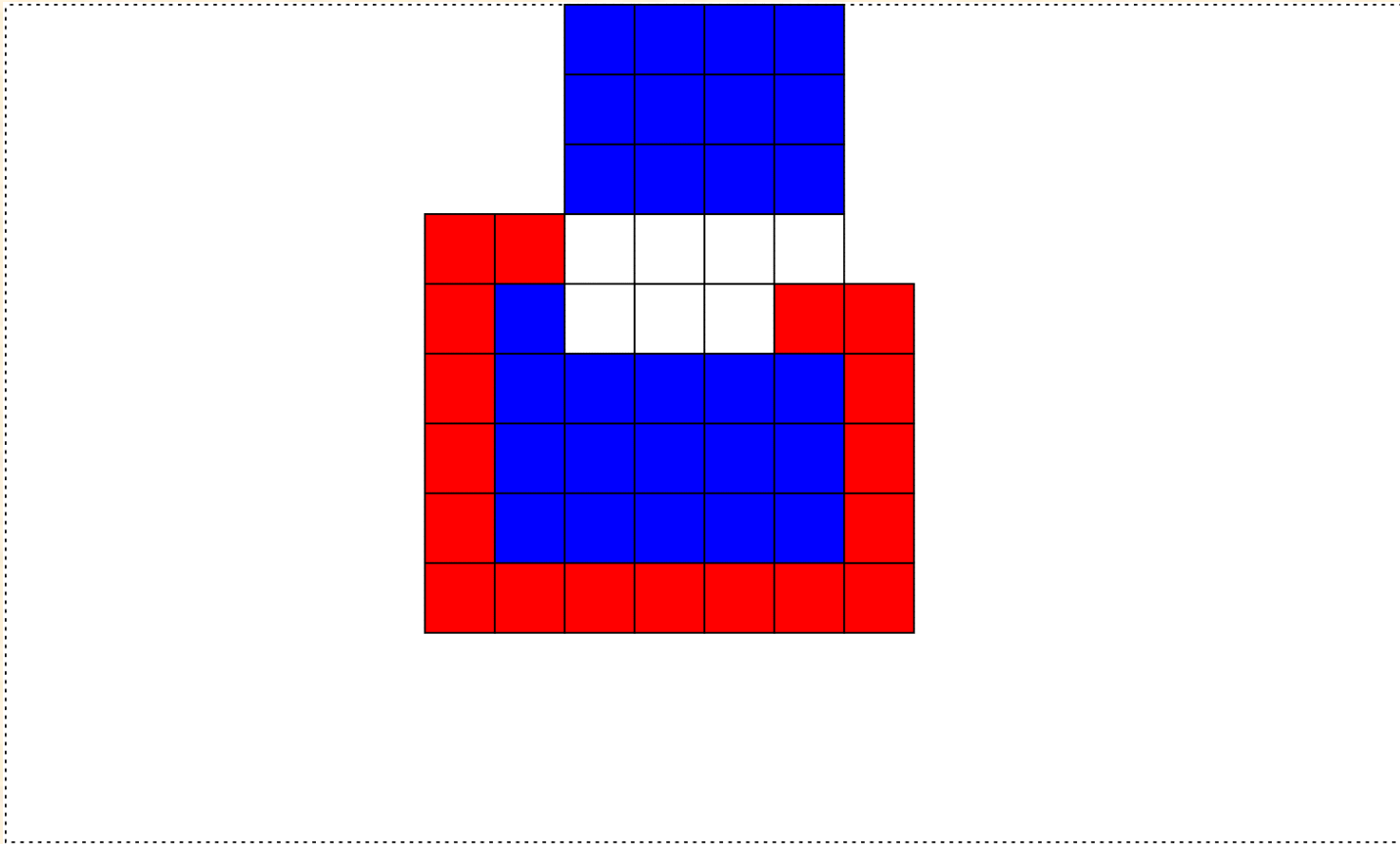




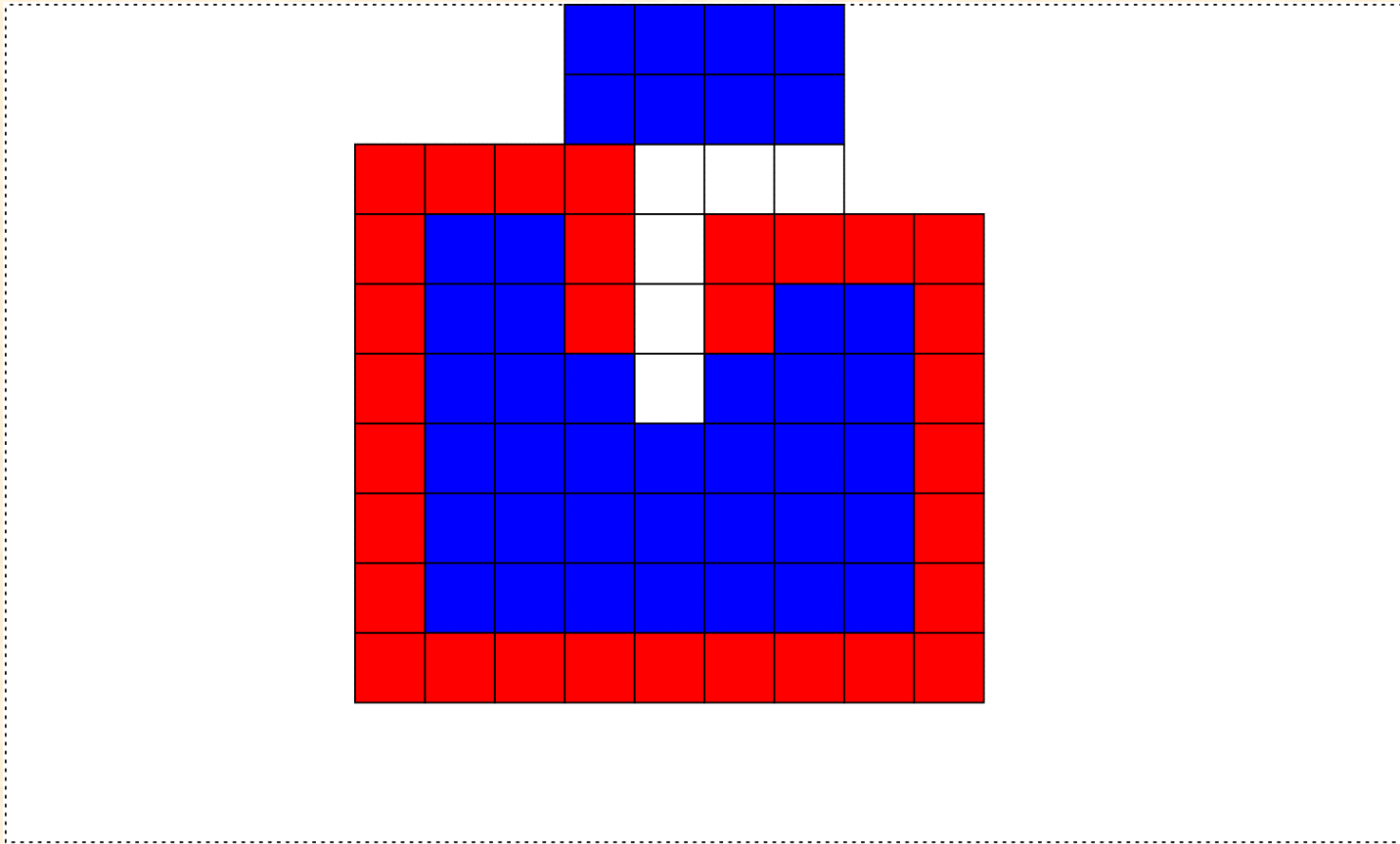
# Step 3



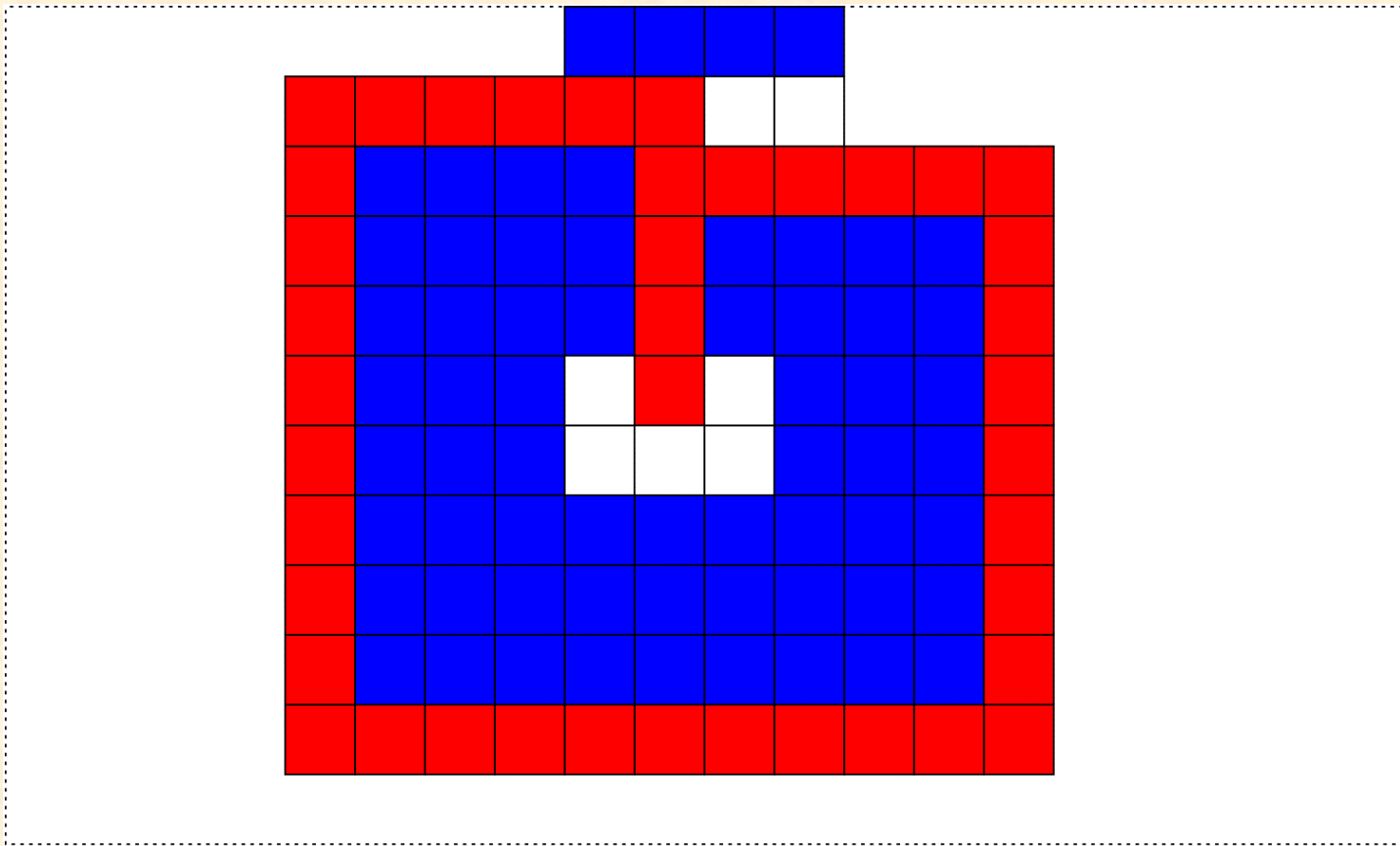
# Step 4



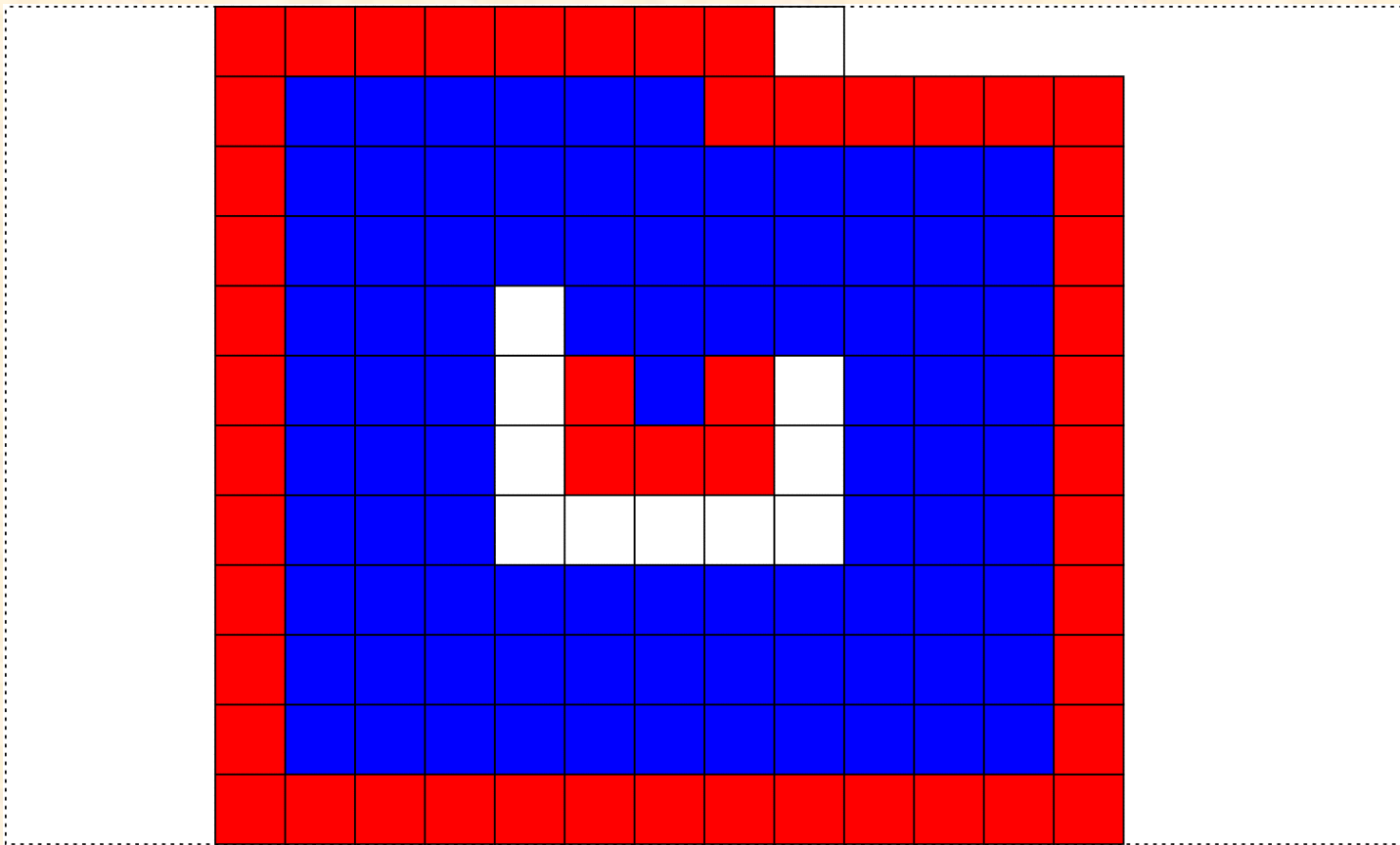
# Step 5



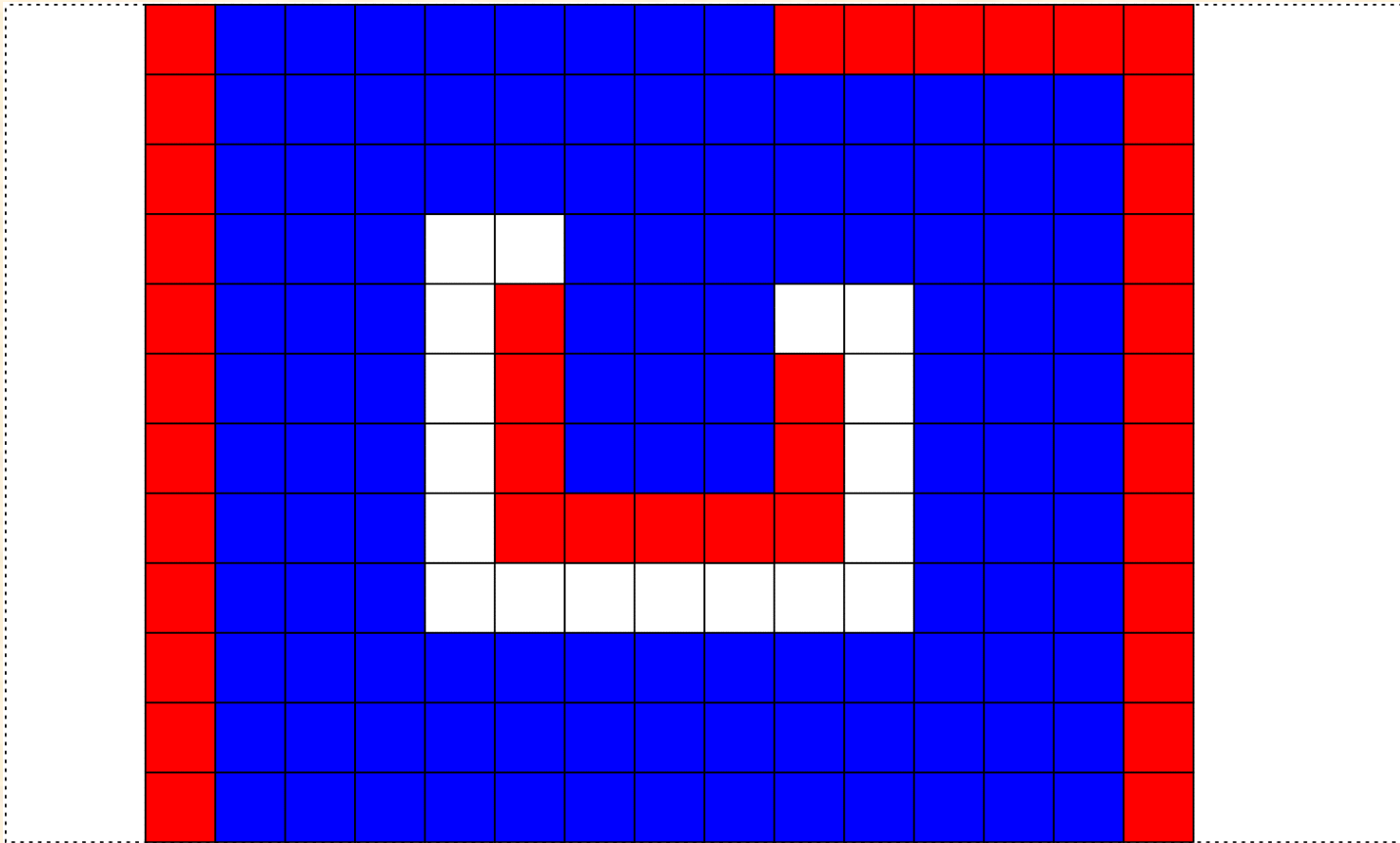
# Step 6: Rejoining & Reinitiation



# Step 7: Beginning of New Spiral

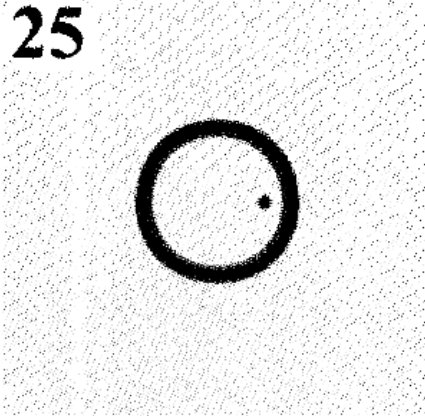


# Step 8

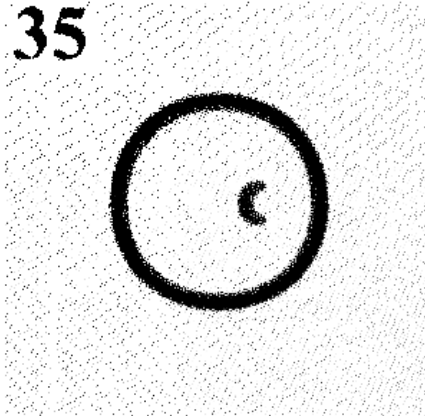


# Formation of Double Spiral

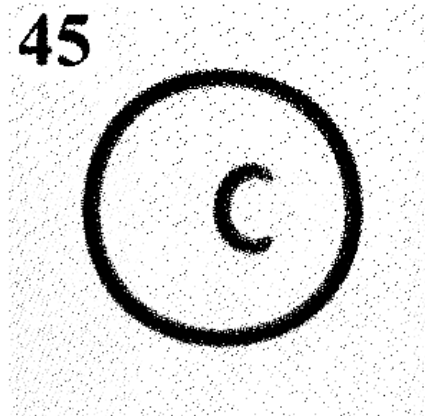
25



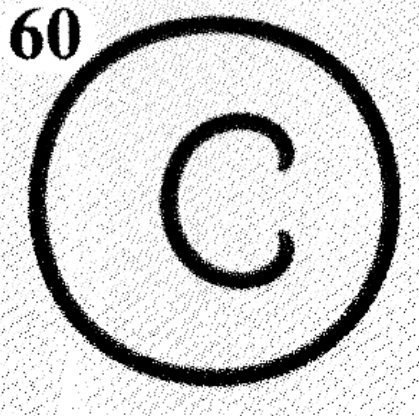
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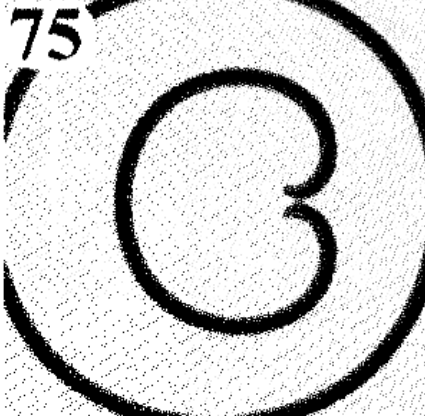
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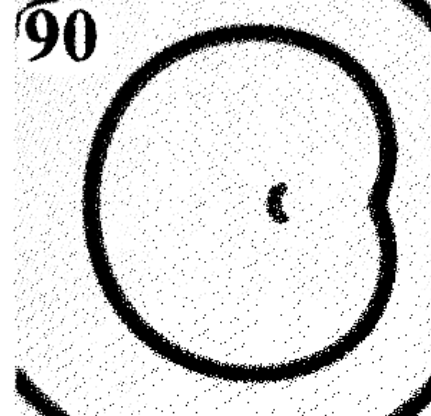
60



75



90



# NetLogo Simulation Of Spiral Formation

- Amoebas are immobile at timescale of wave movement
- A fraction of patches are inert (grey)
- A fraction of patches has initial concentration of cAMP
- At each time step:
  - chemical diffuses
  - each patch responds to local concentration



# Response of Patch

**if** patch is not refractory (**brown**) **then**

**if** local chemical  $>$  threshold **then**

set refractory period

produce pulse of chemical (**red**)

**else**

decrement refractory period

degrade chemical in local area

# Demonstration of NetLogo Simulation of Spiral Formation

[Run SlimeSpiral.nlogo](#)

# Demonstration of NetLogo Simulation of Spiral Formation (a closer look)

[Run SlimeSpiralBig.nlogo](#)

# Observations

- Excitable media can support circular and spiral waves
- Spiral formation can be triggered in a variety of ways
- All seem to involve inhomogeneities (broken symmetries):
  - in space
  - in time
  - in activity
- Amplification of random fluctuations
- Circles & spirals are to be expected

# NetLogo Simulation of Streaming Aggregation

1. chemical diffuses
2. **if** cell is refractory (**yellow**)
3. **then** chemical degrades
4. **else** (it's excitable, colored white)
  1. **if** chemical > movement threshold **then**  
take step up chemical gradient
  2. **else if** chemical > relay threshold **then**  
produce more chemical (**red**)  
become refractory
  3. **else** wait

# Demonstration of NetLogo Simulation of Streaming

[Run SlimeStream.nlogo](#)

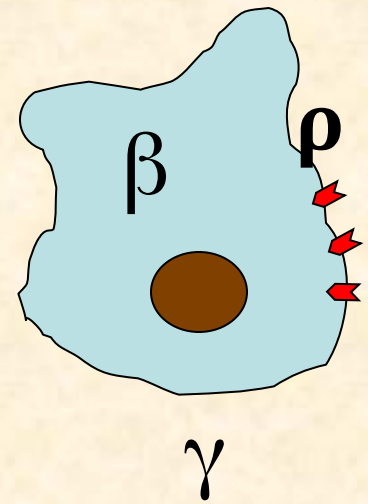
# Modified Martiel & Goldbeter Model for Dicty Signalling

Variables (functions of  $x, y, t$ ):

$\beta$  = intracellular concentration  
of cAMP

$\gamma$  = extracellular concentration  
of cAMP

$\rho$  = fraction of receptors in active state



# Equations

$$\frac{d\beta(x,y,t)}{dt} = s\Phi(\rho,\gamma) - \beta k_i - \beta k_t \quad [1]$$

Rate of change in intracellular [cAMP] = Production of cAMP - Intracellular hydrolysis - Secretion of cAMP

$$\frac{d\gamma(x,y,t)}{dt} = \frac{k_t}{h}\beta - k_e\gamma + D\nabla^2\gamma \quad [2]$$

Rate of change in extracellular [cAMP] = Secretion of cAMP - Extracellular hydrolysis + Diffusion of cAMP

$$\frac{d\rho(x,y,t)}{dt} = f_2(\gamma)(1 - \rho) - f_1(\gamma)\rho \quad [3]$$

Rate of change in fraction of active receptor = Dephosphorylation of receptor - Phosphorylation of receptor



# Positive Feedback Loop

- Extracellular cAMP increases  
( $\gamma$  increases)
- $\Rightarrow$  Rate of synthesis of intracellular cAMP increases  
( $\Phi$  increases)
- $\Rightarrow$  Intracellular cAMP increases  
( $\beta$  increases)
- $\Rightarrow$  Rate of secretion of cAMP increases
- ( $\Rightarrow$  Extracellular cAMP increases)

# Negative Feedback Loop

- Extracellular cAMP increases  
( $\gamma$  increases)
- $\Rightarrow$  cAMP receptors desensitize  
( $f_1$  increases,  $f_2$  decreases,  $\rho$  decreases)
- $\Rightarrow$  Rate of synthesis of intracellular cAMP decreases  
( $\Phi$  decreases)
- $\Rightarrow$  Intracellular cAMP decreases  
( $\beta$  decreases)
- $\Rightarrow$  Rate of secretion of cAMP decreases
- $\Rightarrow$  Extracellular cAMP decreases  
( $\gamma$  decreases)

# Dynamics of Model

- Unperturbed  
⇒ cAMP concentration reaches steady state
- Small perturbation in extracellular cAMP  
⇒ returns to steady state
- Perturbation  $>$  threshold ⇒
  - large transient in cAMP, and then return to steady state
  - *or* oscillation (depending on model parameters)

# Typical Equations for Excitable Medium (ignoring diffusion)

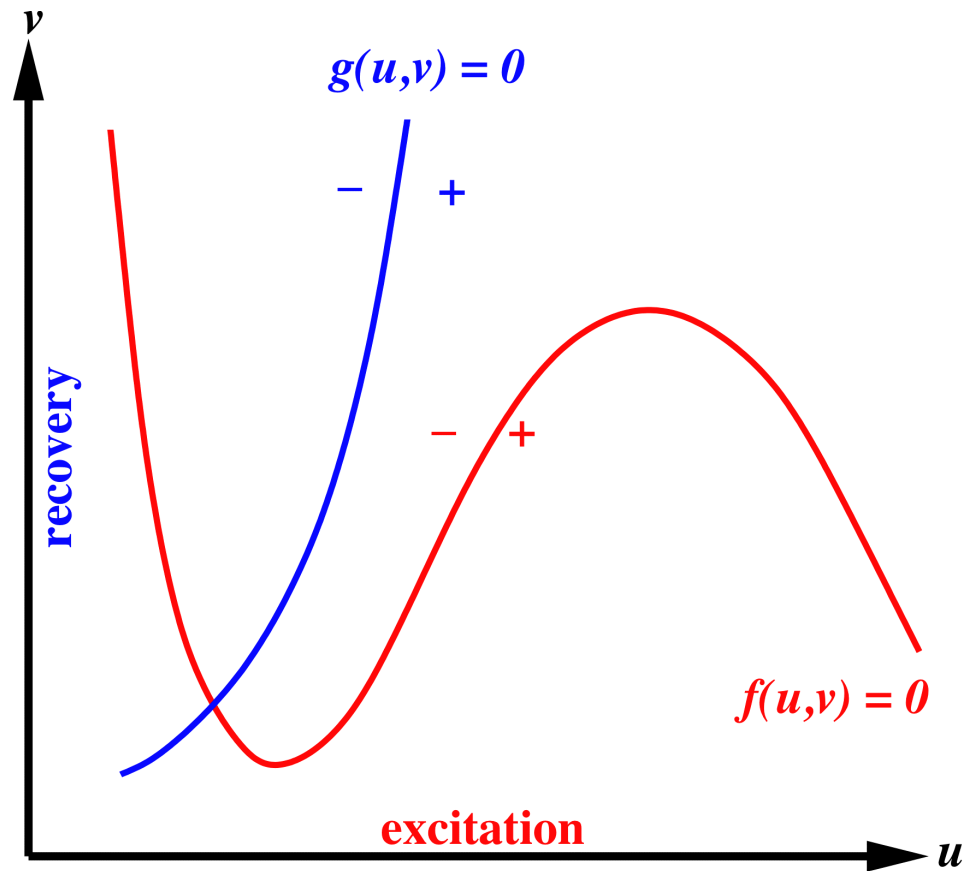
- Excitation variable:

$$\dot{u} = f(u, v)$$

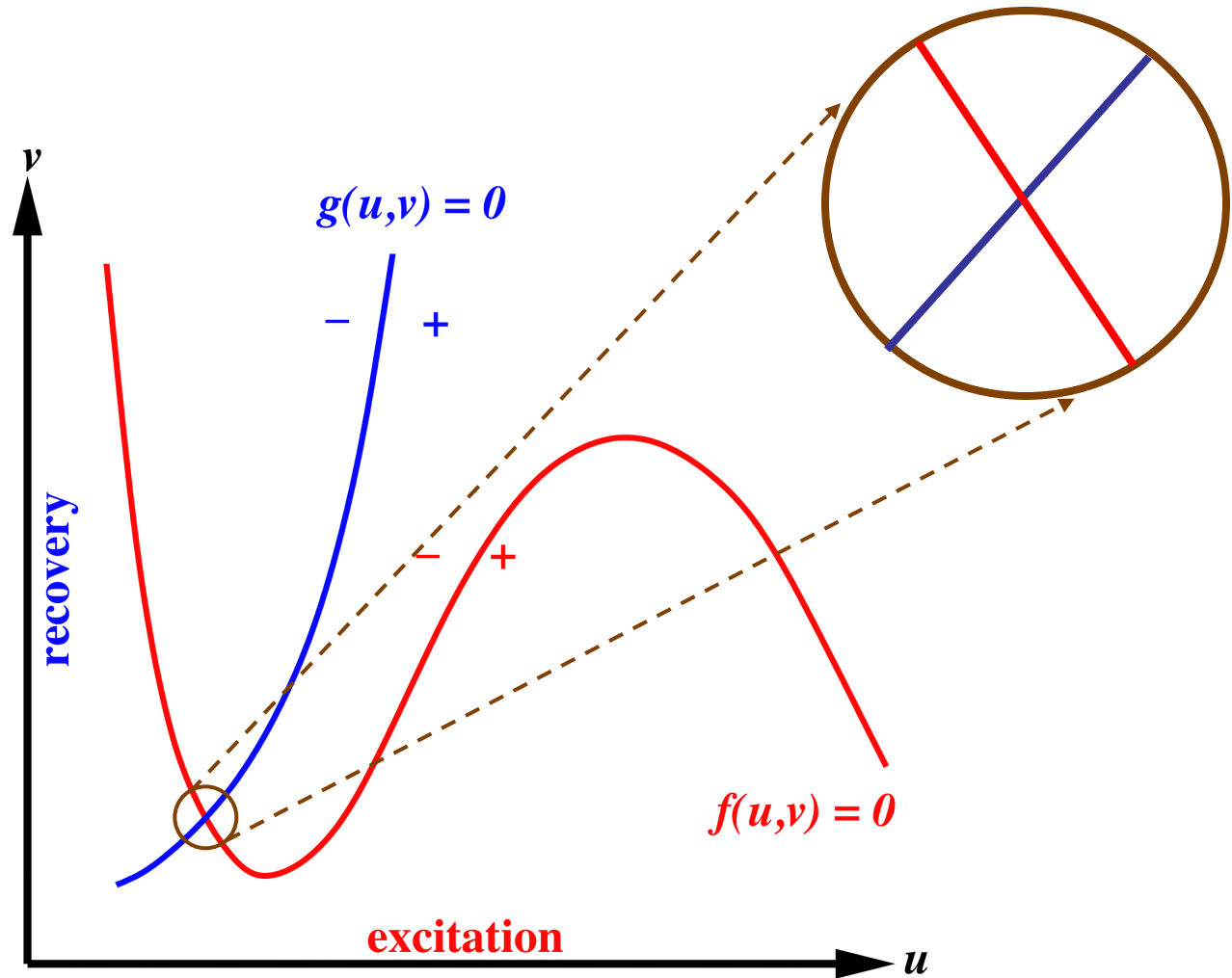
- Recovery variable:

$$\dot{v} = g(u, v)$$

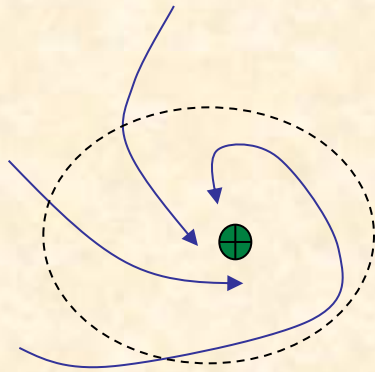
# Nullclines



# Local Linearization

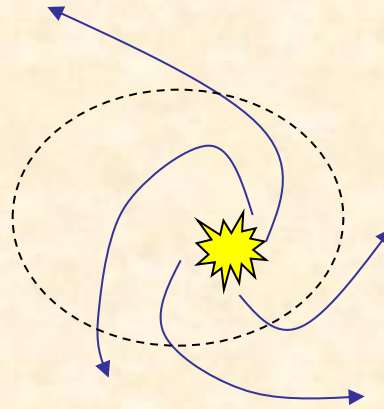


# Fixed Points & Eigenvalues



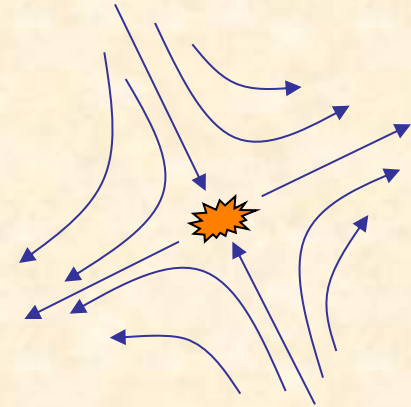
**stable  
fixed point**

real parts of  
eigenvalues  
are negative



**unstable  
fixed point**

real parts of  
eigenvalues  
are positive



**saddle point**

one positive real &  
one negative real  
eigenvalue

# Neural Impulse Propagation

$$C \frac{dv}{dt} = I - g_{Na} m^3 h (V - V_{Na}) - g_K n^4 (V - V_K) - g_L (V - V_L)$$

$$\frac{dm}{dt} = a_m(V)(1 - m) - b_m(V)m$$

$$\frac{dh}{dt} = a_h(V)(1 - h) - b_h(V)h$$

$$\frac{dn}{dt} = a_n(V)(1 - n) - b_n(V)n$$

$$a_m(V) = .1(V + 40)/(1 - \exp(-(V + 40)/10))$$

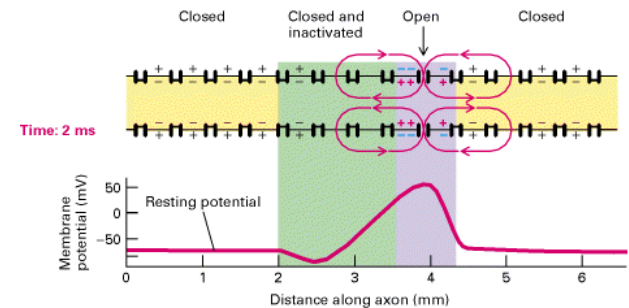
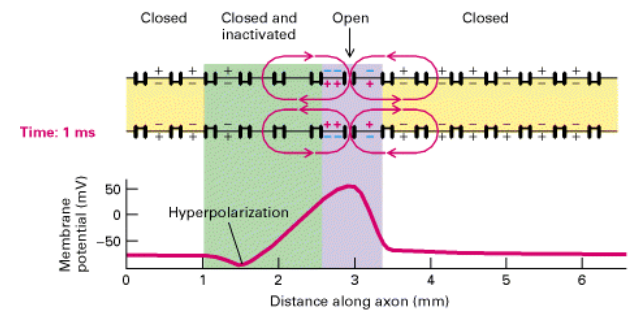
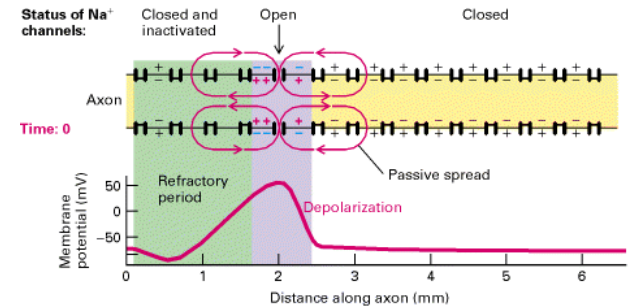
$$b_m(V) = 4 \exp(-(V + 65)/18)$$

$$a_h(V) = .07 \exp(-(V + 65)/20)$$

$$b_h(V) = 1/(1 + \exp(-(V + 35)/10))$$

$$a_n(V) = .01(V + 55)/(1 - \exp(-(V + 55)/10))$$

$$b_n(V) = .125 \exp(-(V + 65)/80)$$



👉 Hodgkin-Huxley equations



# FitzHugh-Nagumo Model

- A simplified model of action potential generation in neurons
- The neuronal membrane is an excitable medium
- $B$  is the input bias:

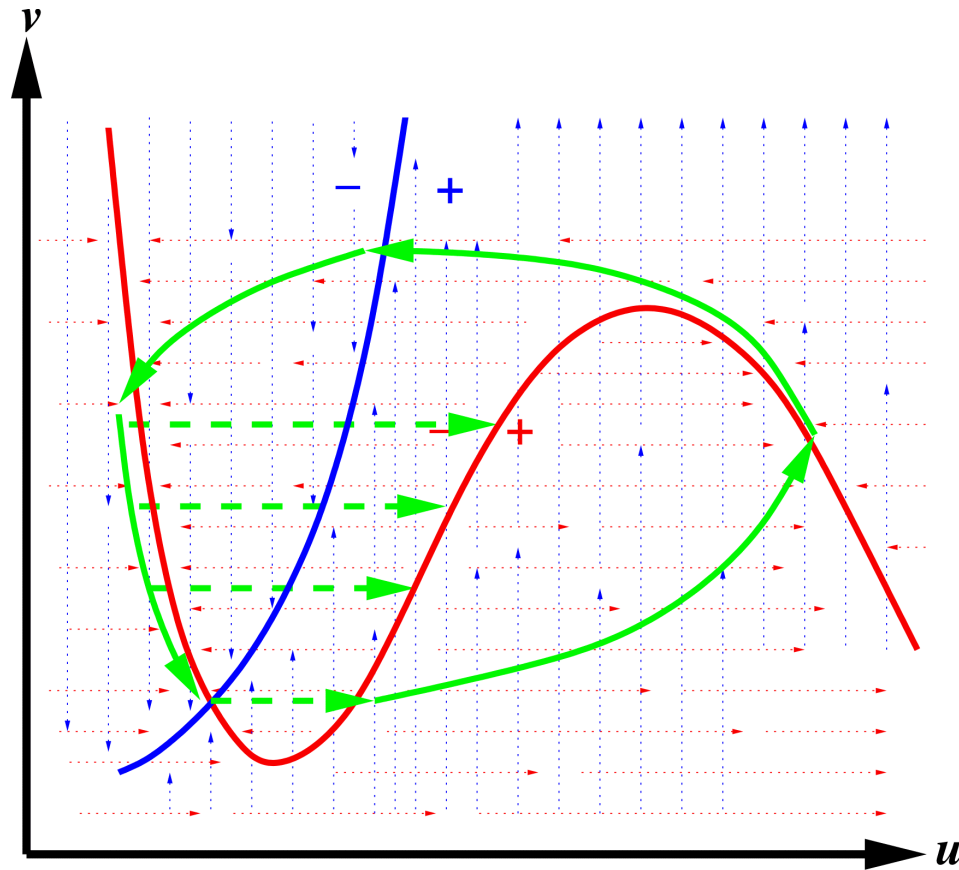
$$\dot{u} = u - \frac{u^3}{3} - v + B$$

$$\dot{v} = \varepsilon(b_0 + b_1 u - v)$$

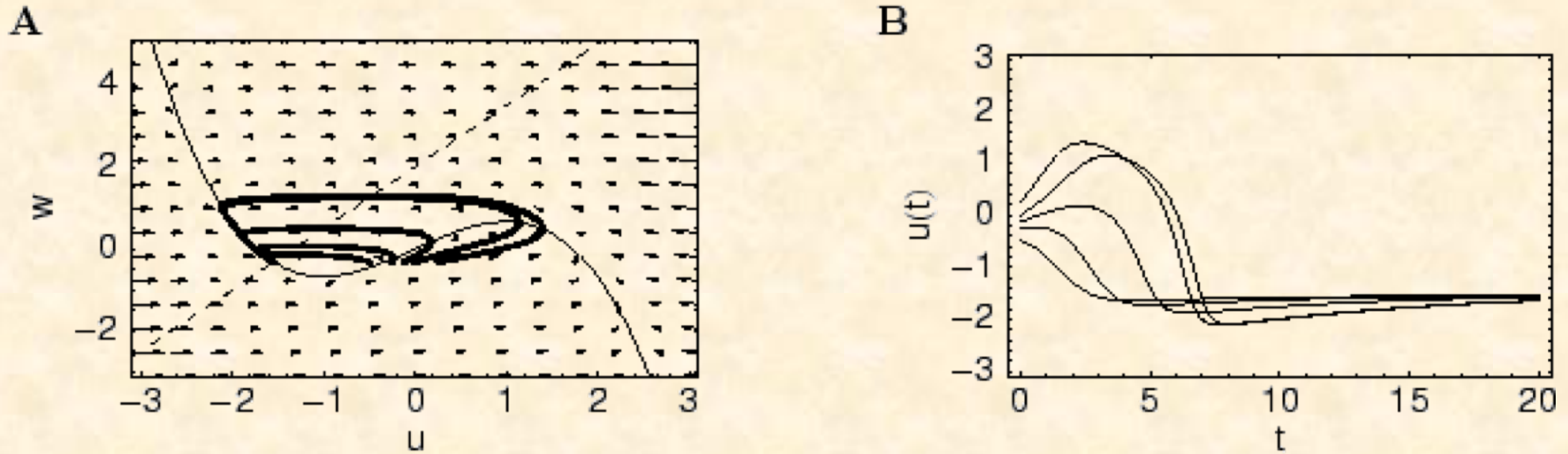
NetLogo Simulation of  
Excitable Medium  
in 2D Phase Space

(EM-Phase-Plane.nlogo)

# Elevated Thresholds During Recovery

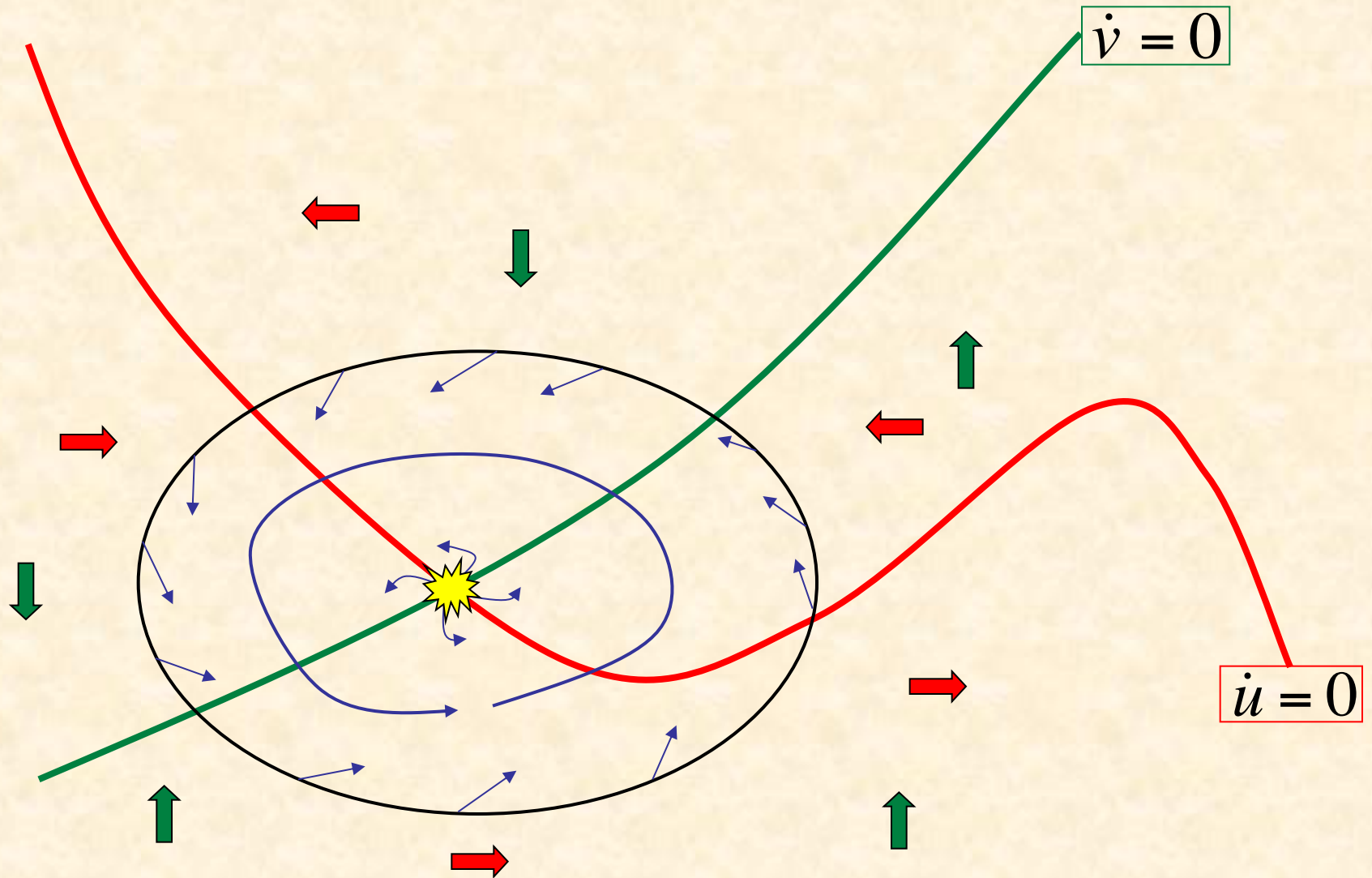


# Type II Model

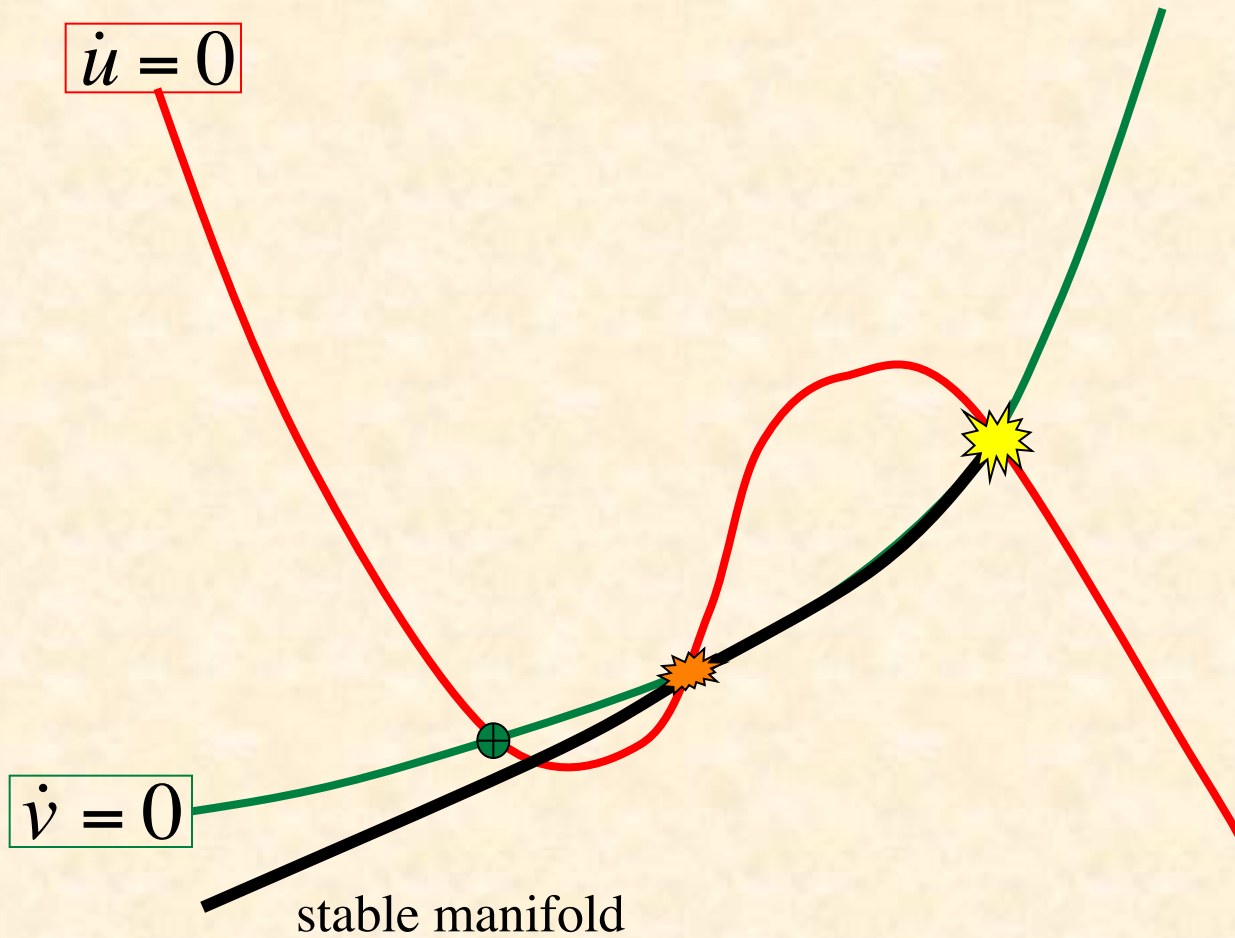


- Soft threshold with critical regime
- Bias can destabilize fixed point

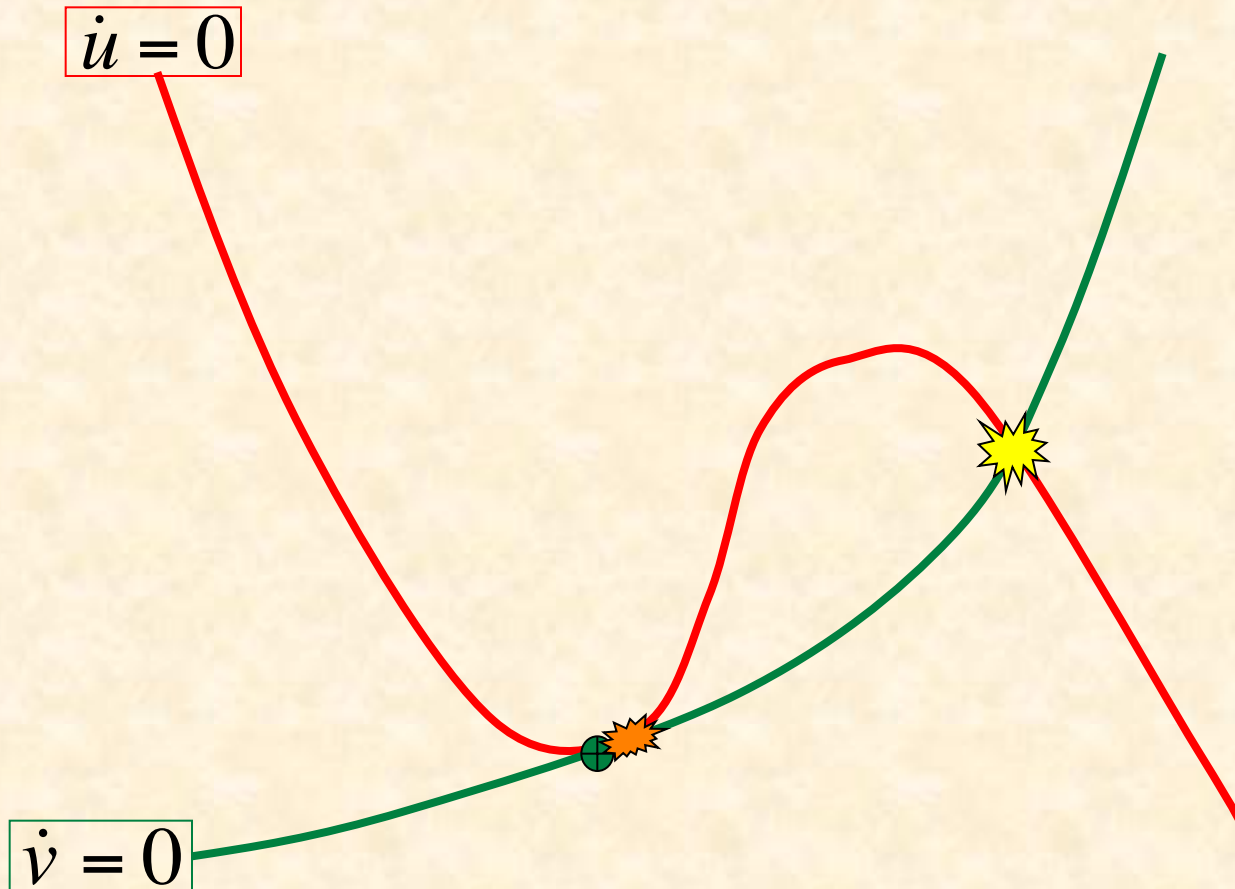
# Poincaré-Bendixson Theorem



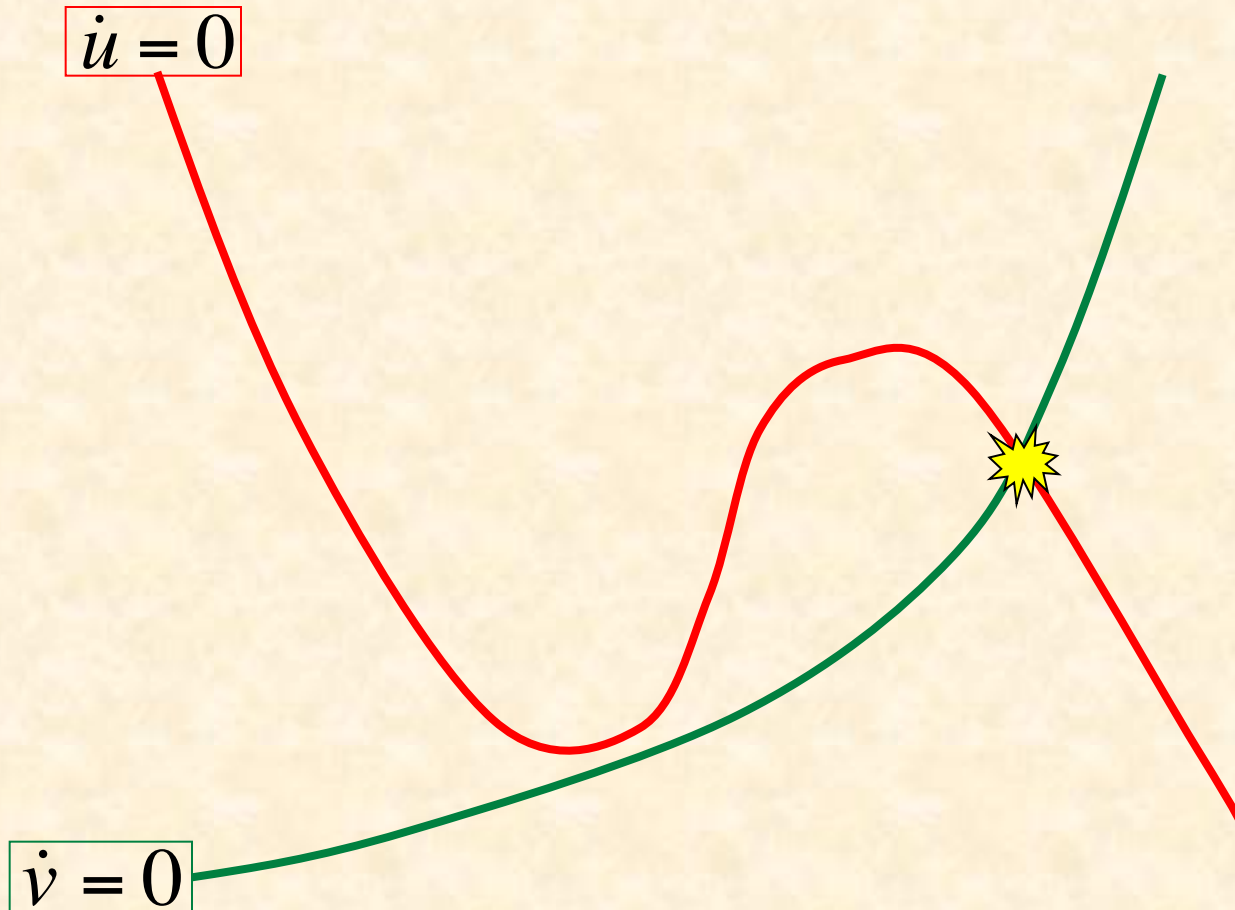
# Type I Model



# Type I Model (Elevated Bias)

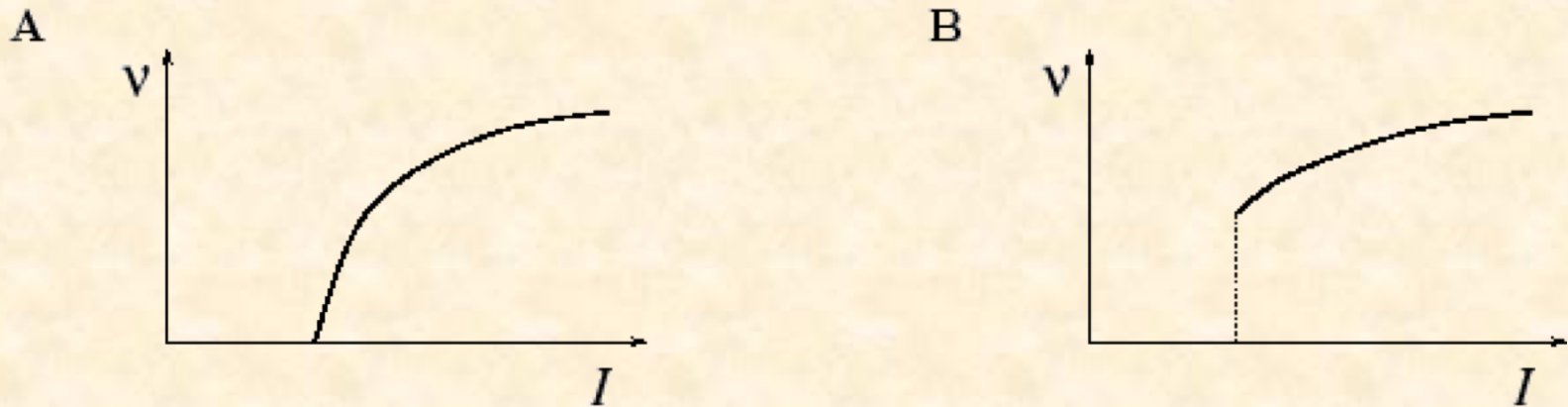


# Type I Model (Elevated Bias 2)





# Type I vs. Type II



- Continuous vs. threshold behavior of frequency
- Slow-spiking vs. fast-spiking neurons

fig. < Gerstner & Kistler

# Additional Bibliography

1. Kessin, R. H. *Dictyostelium: Evolution, Cell Biology, and the Development of Multicellularity*. Cambridge, 2001.
2. Gerhardt, M., Schuster, H., & Tyson, J. J. “A Cellular Automaton Model of Excitable Media Including Curvature and Dispersion,” *Science* **247** (1990): 1563-6.
3. Tyson, J. J., & Keener, J. P. “Singular Perturbation Theory of Traveling Waves in Excitable Media (A Review),” *Physica D* **32** (1988): 327-61.
4. Camazine, S., Deneubourg, J.-L., Franks, N. R., Sneyd, J., Theraulaz, G., & Bonabeau, E. *Self-Organization in Biological Systems*. Princeton, 2001.
5. Pálsson, E., & Cox, E. C. “Origin and Evolution of Circular Waves and Spiral in *Dictyostelium discoideum* Territories,” *Proc. Natl. Acad. Sci. USA*: **93** (1996): 1151-5.
6. Solé, R., & Goodwin, B. *Signs of Life: How Complexity Pervades Biology*. Basic Books, 2000.