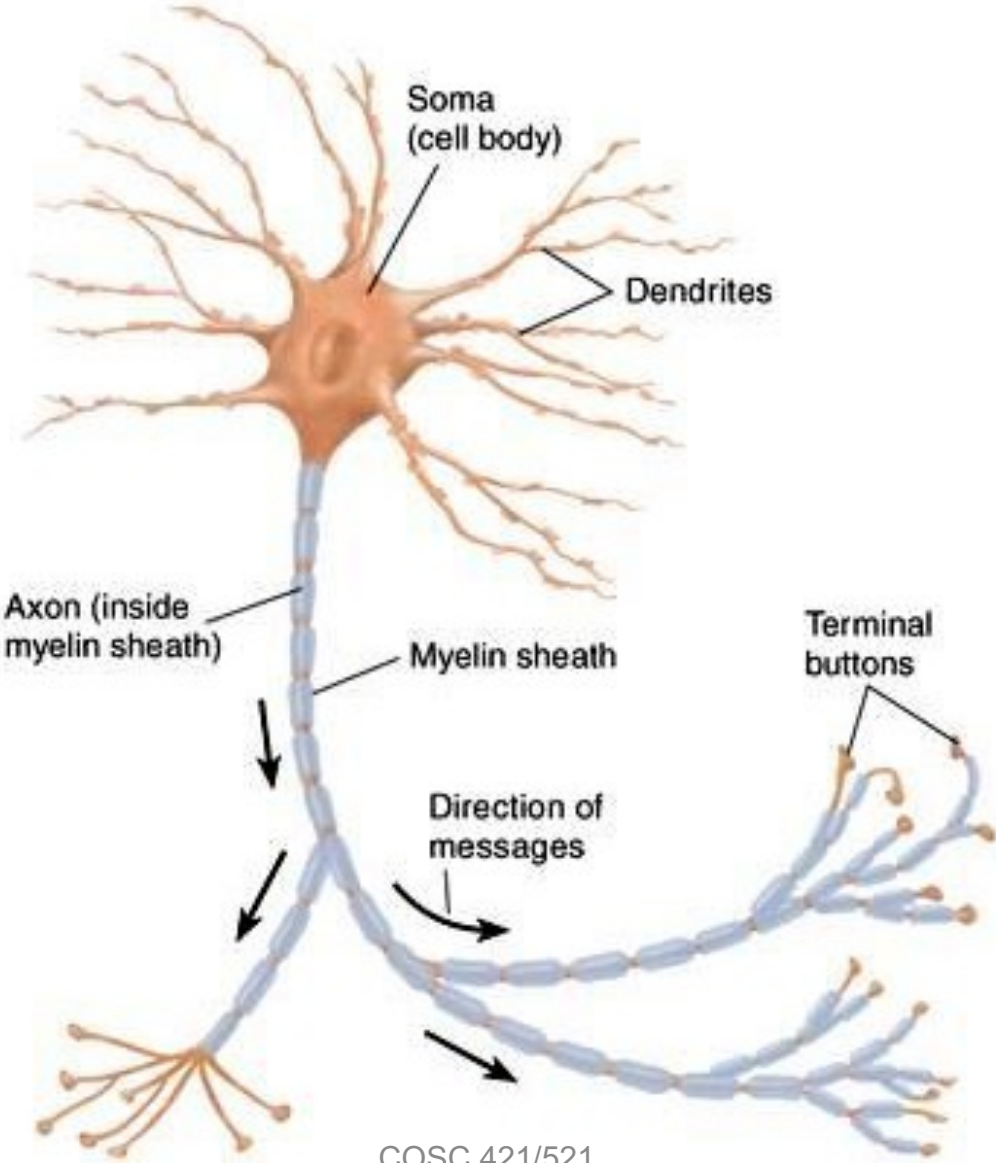


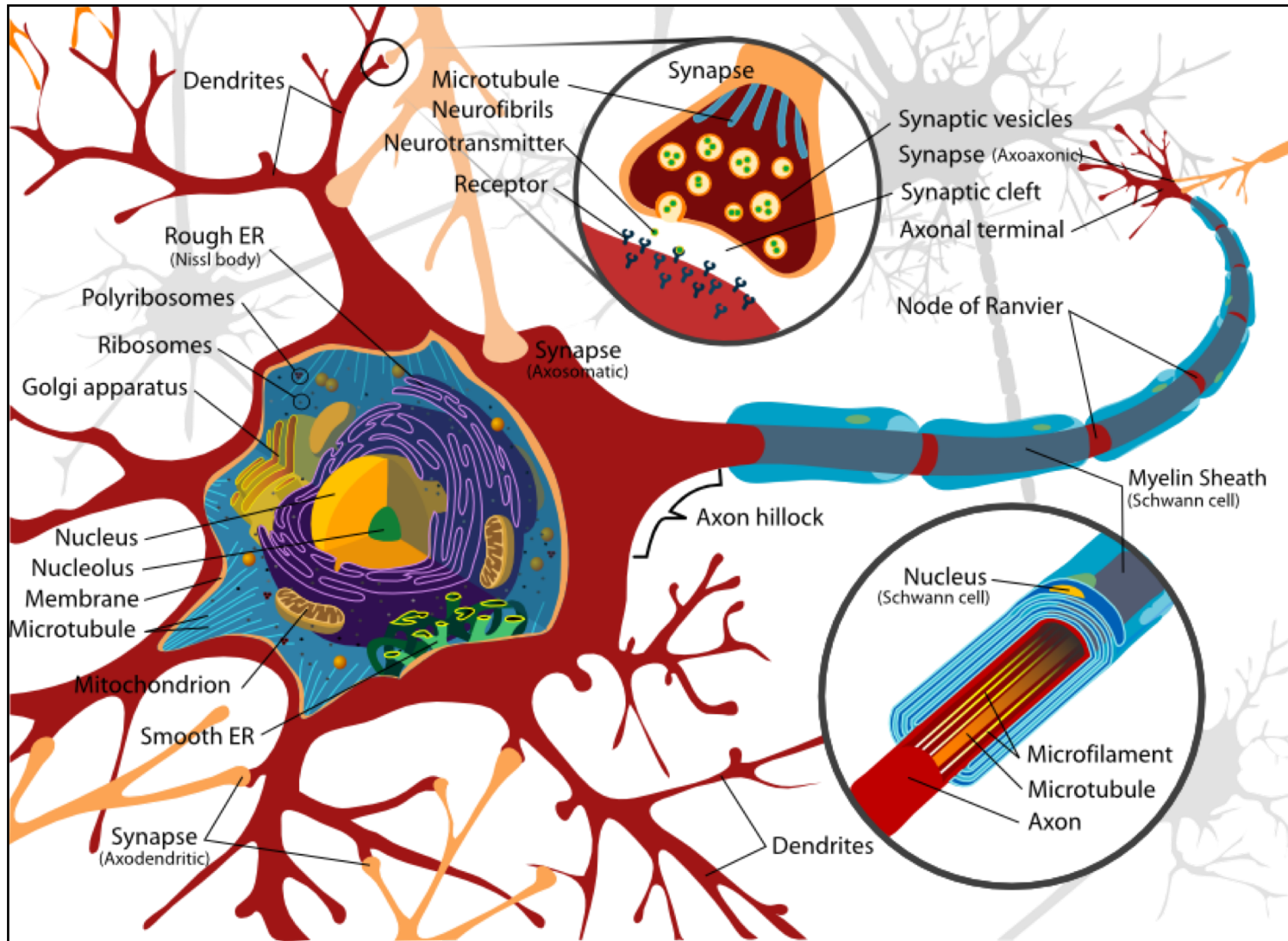
## 2. Neurons

# Typical Neuron

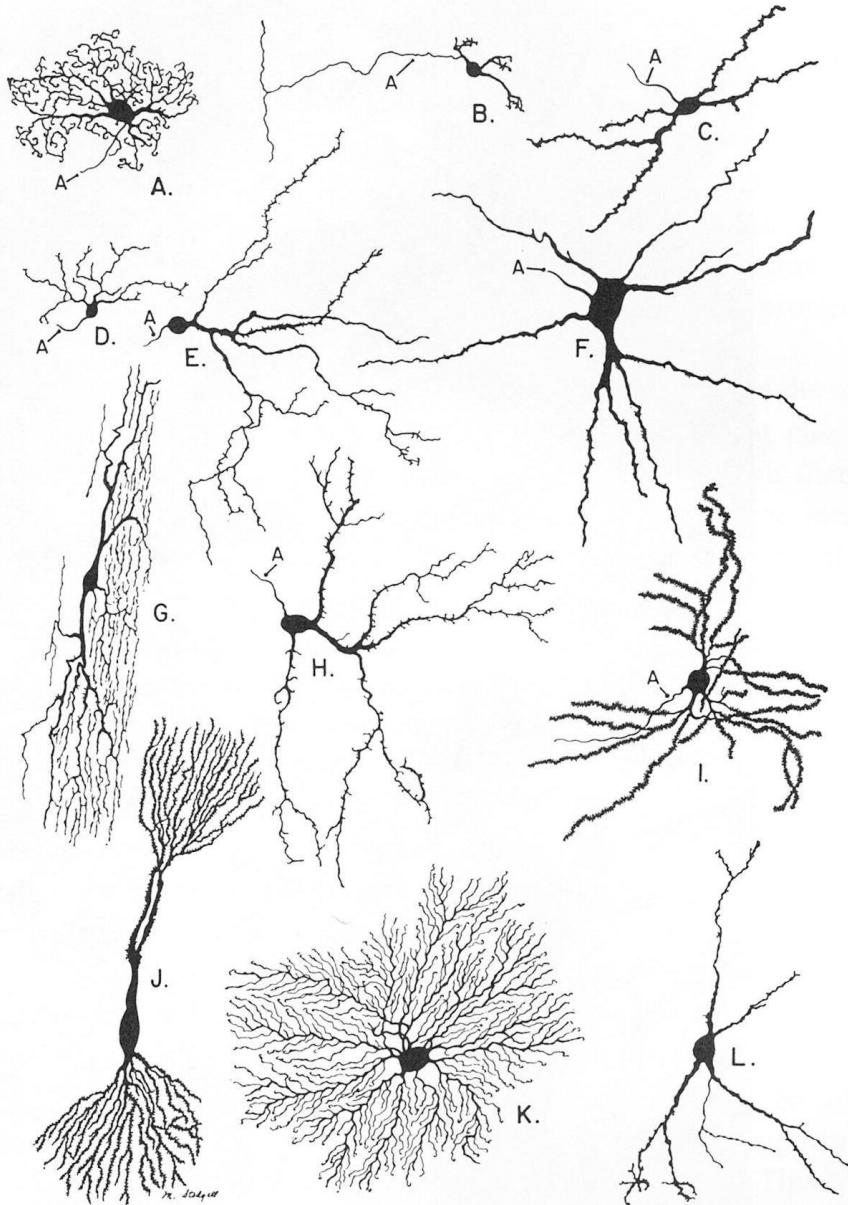




# Typical Neuron



# Dendritic Trees of Some Neurons

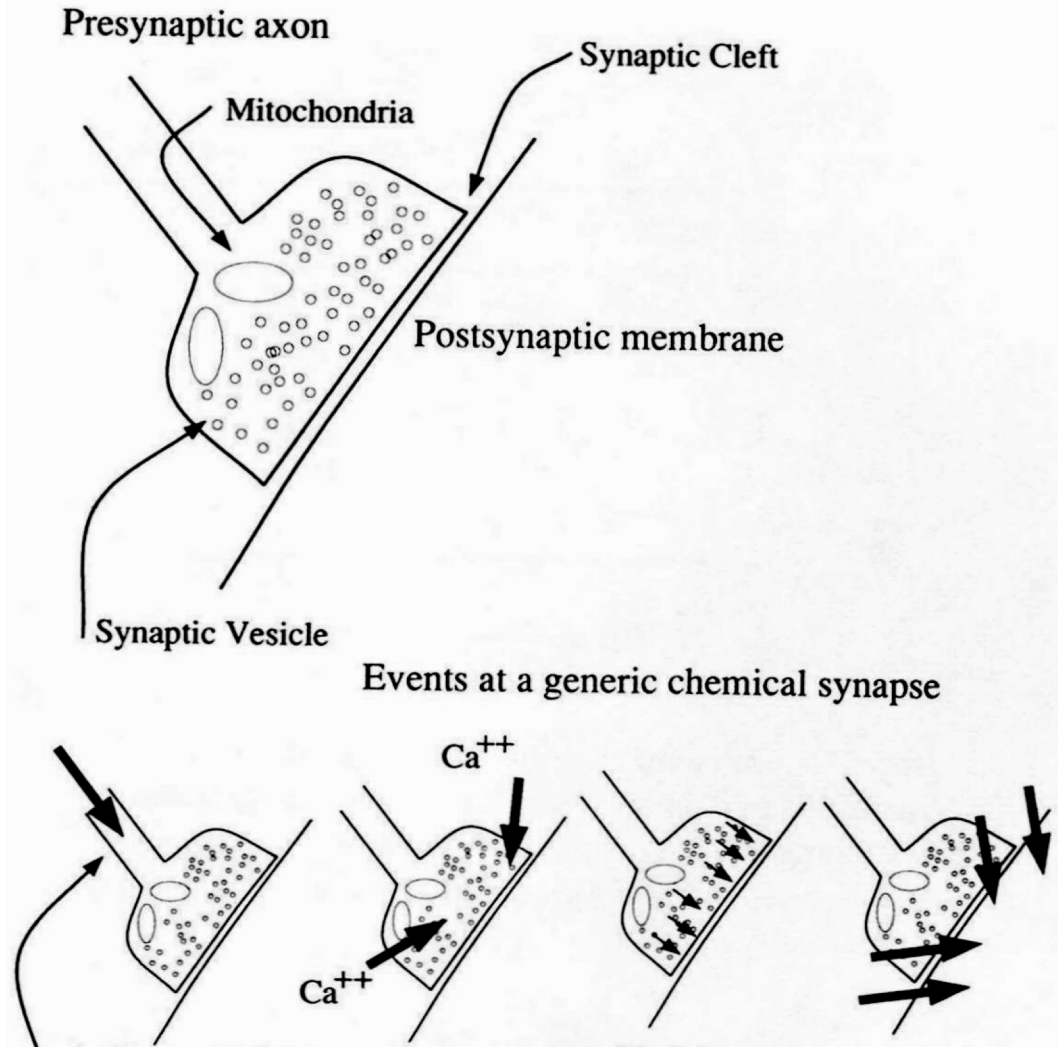


- A. inferior olivary nucleus
- B. granule cell of cerebellar cortex
- C. small cell of reticular formation
- D. small gelatinosa cell of spinal trigeminal nucleus
- E. ovoid cell, nucleus of tractus solitarius
- F. large cell of reticular formation
- G. spindle-shaped cell, substantia gelatinosa of spinal chord
- H. large cell of spinal trigeminal nucleus
- I. putamen of lenticular nucleus
- J. double pyramidal cell, Ammon's horn of hippocampal cortex
- K. thalamic nucleus
- L. globus pallidus of lenticular nucleus

# Synapses

Animation of synapses:  
[Hurd Studios Nicotine.flv](#)

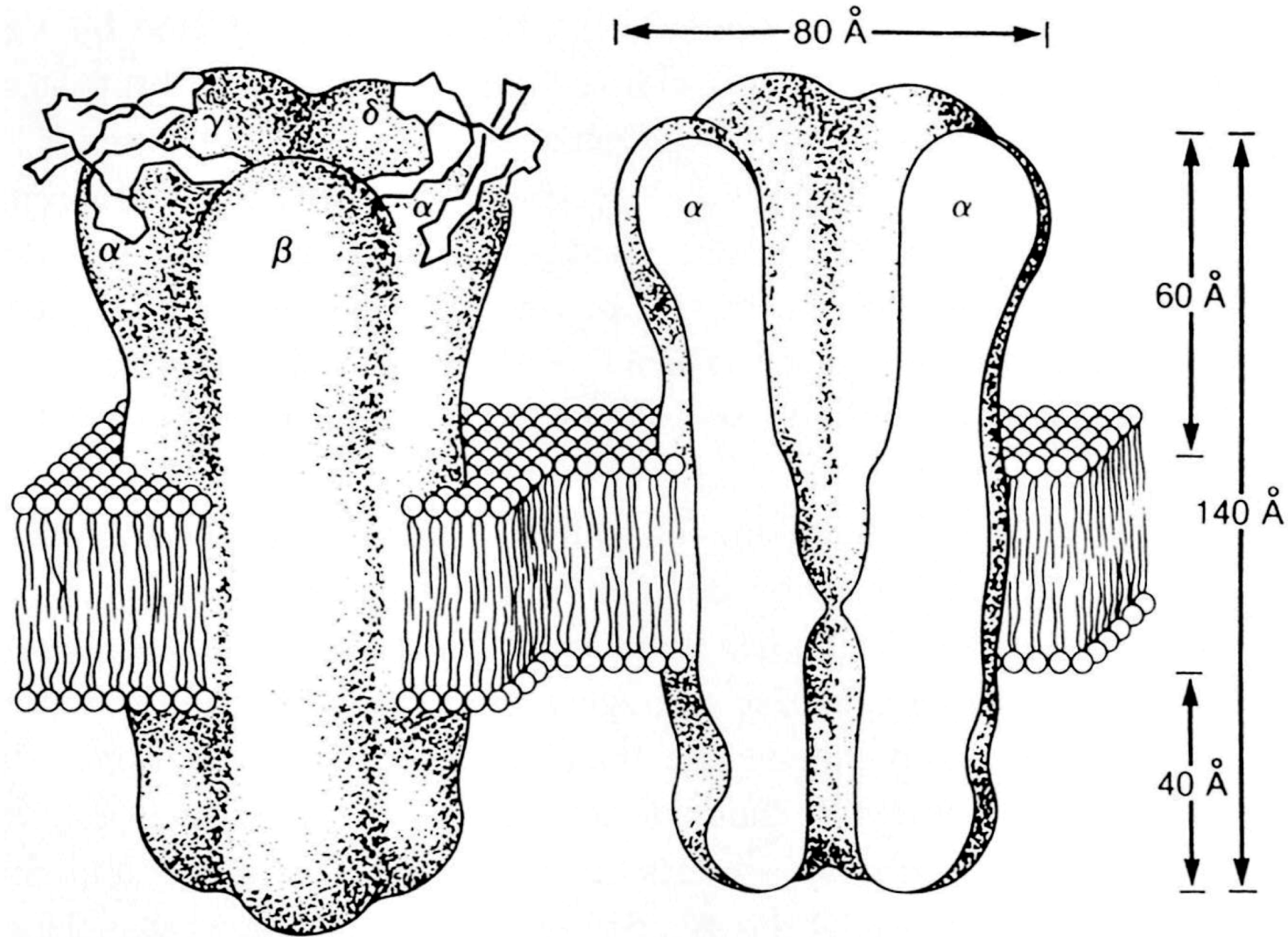
# Chemical Synapse



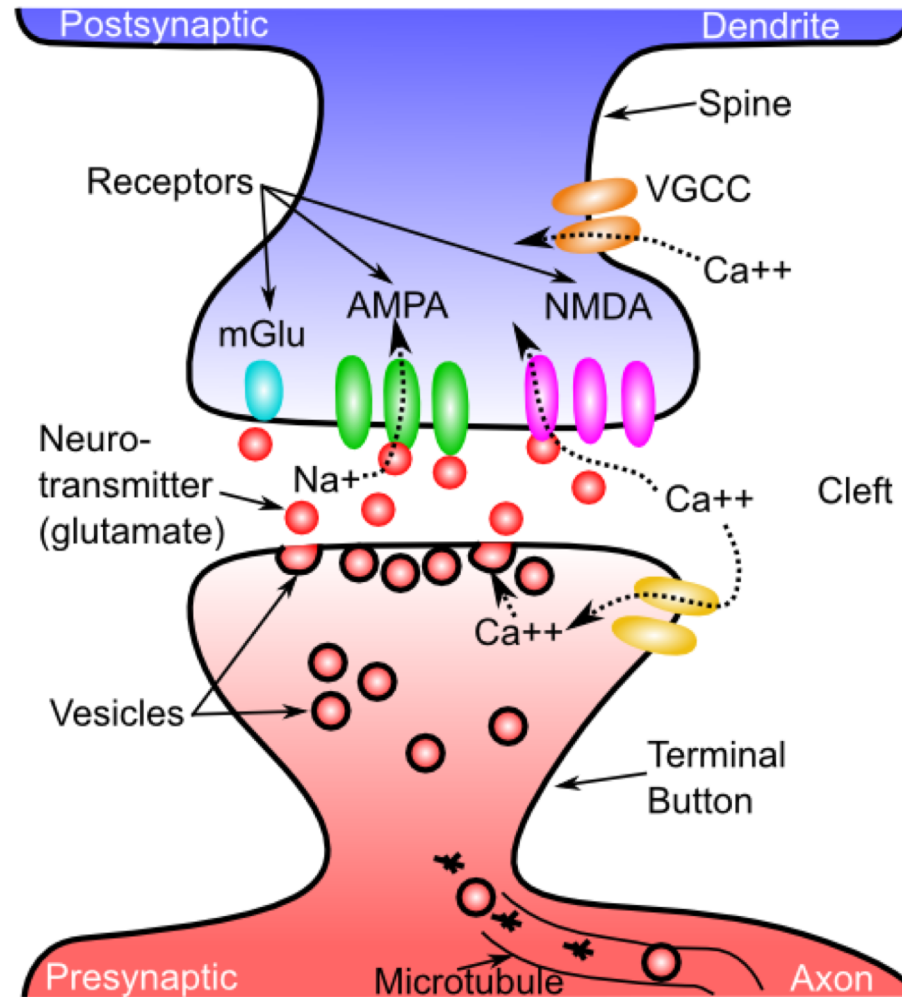
1. Action potential arrives at synapse
2. Opens Ca ion channels and Ca<sup>++</sup> ions enter cell
3. Vesicles move to membrane, release neurotransmitter
4. Transmitter crosses cleft, causes postsynaptic voltage change



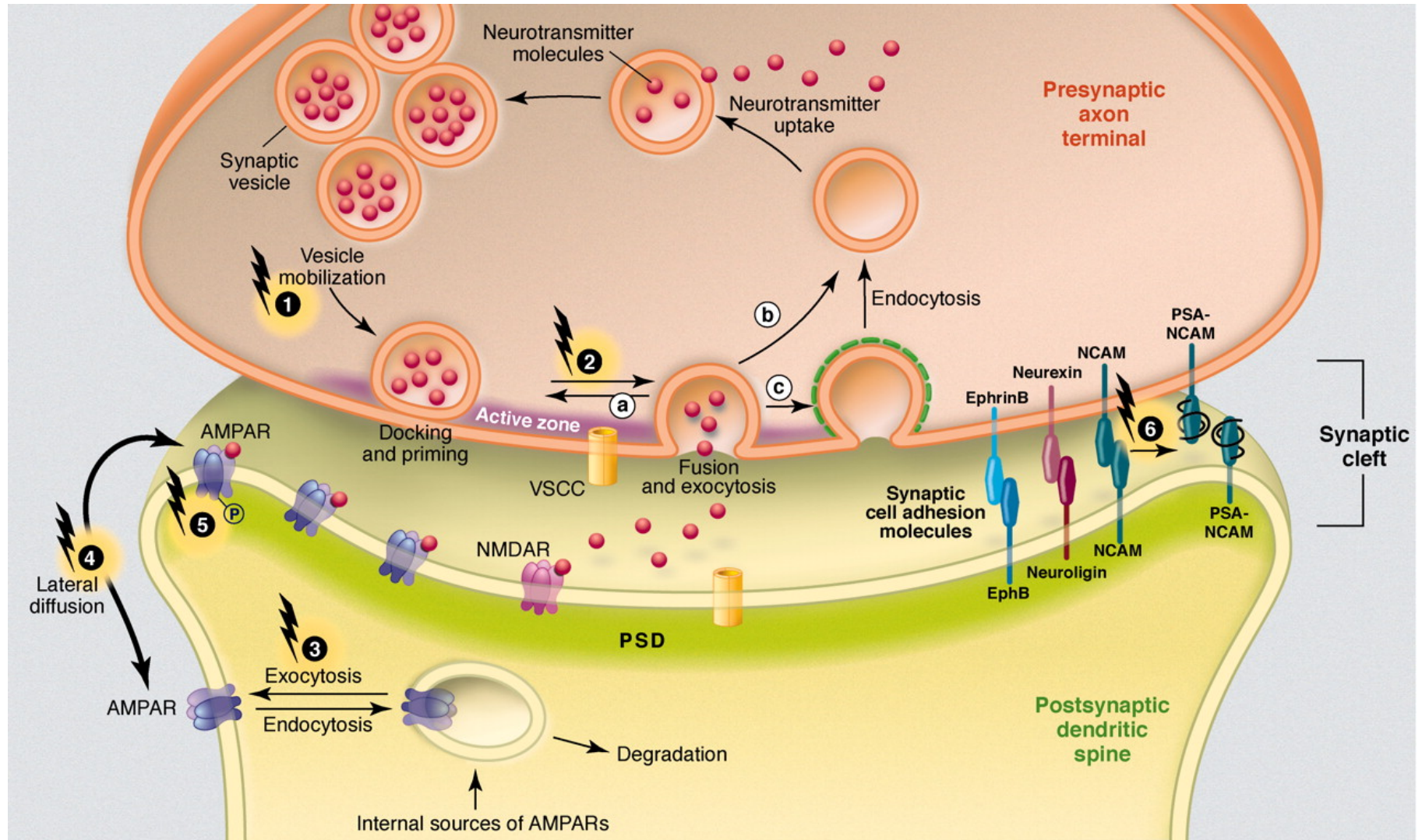
# Typical Receptor



# Synapse with Receptors



**Fig. 3 Activity-dependent modulation of pre-, post-, and trans-synaptic components.**



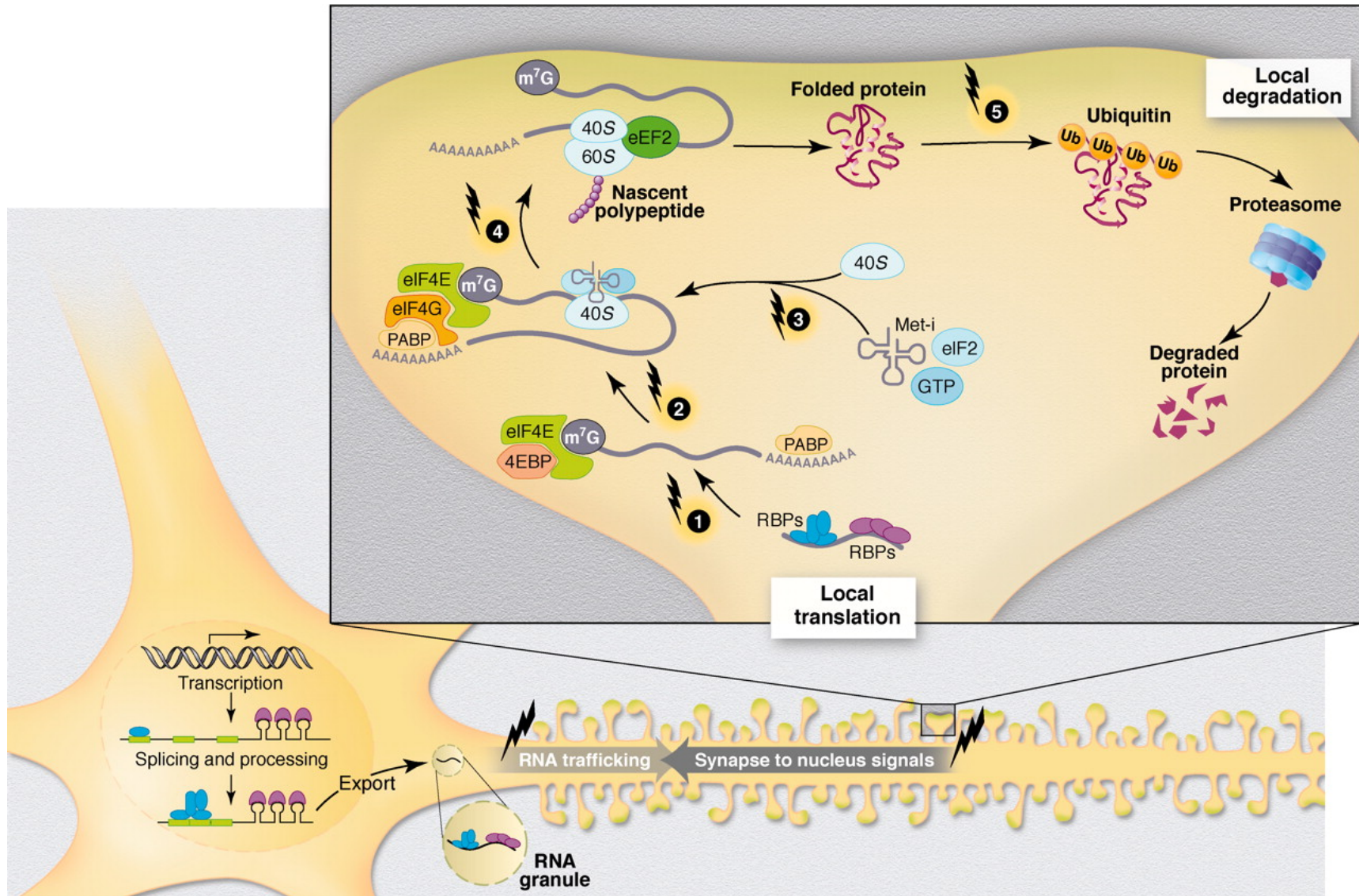
V M Ho et al. Science 2011;334:623-628

1/15/20

9



**Fig. 4 Local regulation of the synaptic proteome.**

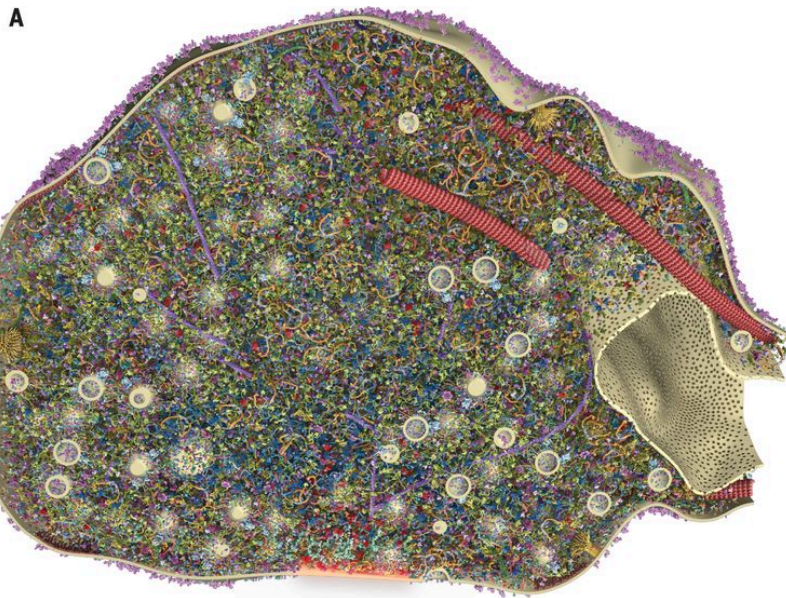


V M Ho et al. Science 2011;334:623-628

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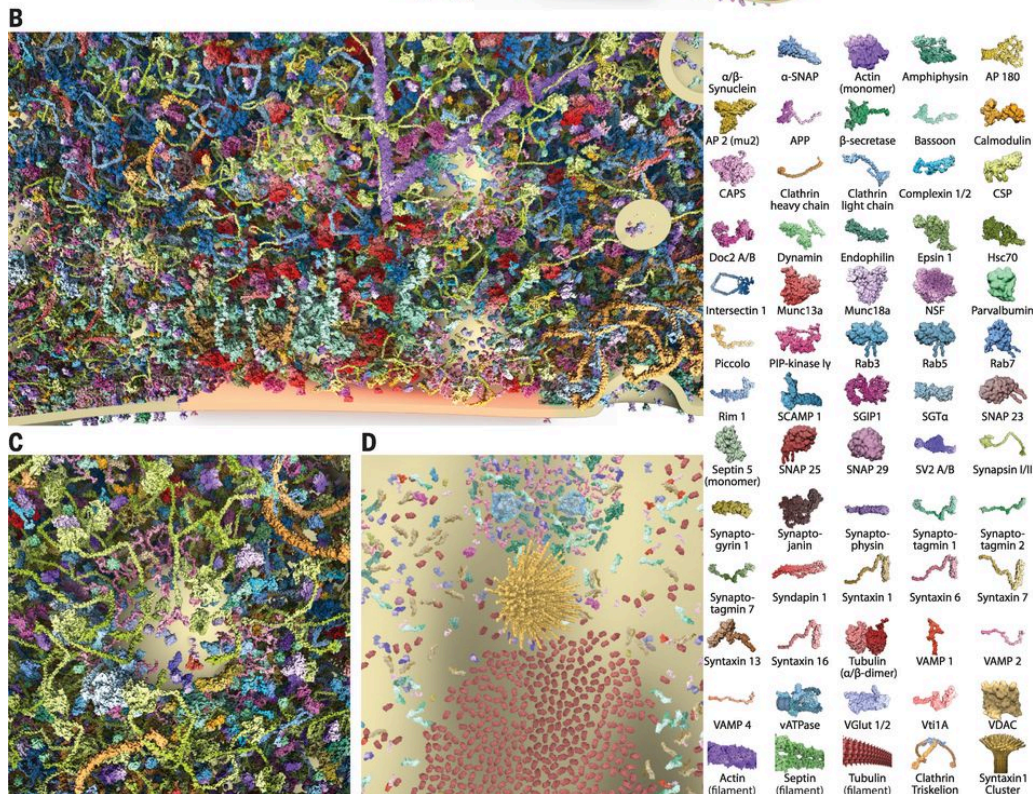
10





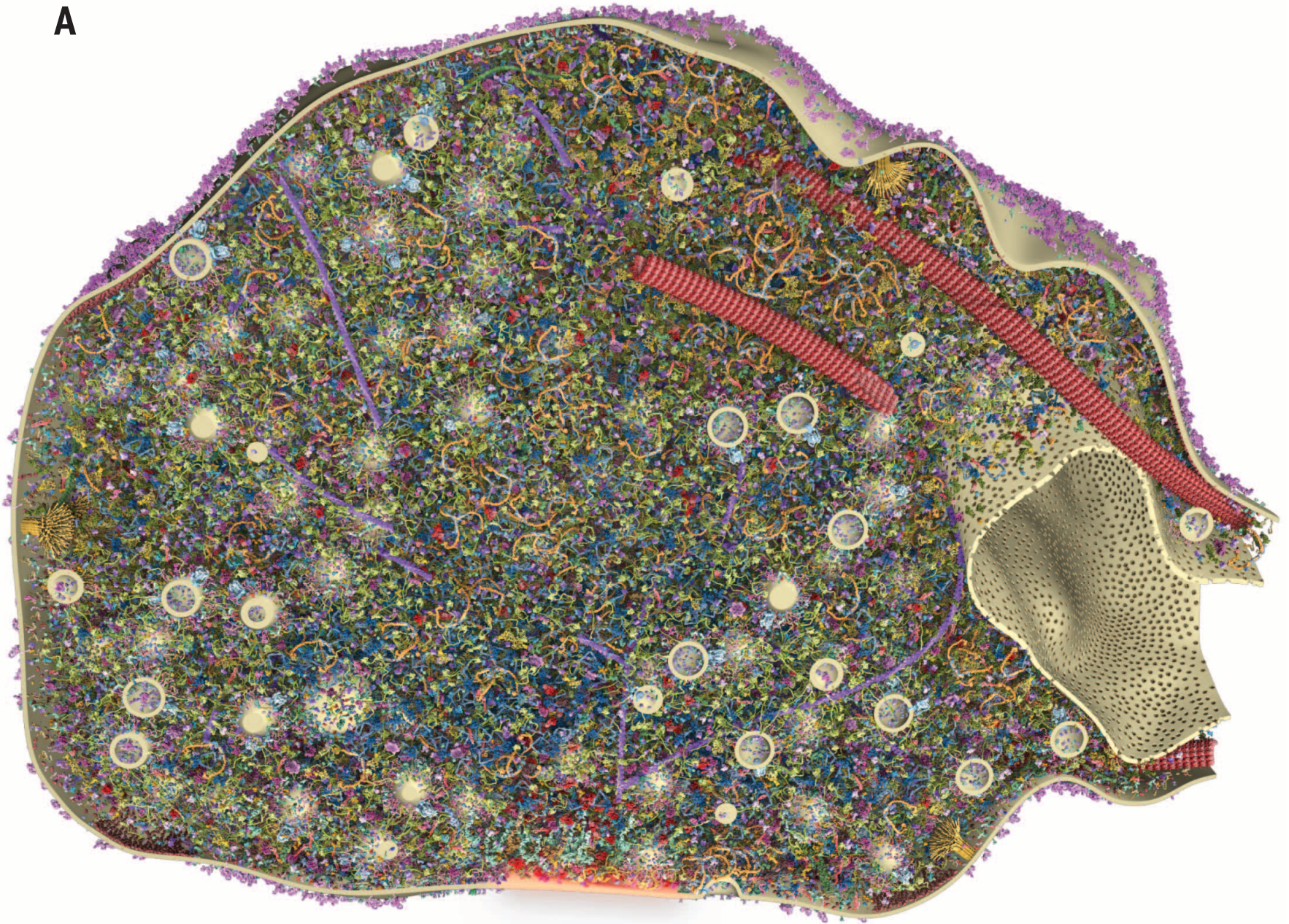
**Fig. 3: A 3D model of synaptic architecture.**

- A section through the synaptic bouton, indicating 60 proteins.
- High-zoom view of the active zone area.
- High-zoom view of one vesicle within the vesicle cluster.
- High-zoom view of a section of the plasma membrane in the vicinity of the active zone. Clusters of syntaxin (yellow) and SNAP 25 (red) are visible, as well as a recently fused synaptic vesicle (top). The graphical legend indicates the different proteins (right). Displayed synaptic vesicles have a diameter of 42 nm.





A



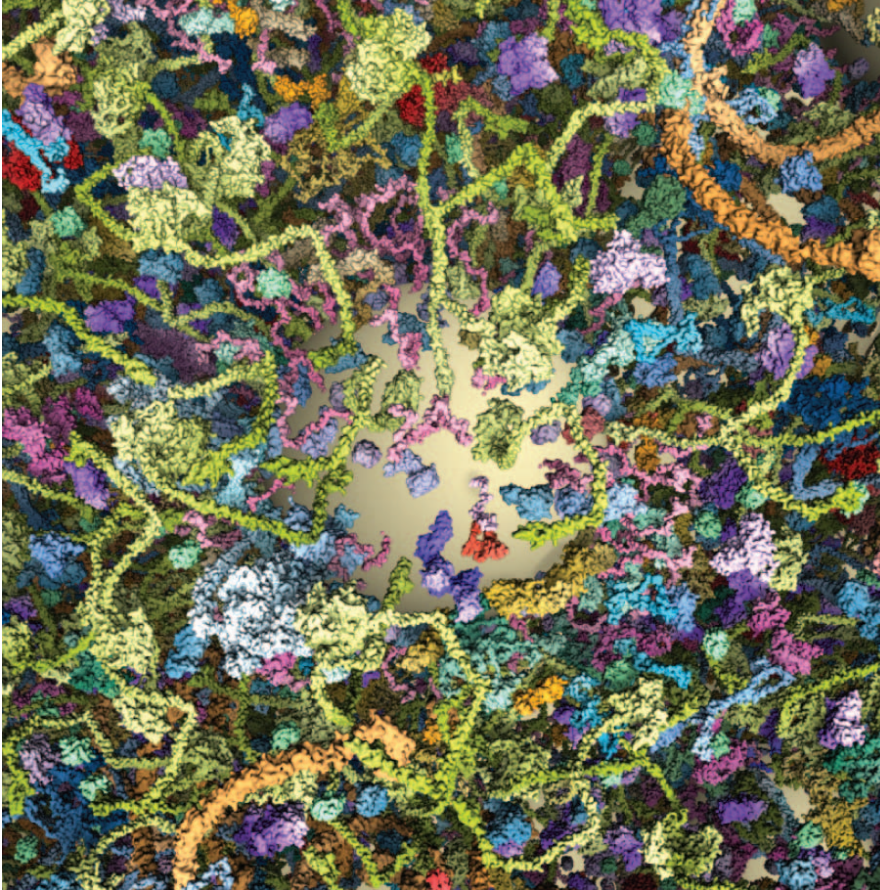


**B**

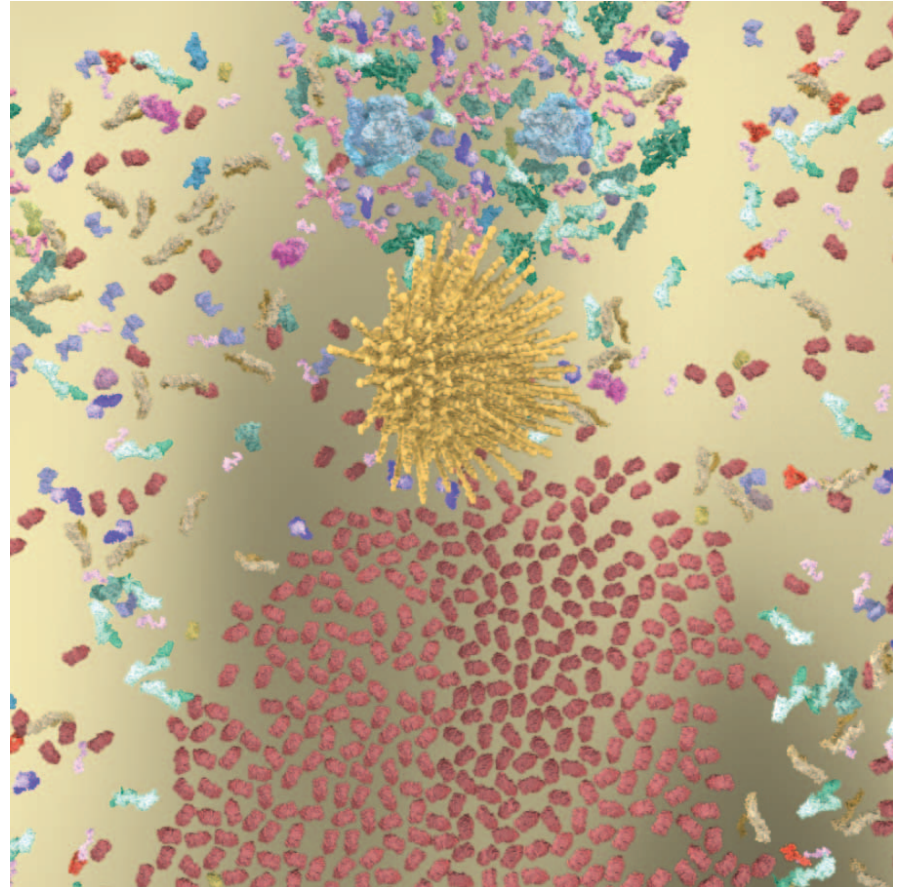




**C**

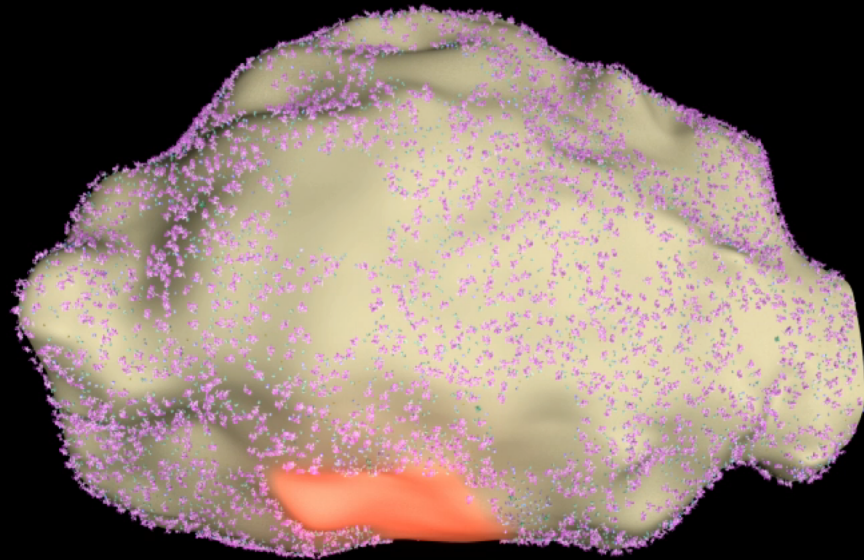


**D**

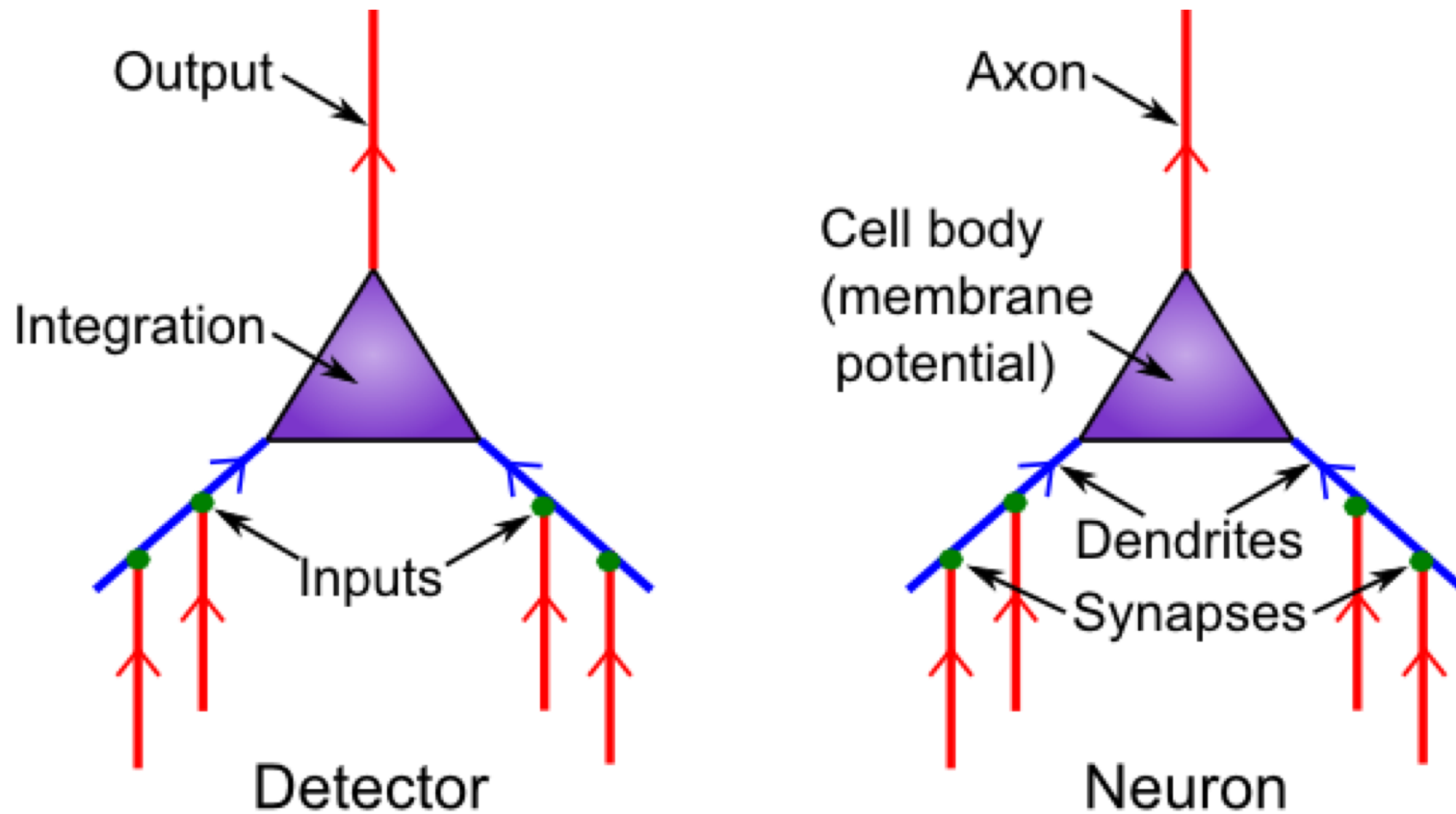


# Video of 3D Model

Lateral view of the synaptic bouton. Pink colored protein is APP.  
The active zone area is shown by the fire-red shading.



# Detector Model

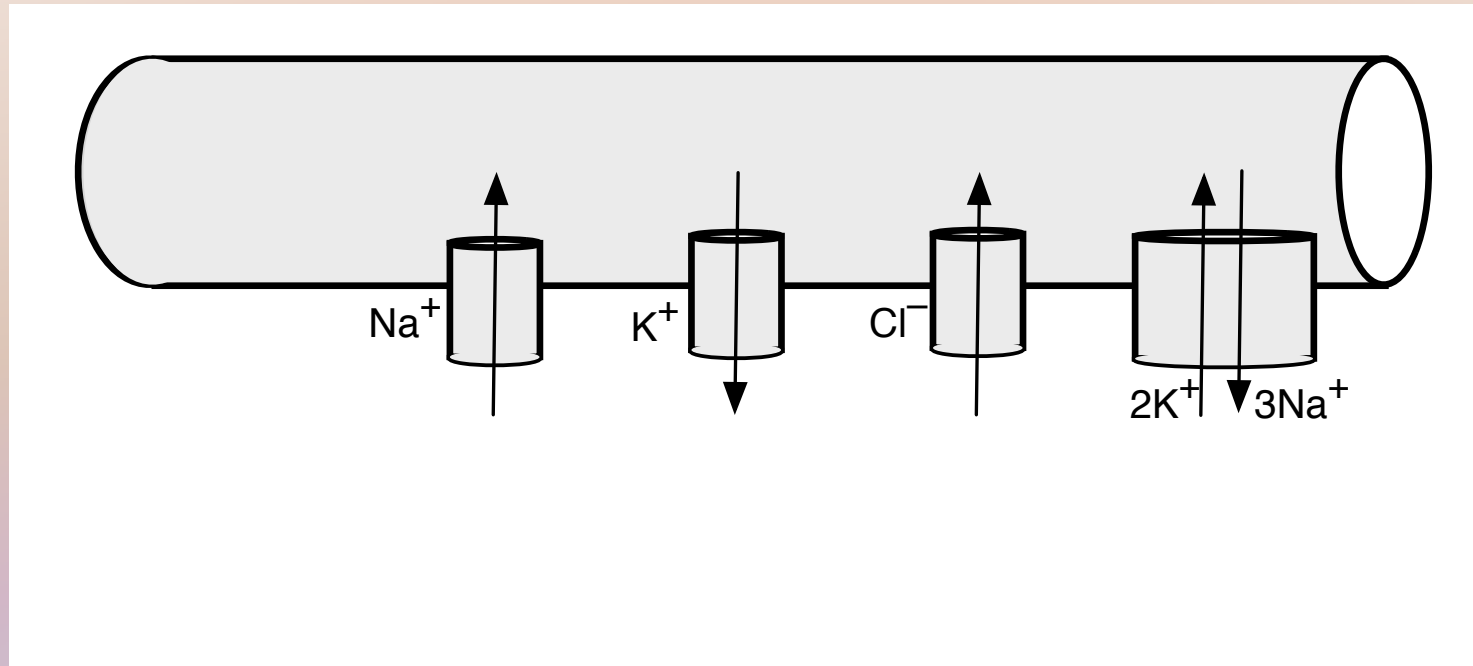




# Overall Strategy

- Neurons are electrical systems and can be described using basic electrical equations.
- Use these equations to simulate on a computer.
- Need a fair bit of math to get a full working model (more here than most chapters), but you only really need to understand conceptually.

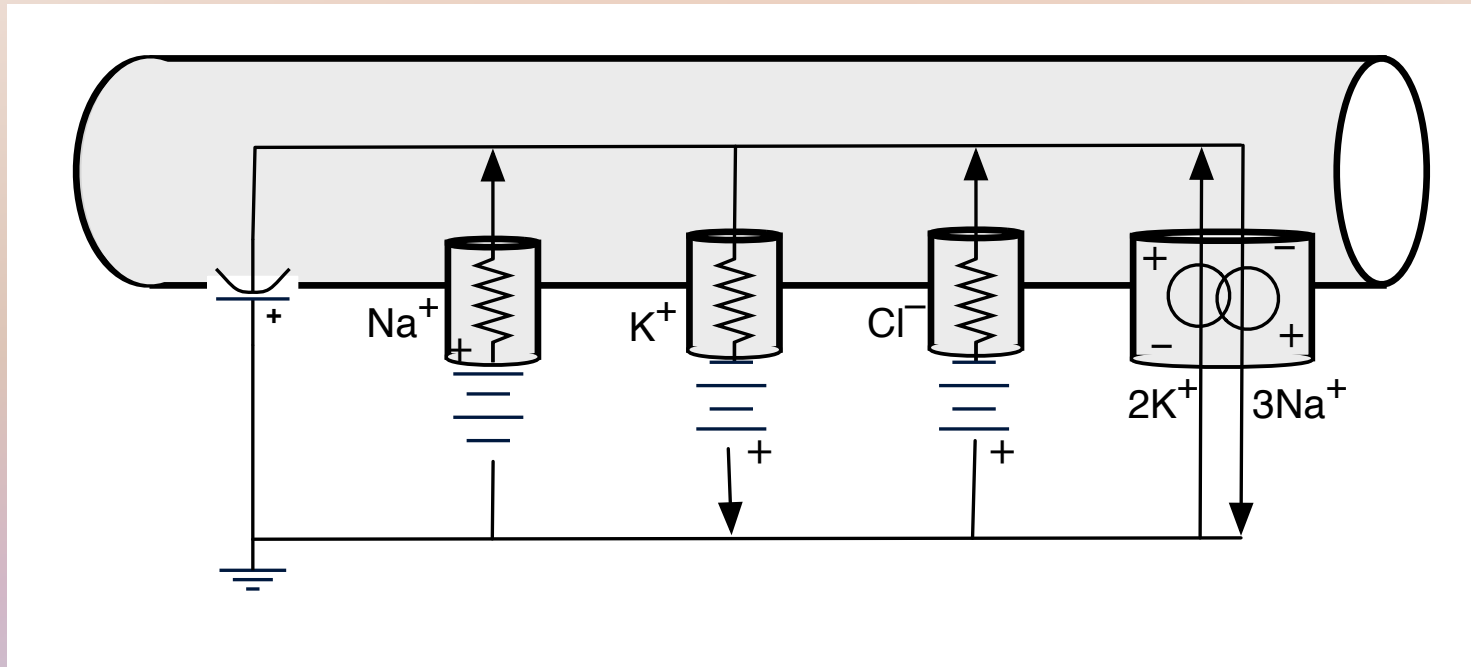
# Membrane Potential: Channels



- Na-K pump  $\Rightarrow$  intracellular negative
- $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Cl}^-$  diffusing through their channels
- create potentials across channels

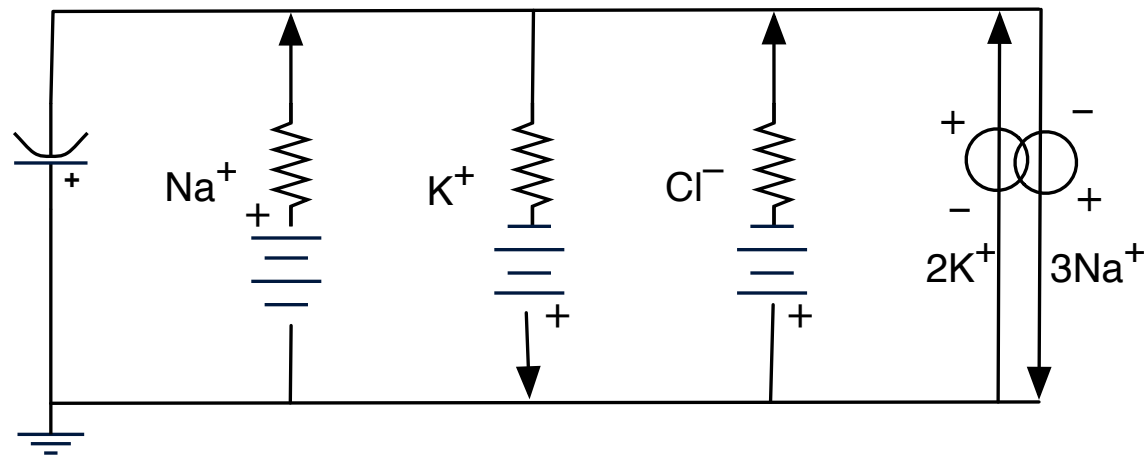


# Membrane Potential: Channels & Equivalent Circuit



- Open channels define resistance to ion flow
- Membrane acts like insulator
- Ion pump charges membrane capacitance

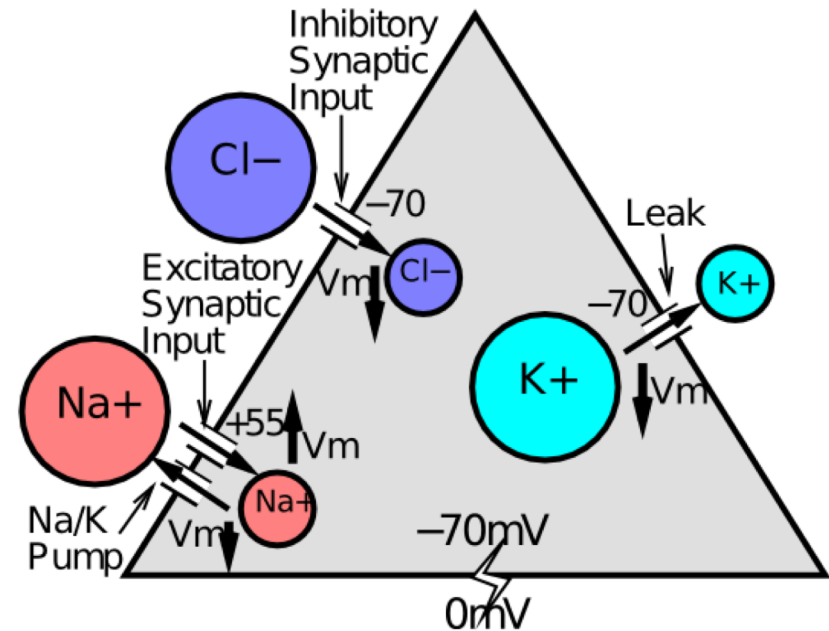
# Membrane Potential: Equivalent Circuit



- Ion pump is constant
- Change in conductance of channels
- $\Rightarrow$  change in membrane potential

# Neurophysiology of Membrane

- Na-K pump pumps  $\text{Na}^+$  out of the neuron and pumps a lesser amount of  $\text{K}^+$  into the neuron
- Creates negative resting potential ( $-70 \text{ mV}$ )
- $\text{Na}^+$  wants in (can't, due to closed channels)
- $\text{Cl}^-$  is in balance (diffusion pushes in, electrical pushes out)
- $\text{K}^+$  is in balance (diffusion pushes out, electrical pushes in)



# Ions Summary

- Excitatory synaptic input boosts the membrane potential by allowing  $\text{Na}^+$  ions to enter the neuron (depolarization)
- Inhibitory synaptic input serves to counteract this increase in membrane potential by allowing  $\text{Cl}^-$  ions to enter the neuron
- The leak current ( $\text{K}^+$  flowing out of the neuron through open channels) acts as a drag on the membrane potential. Functionally speaking, it makes it harder for excitatory input to increase the membrane potential.

(slide based on Frank)

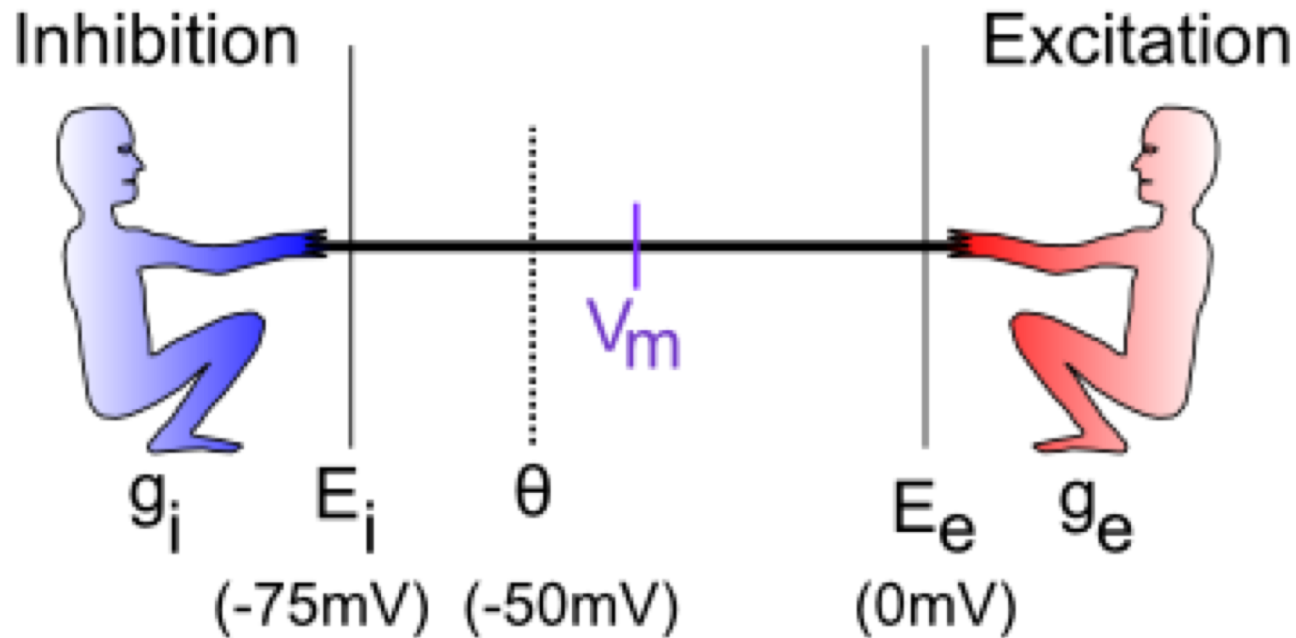
# Input Signals

- Excitatory
  - about 85% of inputs
  - AMPA channels, opened by glutamate
- Inhibitory
  - about 15% of inputs
  - GABA channels, opened by GABA
  - produced by inhibitory interneurons
- Leakage
  - potassium channels
- Synaptic efficacy (weight) is net effect of:
  - presynaptic neuron to produce neurotransmitter
  - postsynaptic channels to bind it

# Membrane Potential (Variables)

- $g_e$  = excitatory conductance
- $E_e$  = excitatory potential ( $\sim 0$  mV)
- $g_i$  = inhibitory conductance
- $E_i$  = inhibitory potential ( $-70$  mV)
- $g_l$  = leakage conductance
- $E_l$  = leakage potential
- $V_m$  = membrane potential
- $\theta$  = threshold

# The Tug-of-War



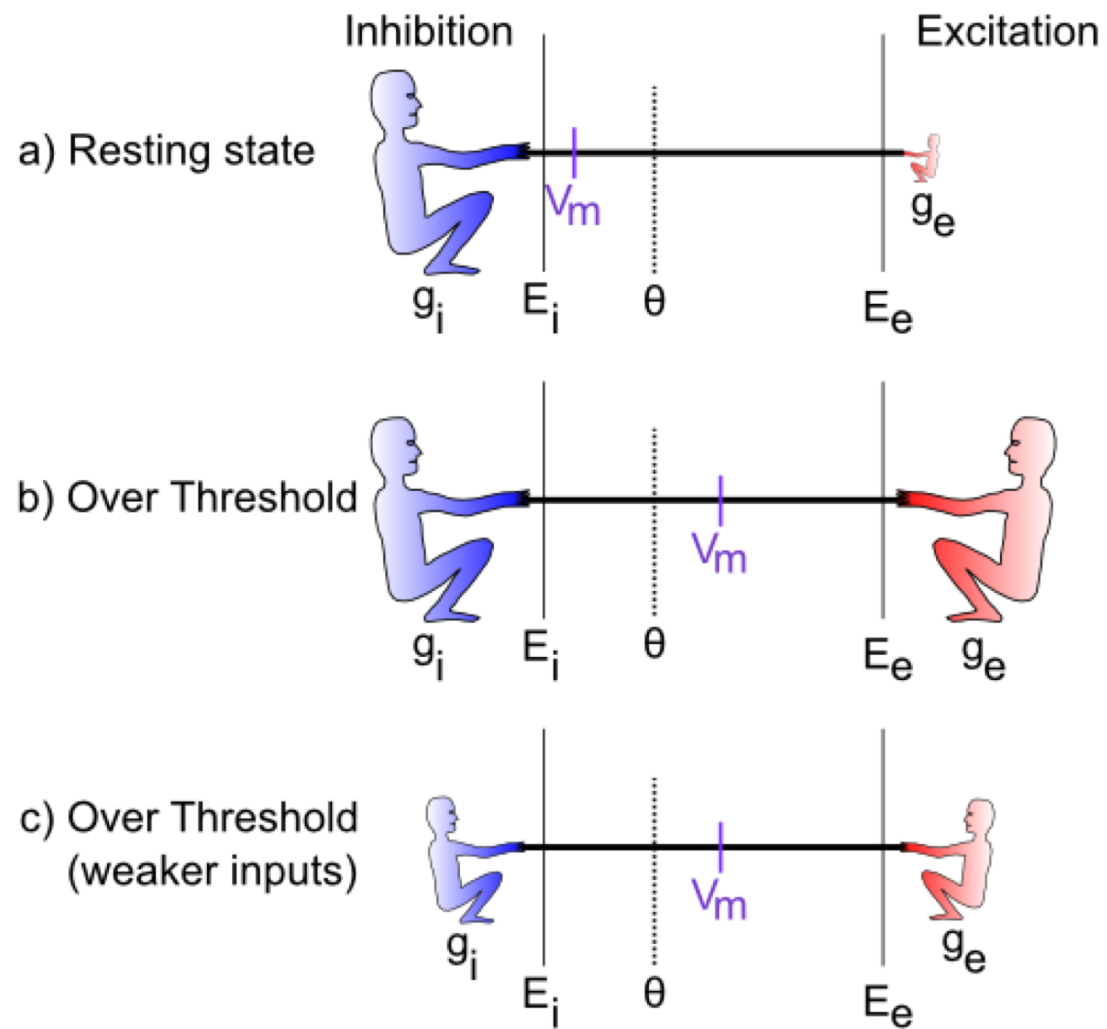
How strongly each guy pulls:  $I = g (E - V_m)$

$g$  = how many input channels are open

$E$  = driving potential (pull down for inhibition, up for excitation)

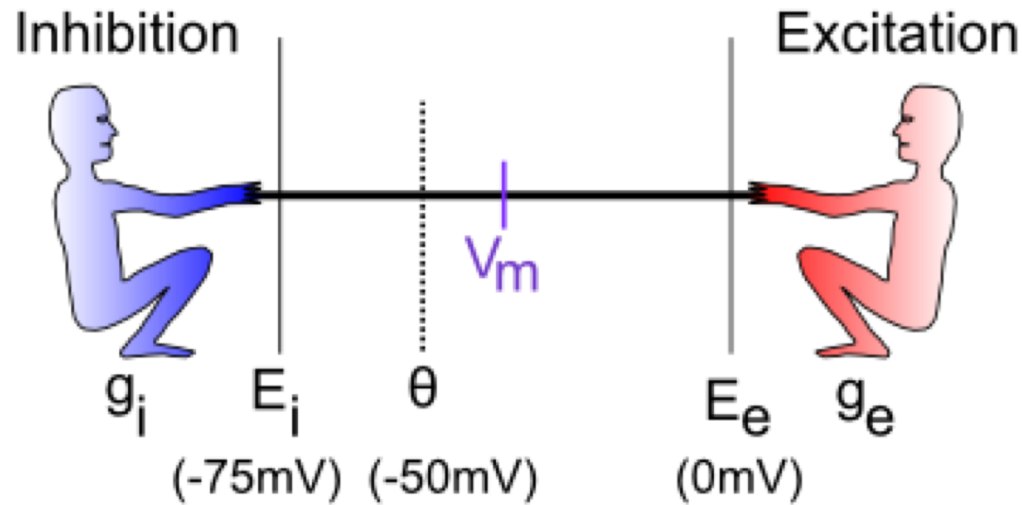
$V_m$  = the “flag” – reflects net balance between two sides

# Relative Balance





# Equations

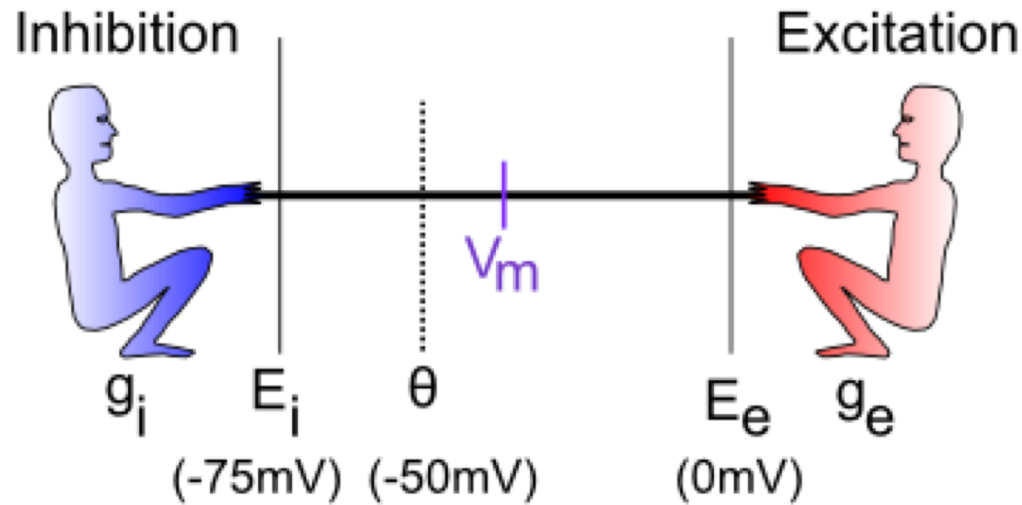


$$I_{net} = I_e + I_i + I_l = g_e (E_e - V_m) + g_i (E_i - V_m) + g_l (E_l - V_m)$$

$$V_m(t) = V_m(t-1) + dt_{vm} I_{net}$$

$$V_m(t) = V_m(t-1) + dt_{vm} [g_e (E_e - V_m) + g_i (E_i - V_m) + g_l (E_l - V_m)]$$

# Equilibrium



$$V_m = \frac{g_e}{g_e + g_i + g_l} E_e + \frac{g_i}{g_e + g_i + g_l} E_i + \frac{g_l}{g_e + g_i + g_l} E_l$$

This is just the balance of forces

# Input Conductances and Weights

- Just add them up (and take the average)

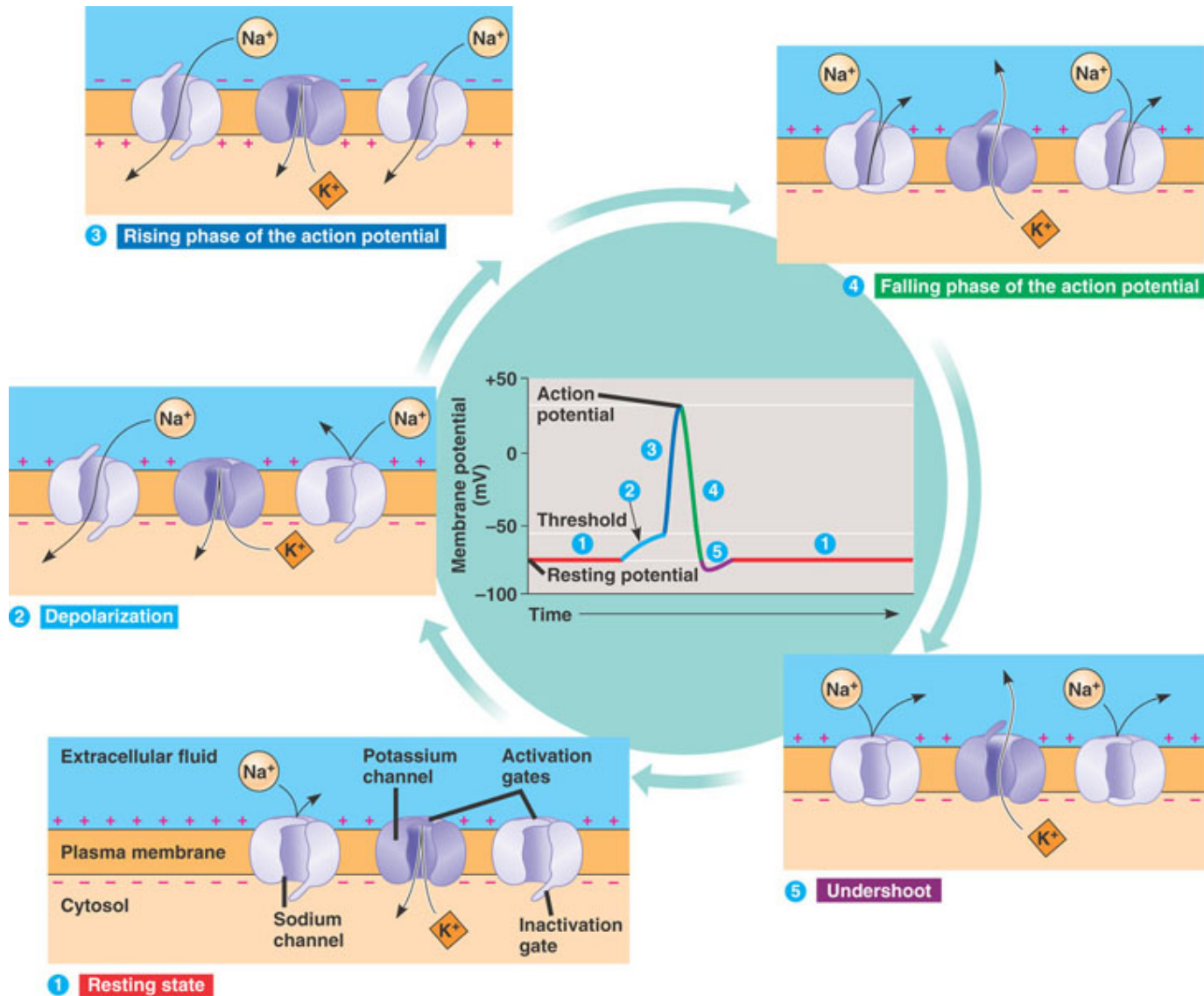
$$g_e(t) = \frac{1}{n} \sum_i x_i w_i$$

- Key concept is *weight*: how much unit listens to given input
- Weights determine what the neuron detects
- Everything you know is encoded in your weights

# Generating Output

- If  $V_m$  gets over threshold, neuron fires a spike
- Spike resets membrane potential back to rest
- Has to climb back up to threshold to spike again

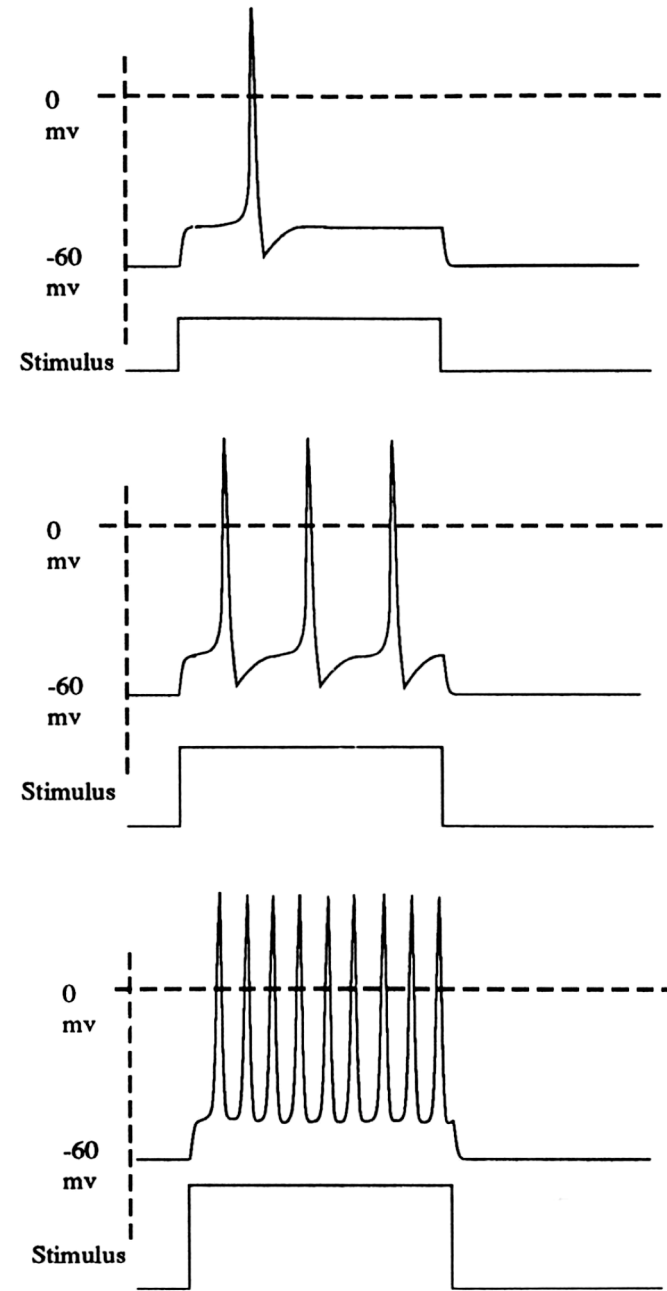
# Action Potential Generation



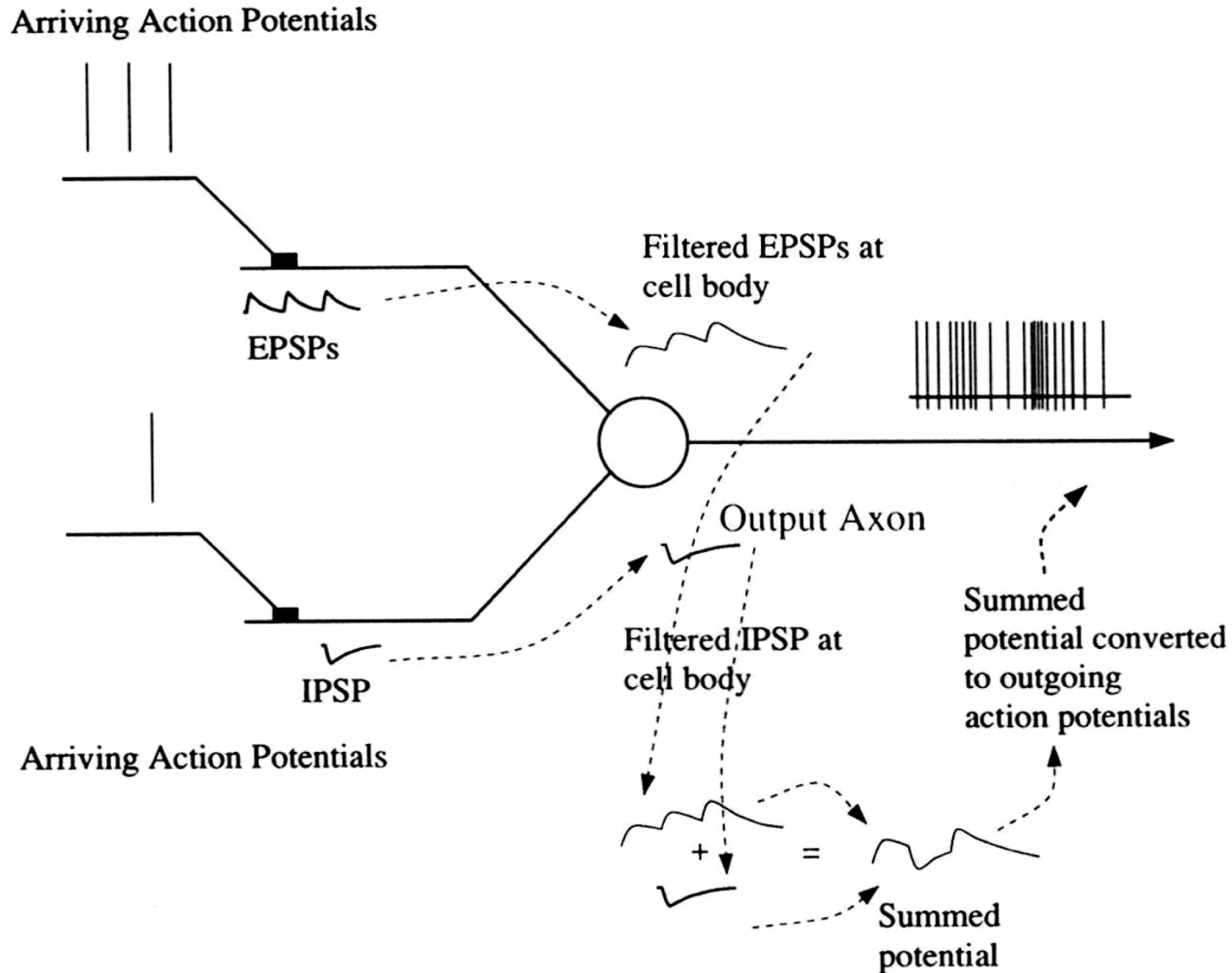
# Action Potential Generation

- From Algonquin College
- <https://www.youtube.com/watch?v=plFOiU7sTO4>

# Frequency Coding

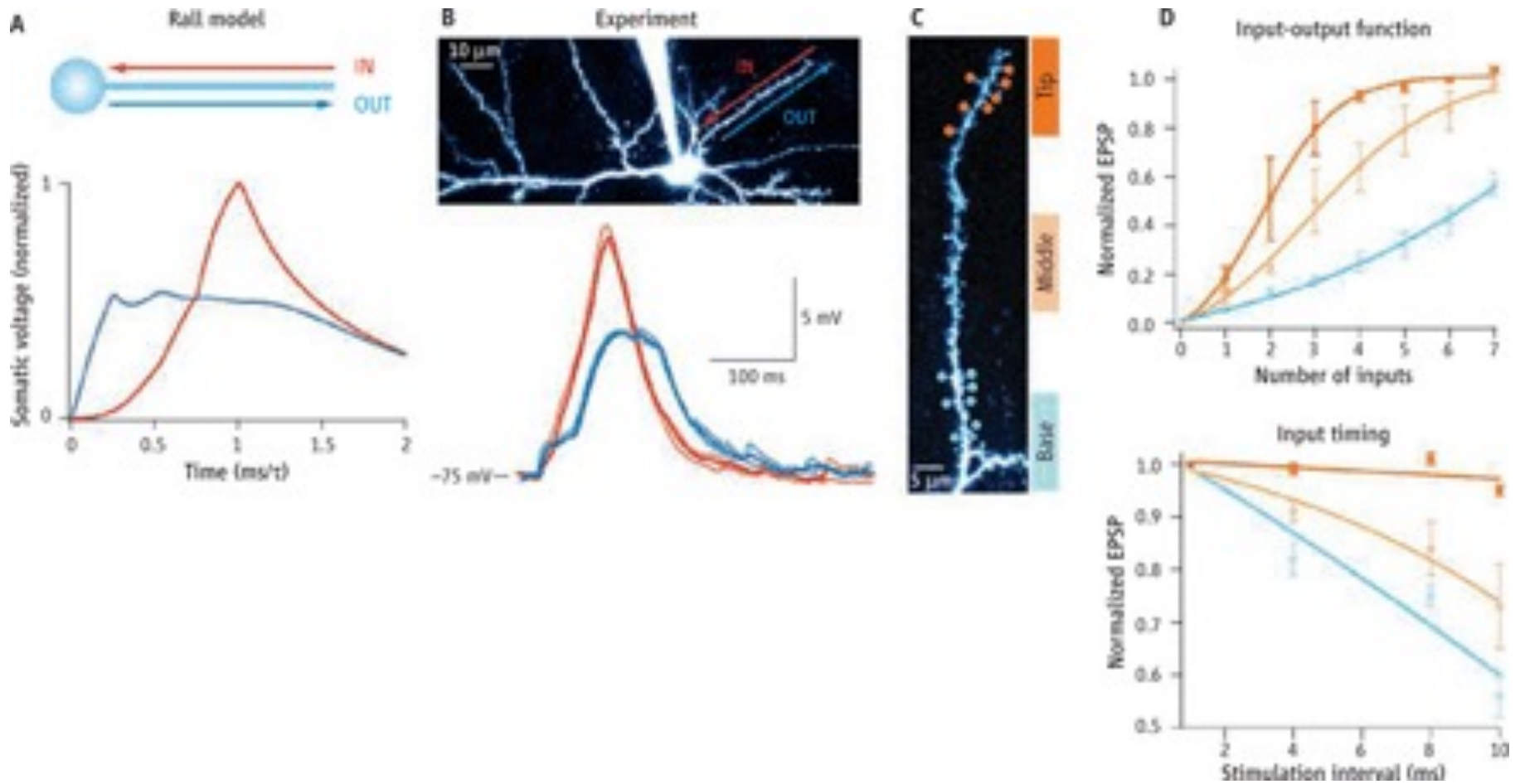


# Slow Potential Neuron



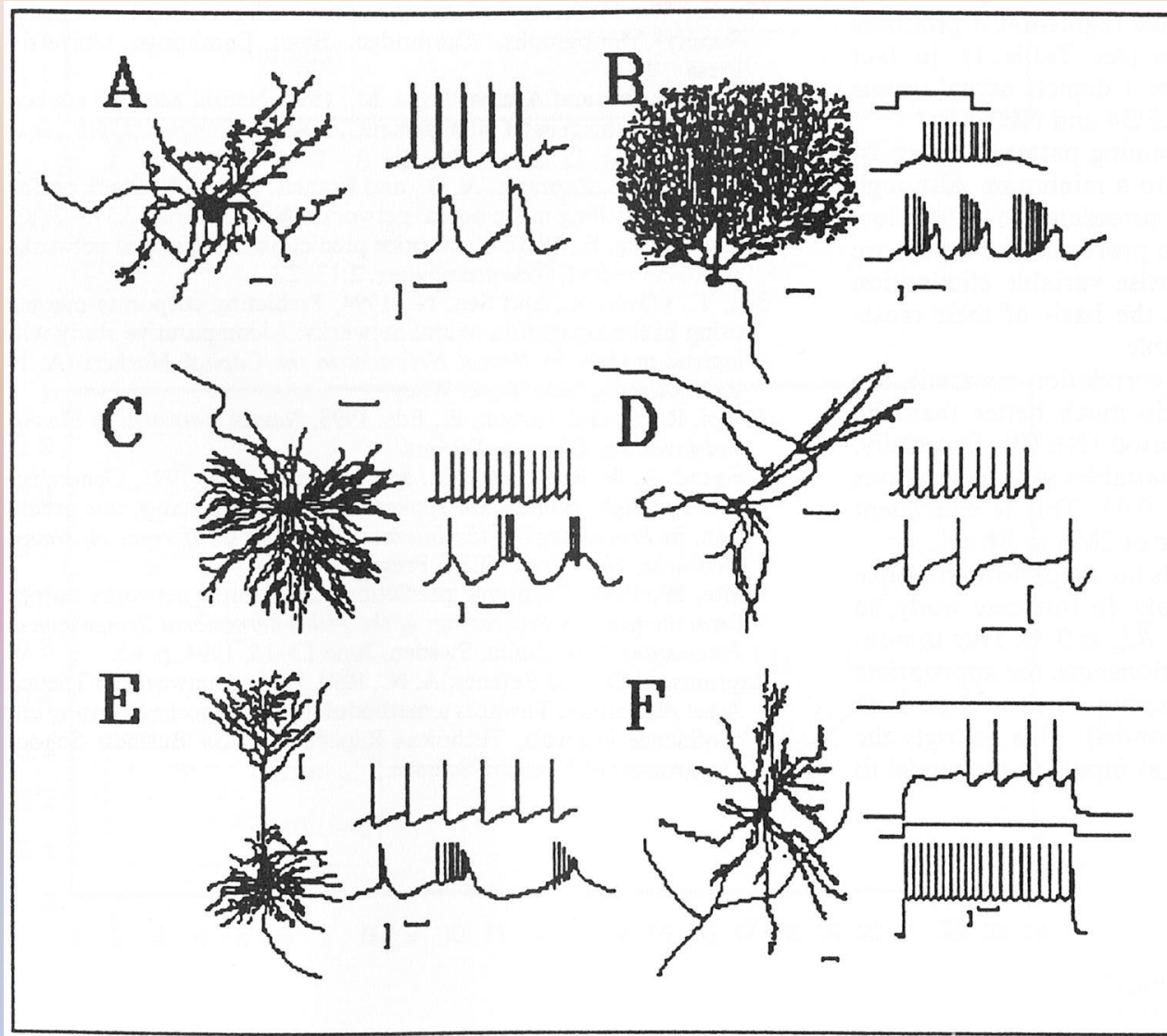


# Dendritic computation in pyramidal cells.



T Branco Science 2011;334:615-616

# Variations in Spiking Behavior



# Computational Formulation

# Membrane Potential

Currents:  $I_x = g_x (E_x - V_m)$ ,  $x = e, i, l$

Net current:  $I_{\text{net}} = I_e + I_i + I_l$

Change in membrane potential:  $\dot{V}_m = C^{-1} I_{\text{net}}$  ( $C^{-1}$  is rate constant)

$$\dot{V}_m = C^{-1} [g_e(E_e - V_m) + g_i(E_i - V_m) + g_l(E_l - V_m)]$$

Equilibrium  $V_m = \frac{g_e E_e + g_i E_i + g_l E_l}{g_e + g_i + g_l}$

# Relative vs. Absolute Conductances

- Previously,  $g_x$  was absolute conductance (measured in nanosiemens)
- More convenient to represent as product  $\bar{g}_x g_x(t)$ 
  - where  $\bar{g}_x$  is the absolute maximum conductance (all channels open)
  - and  $g_x(t)$  is the relative conductance at a given time,  $0 \leq g_x(t) \leq 1$

$$V_m = \frac{\bar{g}_e g_e(t)}{\bar{g}_e g_e(t) + \bar{g}_i g_i(t) + \bar{g}_l} E_e + \frac{\bar{g}_i g_i(t)}{\bar{g}_e g_e(t) + \bar{g}_i g_i(t) + \bar{g}_l} E_i + \frac{\bar{g}_l}{\bar{g}_e g_e(t) + \bar{g}_i g_i(t) + \bar{g}_l} E_l$$

# Discrete Spiking

```
if  $V_m > \theta$  then  
     $y := 1$ ;  
     $V_m := V_{m_r}$ ;  
else  $y := 0$ ;
```

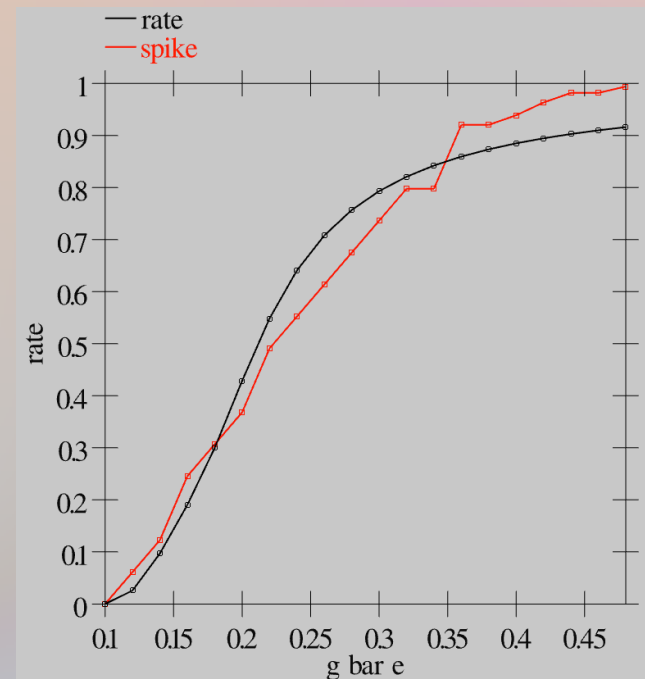
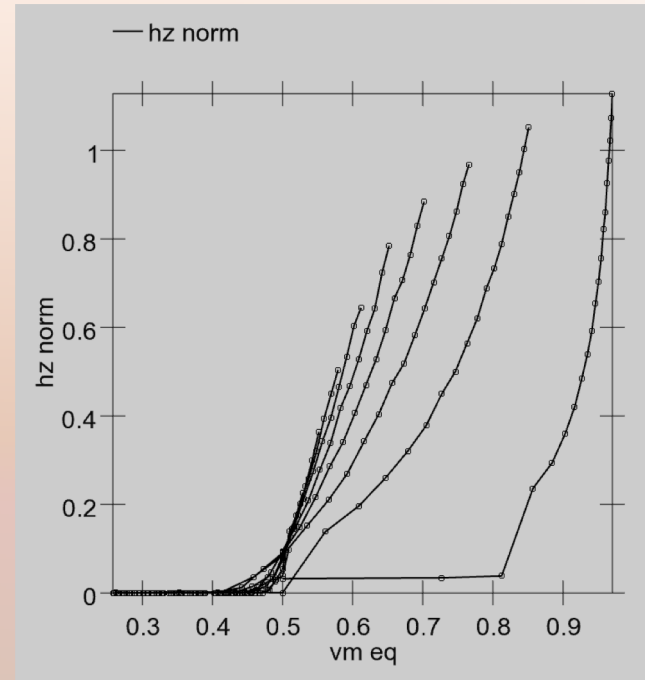


# Rate Code Approximation

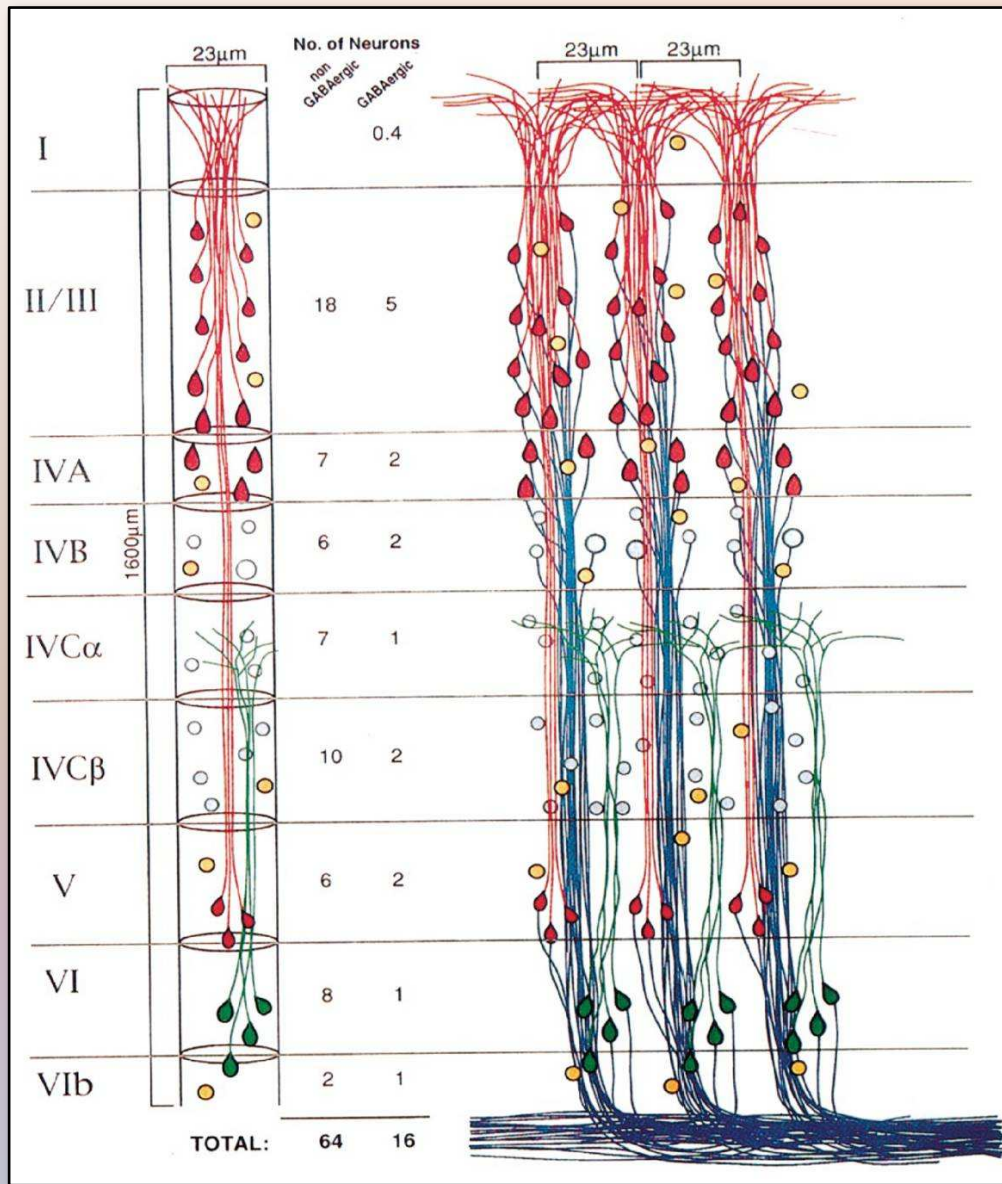
- Brain likes spikes, but rates are more convenient
  - Instantaneous and steady – smaller, faster models
  - But definitely lose several important things
  - Solution: do it both ways, and see the differences
- Goal: equation that makes good approximation of actual spiking rate for same sets of inputs

# Rate Code Approximation

- Rate-coded (simulated) neurons:
  - short-time avg spike frequency  $\approx$
  - avg behavior of minicolumn ( $\sim 100$  neurons) with similar inputs and output behavior
- Rate not predicted well by  $V_m$
- Predicted better by  $g_e$  relative to a threshold value  $g_e^\theta$



# Minicolumn



Up to  $\sim 100$  neurons

75–80% pyramidal

20–25% interneurons

20–50  $\mu\text{m}$  diameter

Length: 0.8 (mouse) to 3mm (human)

$\sim 6 \times 10^5$  synapses

75–90% synapses outside minicolumn

Interacts with  $1.2 \times 10^5$  other minicolumns

Mutually excitable

Also called *microcolumn*

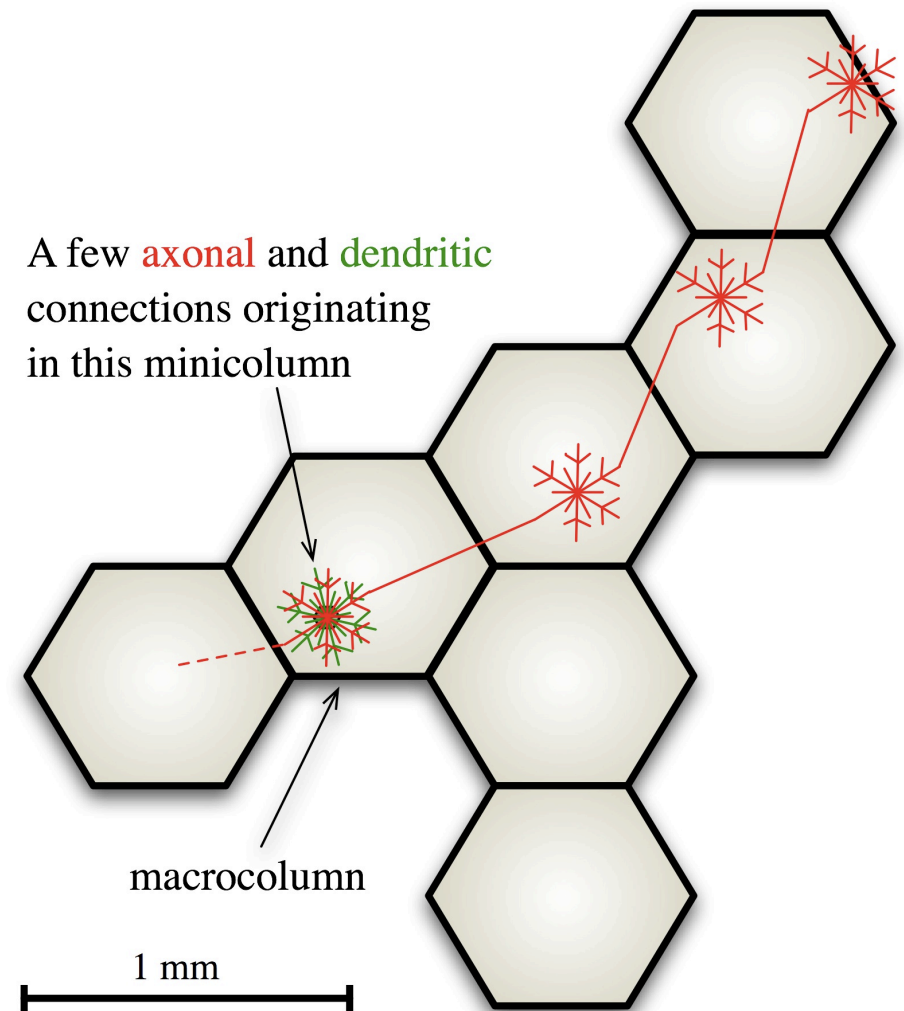
# Intracortical Connections

Dendrites extend 2–4  
minicol. diameters

Axons extend 5× (or even  
30–40× minicol.  
diameter

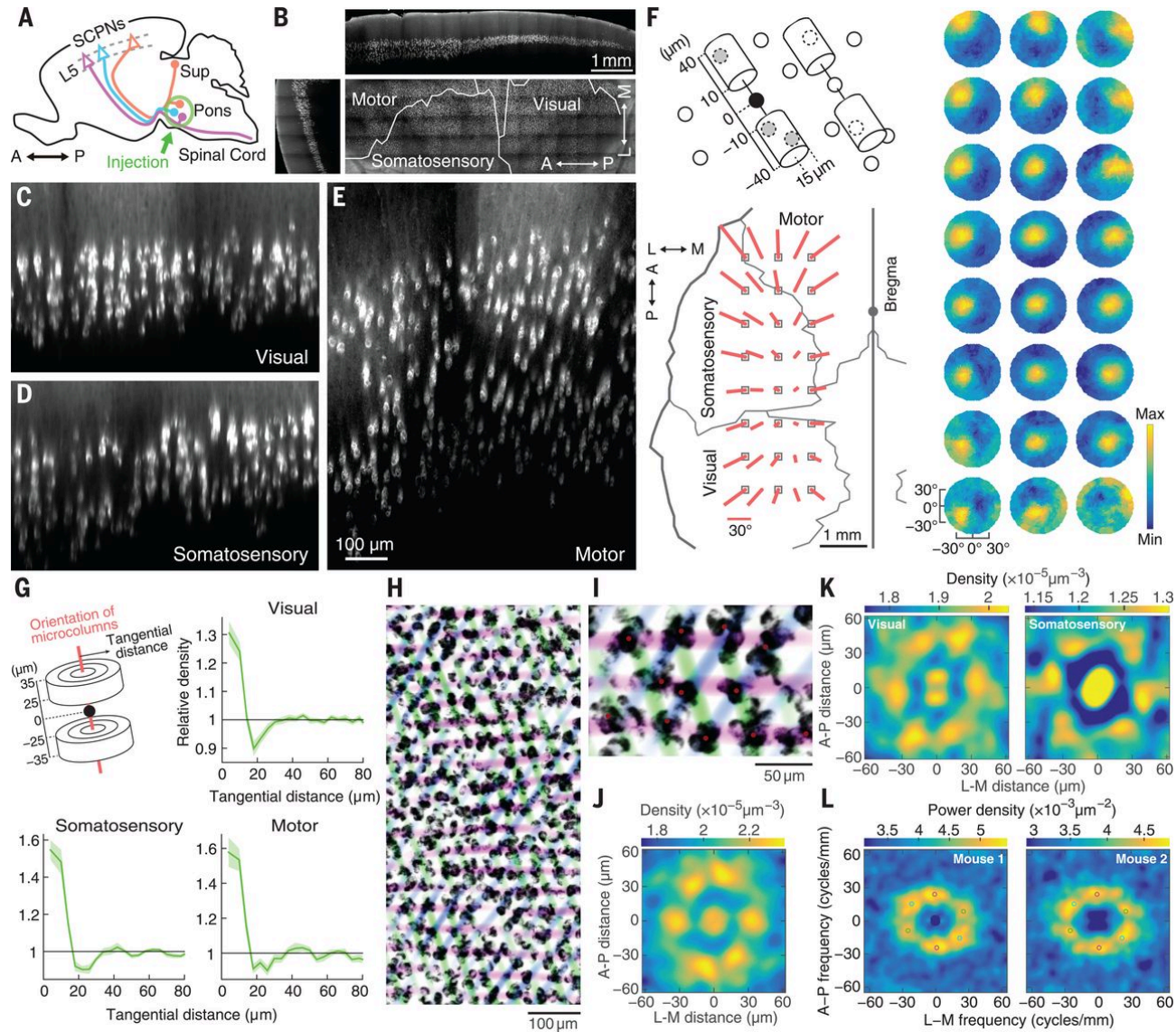
Periodic spacing of axon  
terminal clusters causes  
entrainment

~  $2 \times 10^7$  connections to  
macrocolumn



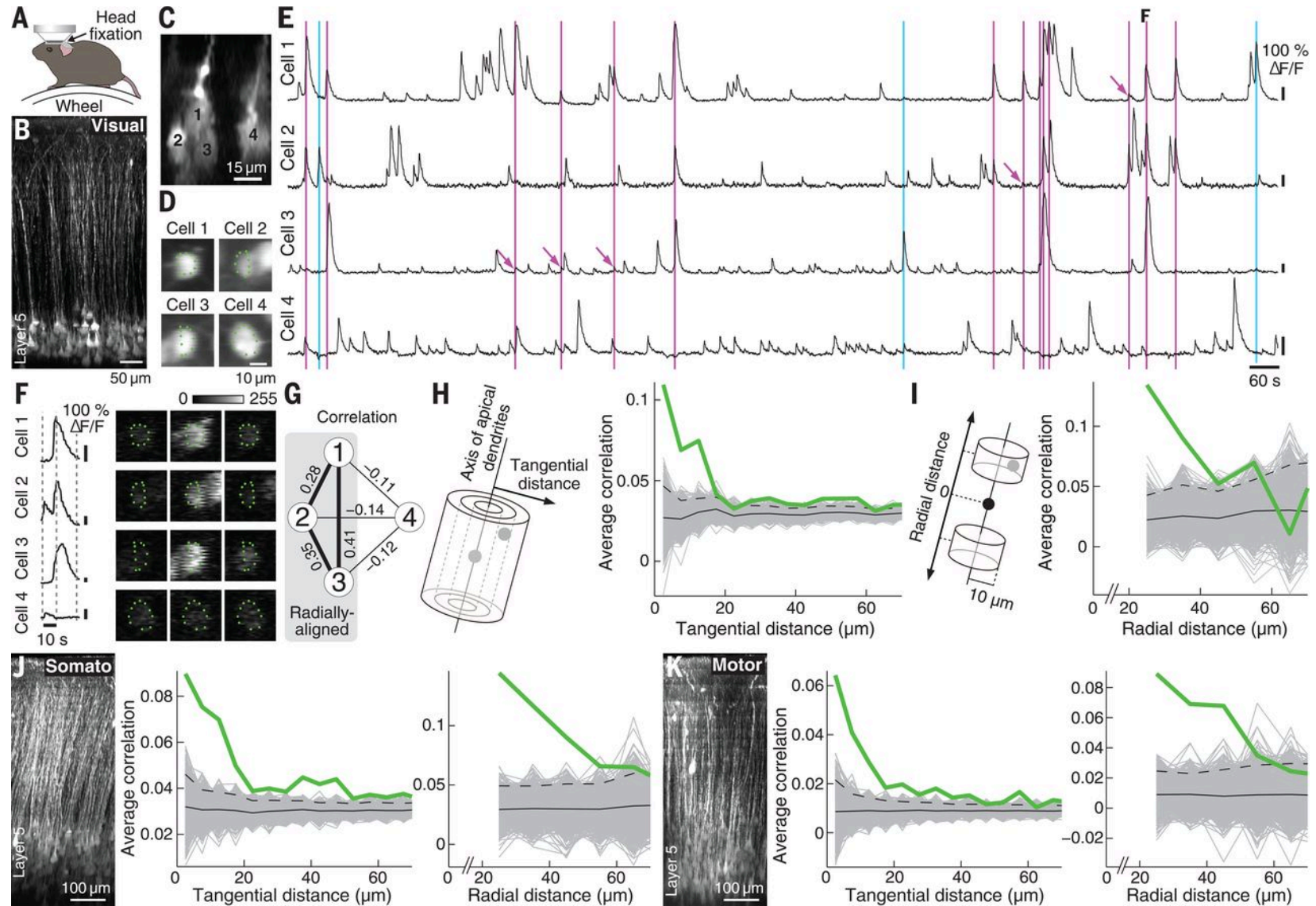


**Fig. 1 Lattice organization of SCPN microcolumns.**



Hisato Maruoka et al. *Science* 2017;358:610-615

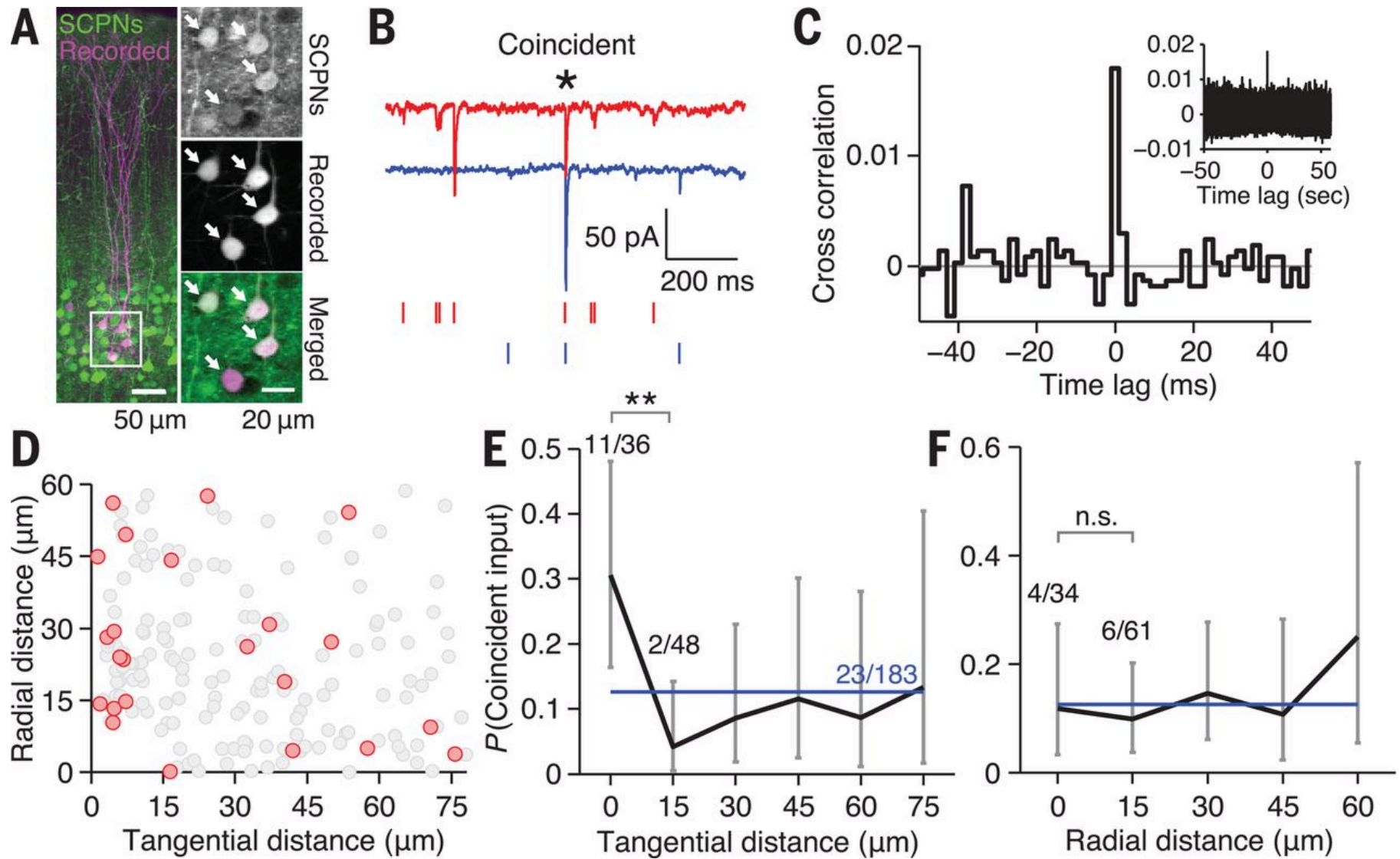
**Fig. 3 Synchronized activity in SCPN microcolumns.**



Hisato Maruoka et al. *Science* 2017;358:610-615



**Fig. 5 Convergent strong inputs to SCPN microcolumns.**



Hisato Maruoka et al. *Science* 2017;358:610-615

# Rate Code Approximation

- $g_e^\theta$  is the conductance when  $V_m = \theta$
- Rate is a nonlinear function of relative conductance
- What is  $f$ ?

$$\theta = \frac{g_e^\theta E_e + g_i E_i + g_l E_l}{g_e^\theta + g_i + g_l}$$

$$g_e^\theta = \frac{g_i (E_i - \theta) + g_l (E_l - \theta)}{\theta - E_e}$$

$$y = f(g_e - g_e^\theta)$$

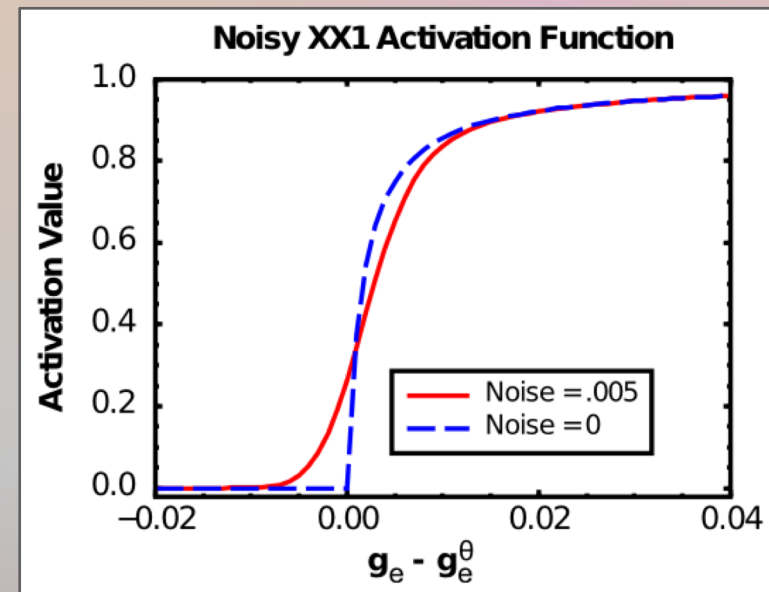
# Activation Function

- Desired properties:
  - threshold ( $\sim 0$  below threshold)
  - saturation
  - smooth
- Smooth by convolution with Gaussian to account for noise
- Activity update:

$$y_{t+1} = y_t + C(y - y_t)$$

$$y = \frac{x}{x+1} \quad \text{where } x = \eta [g_e - g_e^\theta]^+$$

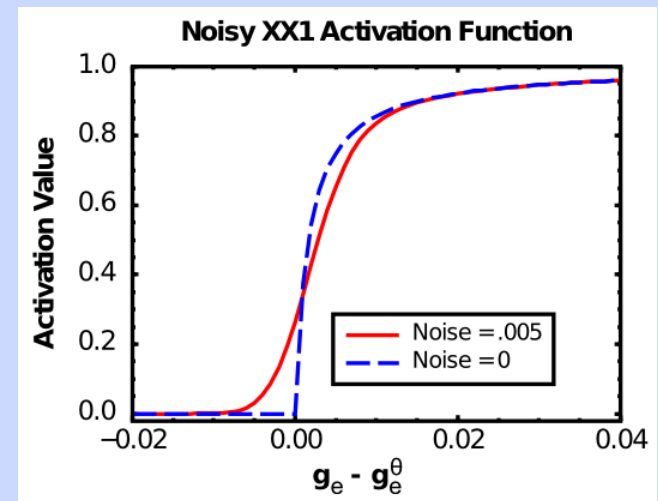
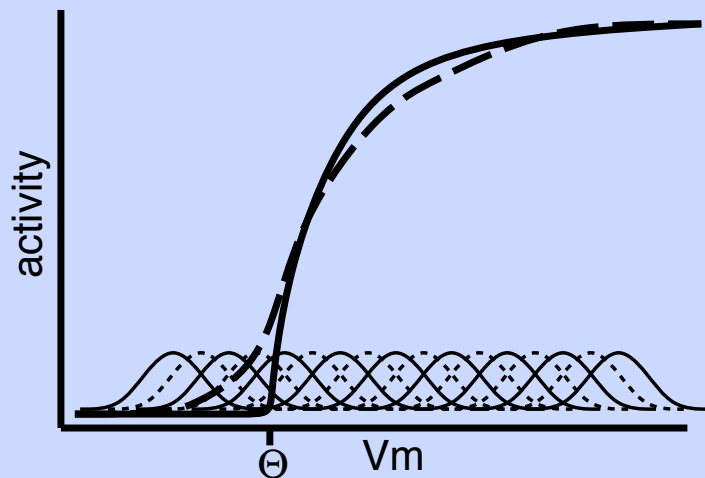
$$y = \frac{1}{1 + \frac{1}{\eta [g_e - g_e^\theta]^+}}$$



# Gaussian Smoothing

X-over-X-plus-1 has a very sharp threshold

Smooth by *convolve* with noise (like “blurring” or “smoothing”):



$$y^*(x) = \int_{-\infty}^{\infty} \frac{1}{\sqrt{2\pi}\sigma} e^{-z^2/(2\sigma^2)} y(x-z) dz$$

(slide based on Frank)

COSC 421/521



# Approximating Continuous Dynamics

- $V_m$  changes gradually when input changes
- Firing rate  $y(t)$  should also change gradually (subject to a time constant)
- Discrete-time update equation:

$$y(t) = y(t - 1) + dt_{vm} (y^*(x) - y(t - 1))$$

# emergent demonstration: Neuron

# Supplementary: Mathematics of Action Potentials

# Neural Impulse Propagation

$$C \frac{dv}{dt} = I - g_{Na} m^3 h (V - V_{Na}) - g_K n^4 (V - V_K) - g_L (V - V_L)$$

$$\frac{dm}{dt} = a_m(V)(1 - m) - b_m(V)m$$

$$\frac{dh}{dt} = a_h(V)(1 - h) - b_h(V)h$$

$$\frac{dn}{dt} = a_n(V)(1 - n) - b_n(V)n$$

$$a_m(V) = .1(V + 40)/(1 - \exp(-(V + 40)/10))$$

$$b_m(V) = 4 \exp(-(V + 65)/18)$$

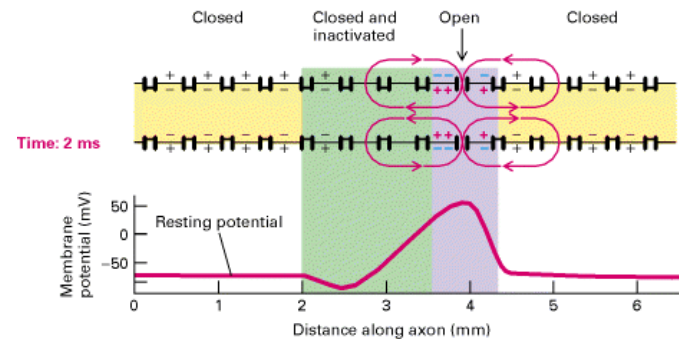
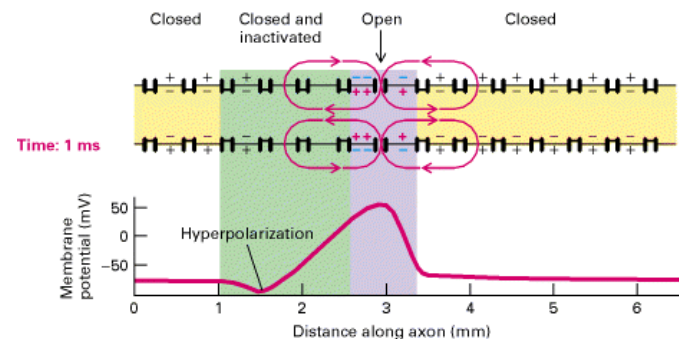
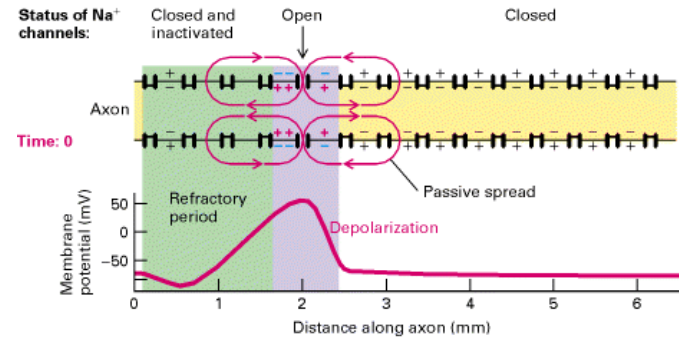
$$a_h(V) = .07 \exp(-(V + 65)/20)$$

$$b_h(V) = 1/(1 + \exp(-(V + 35)/10))$$

$$a_n(V) = .01(V + 55)/(1 - \exp(-(V + 55)/10))$$

$$b_n(V) = .125 \exp(-(V + 65)/80)$$

## Hodgkin-Huxley equations





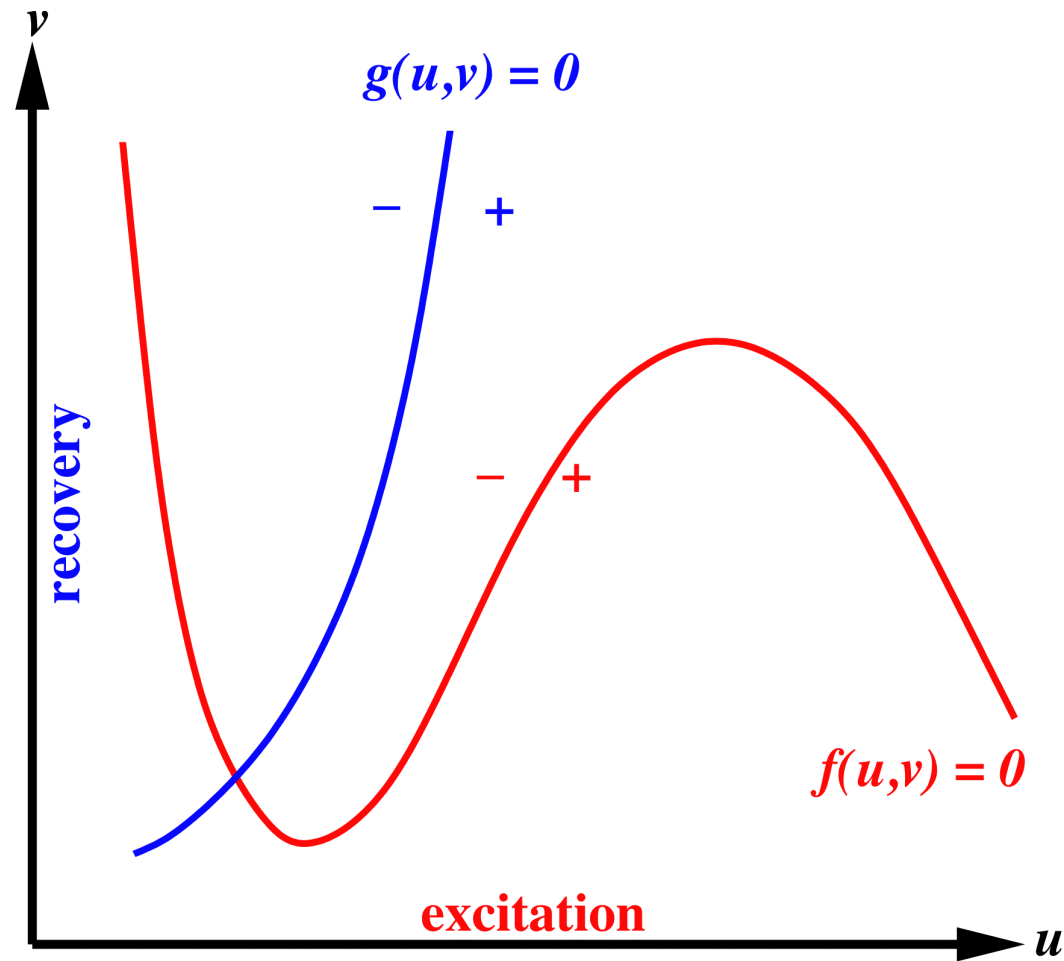
# FitzHugh-Nagumo Model

- A simplified model of action potential generation in neurons
- The neuronal membrane is an excitable medium
- $B$  is the input bias:

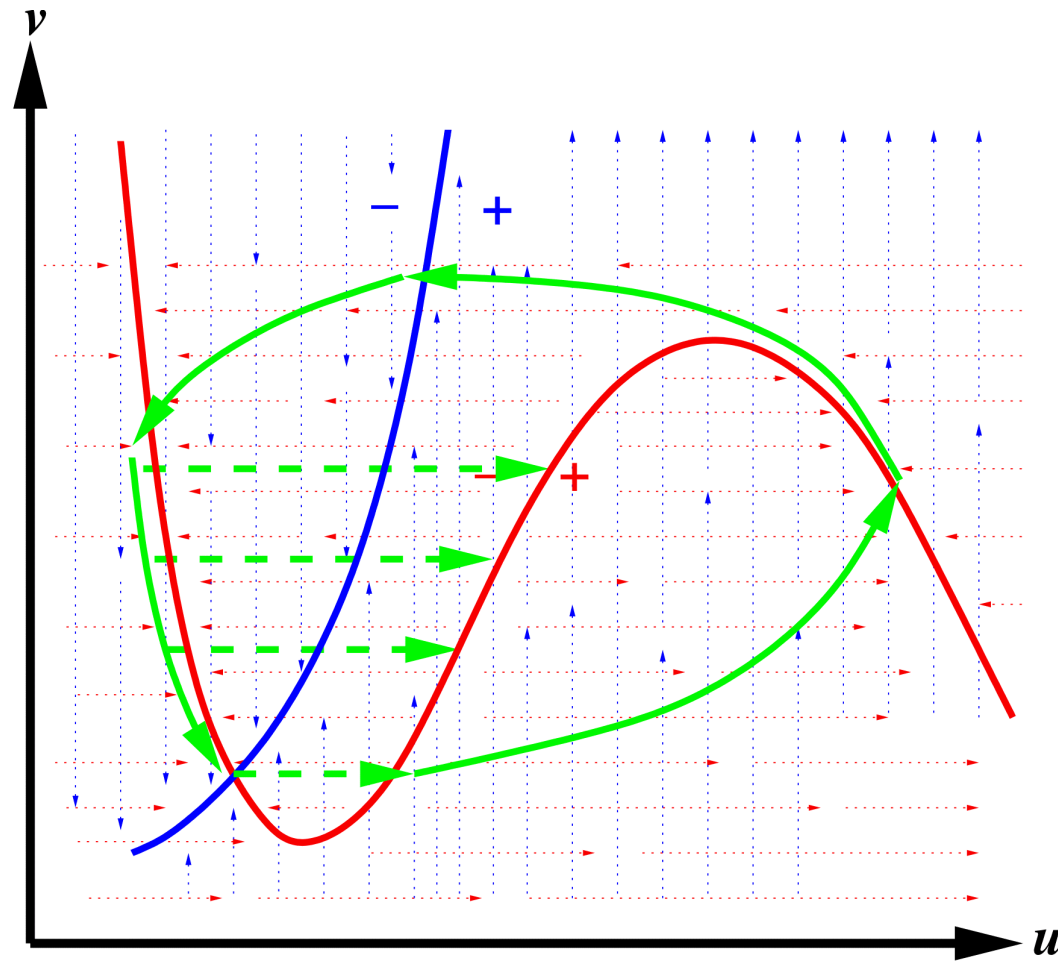
$$\dot{u} = u - \frac{u^3}{3} - v + B$$

$$\dot{v} = \varepsilon(b_0 + b_1 u - v)$$

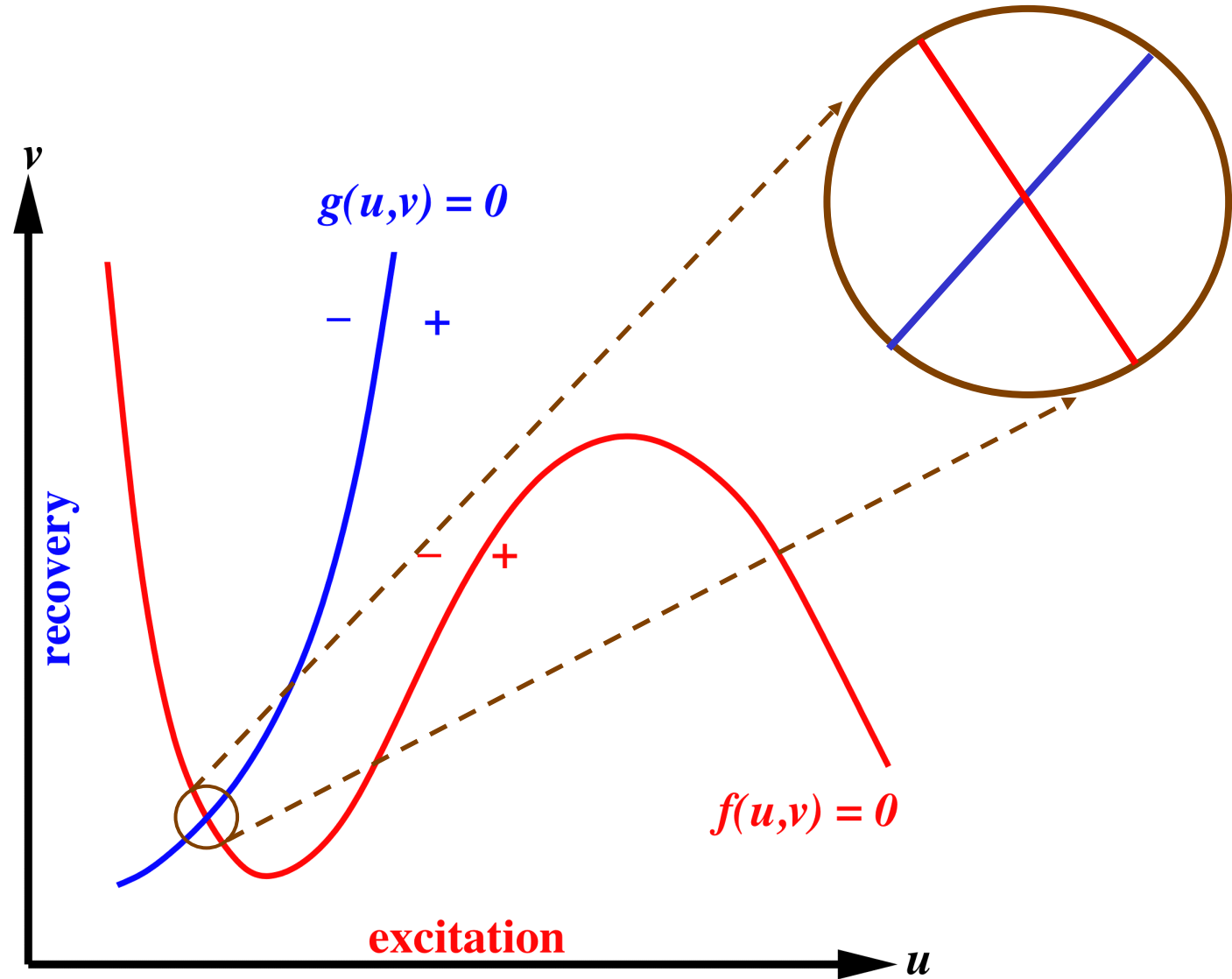
# Nullclines



# Elevated Thresholds During Recovery

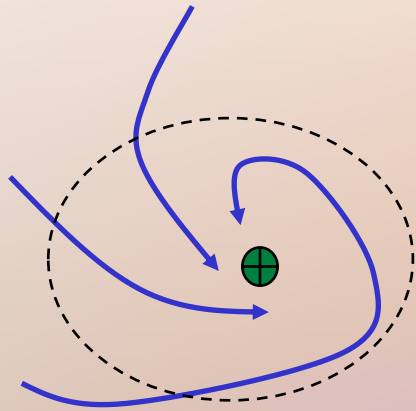


# Local Linearization



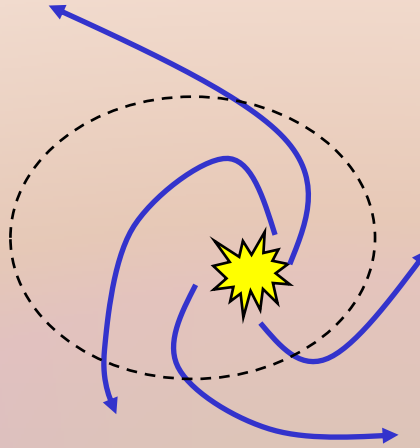


# Fixed Points & Eigenvalues



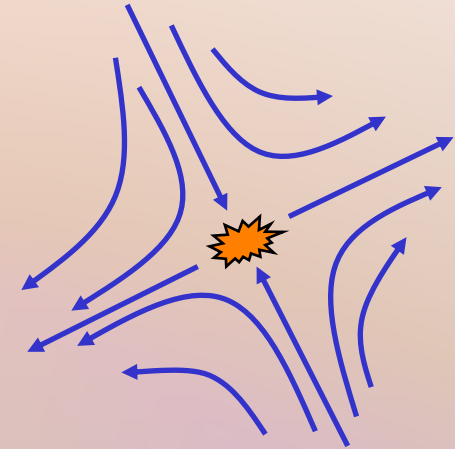
**stable  
fixed point**

real parts of  
eigenvalues  
are negative



**unstable  
fixed point**

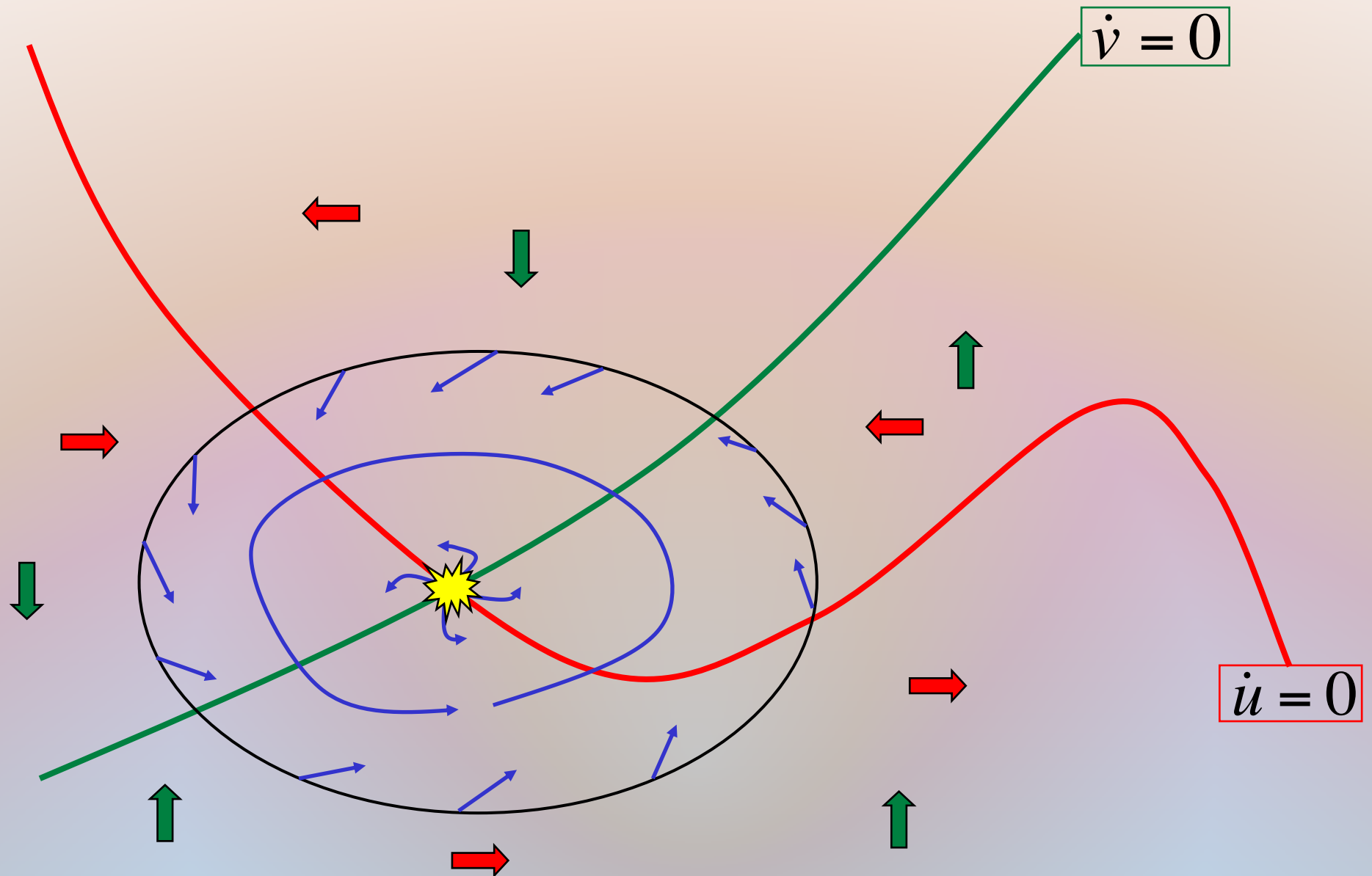
real parts of  
eigenvalues  
are positive



**saddle point**

one positive real &  
one negative real  
eigenvalue

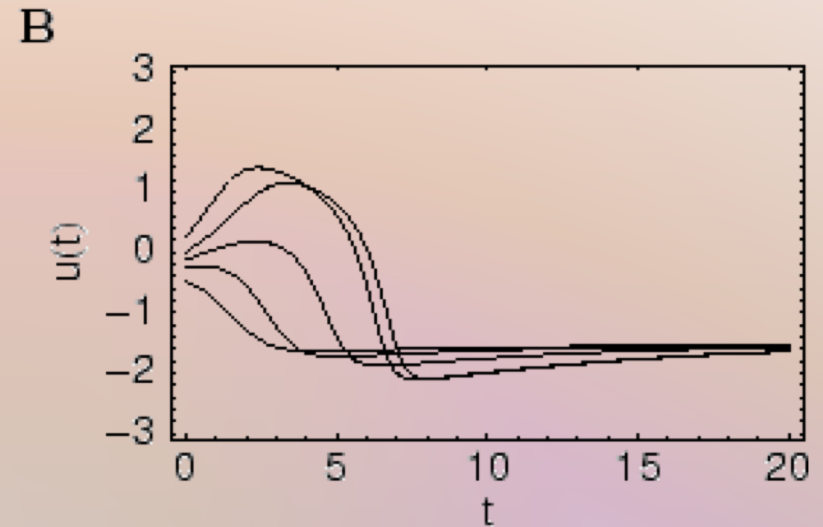
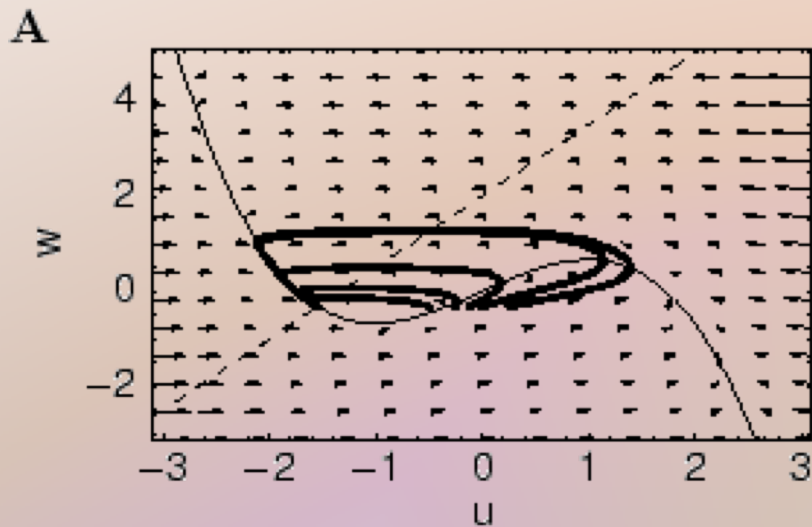
# Poincaré-Bendixson Theorem



NetLogo Simulation of  
Excitable Medium  
in 2D Phase Space

(EM-Phase-Plane.nlogo)

# Type II Model

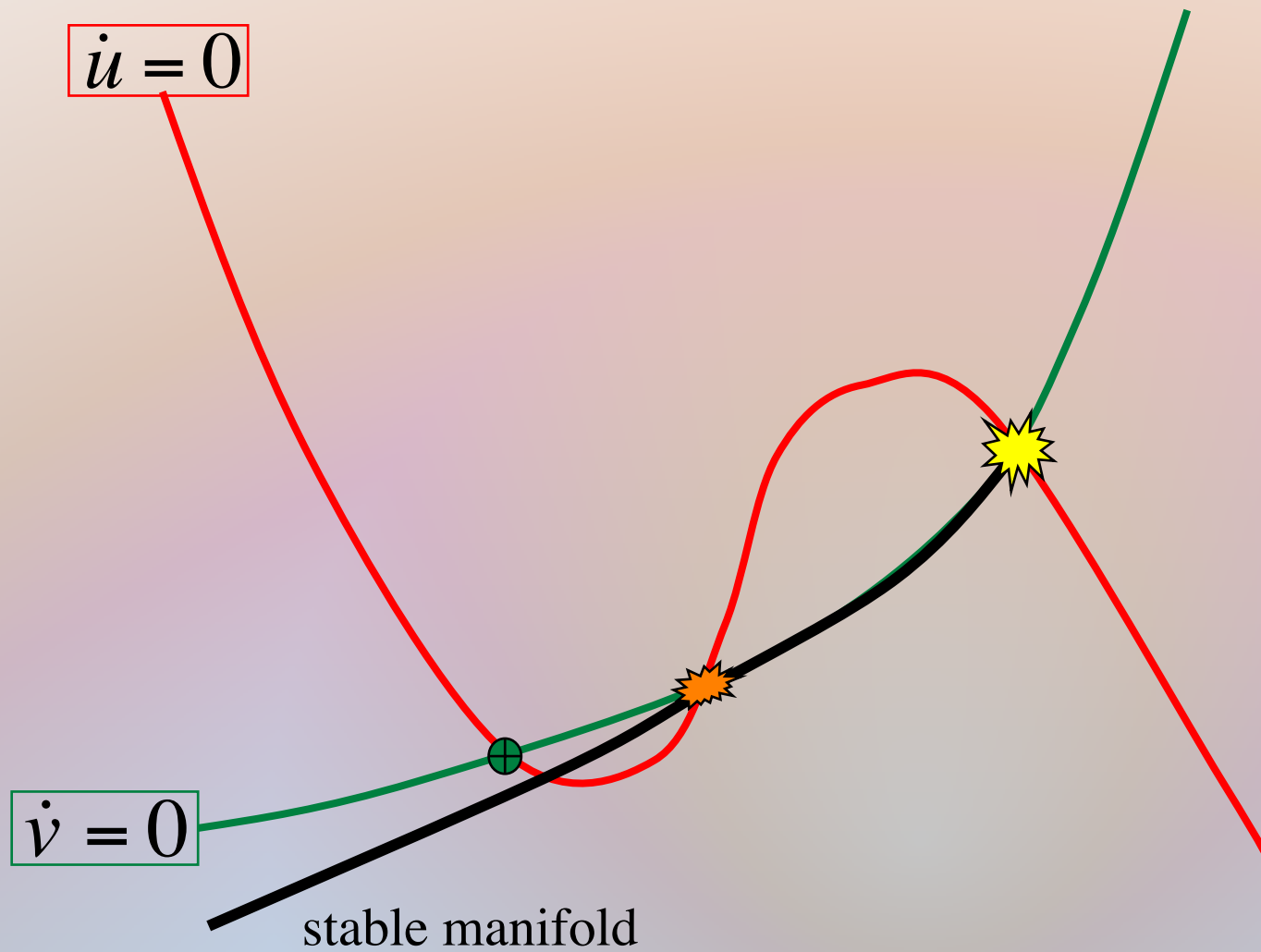


- Soft threshold with critical regime
- Bias can destabilize fixed point

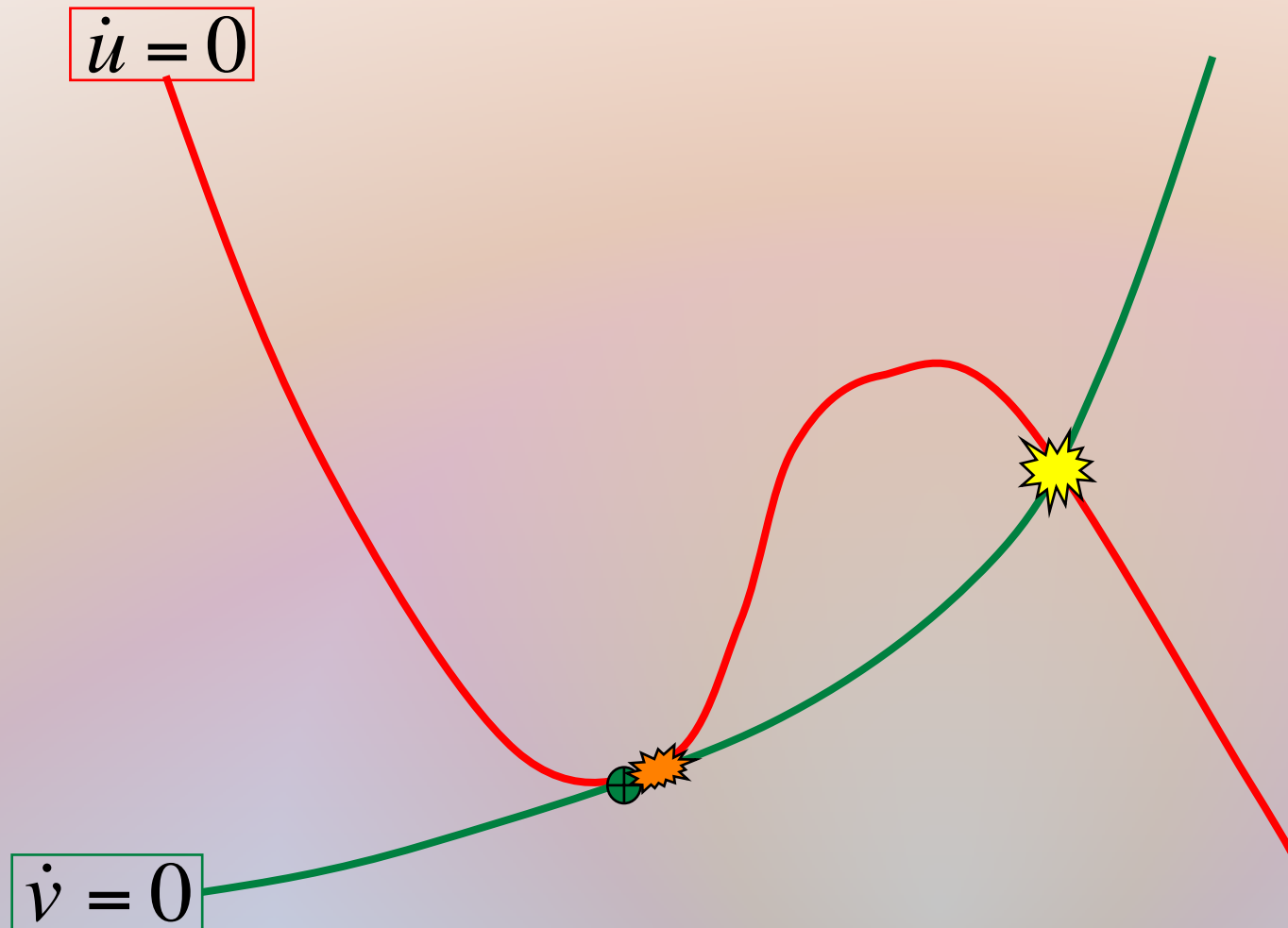
(figs. < Gerstner & Kistler)



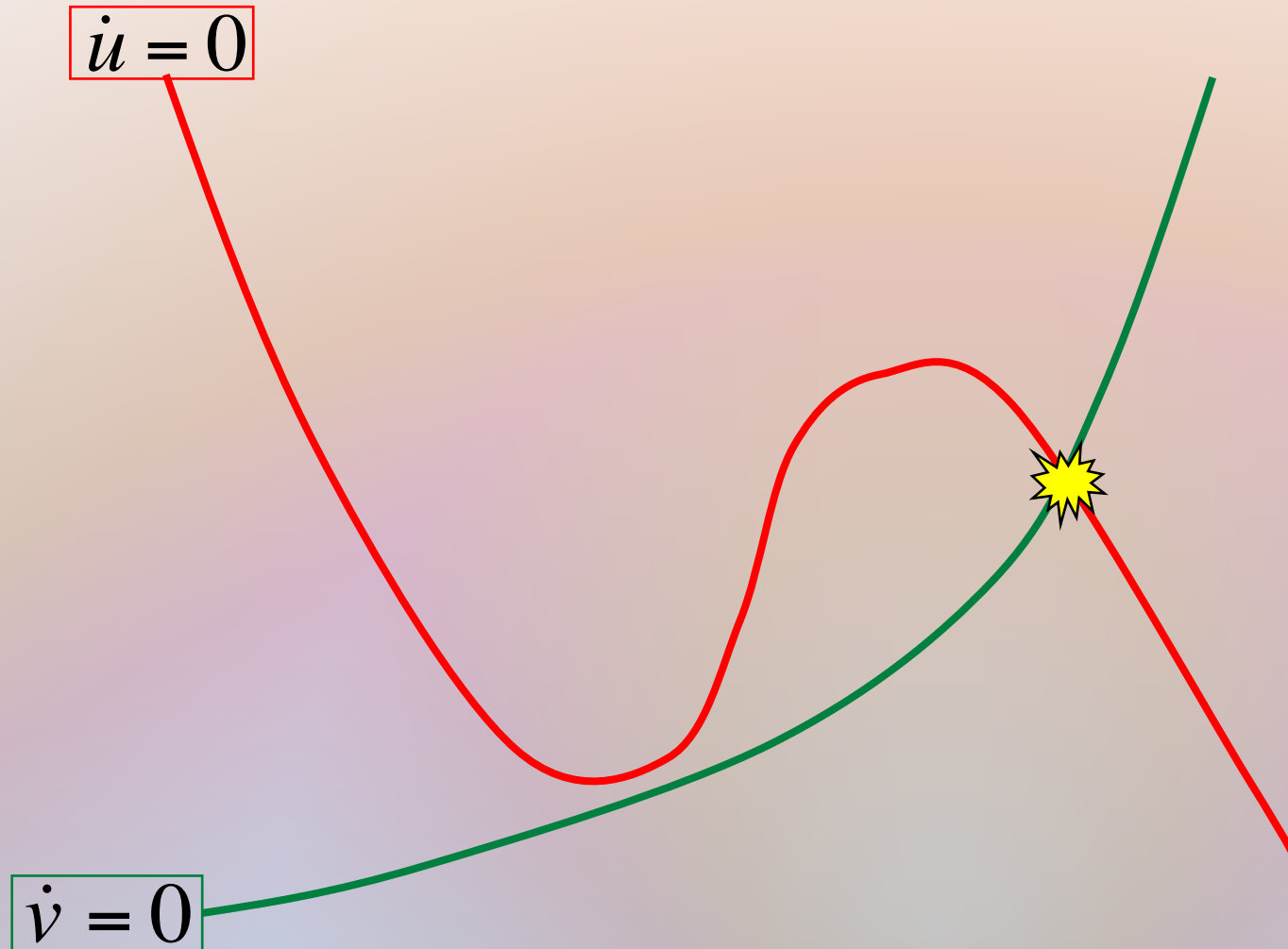
# Type I Model



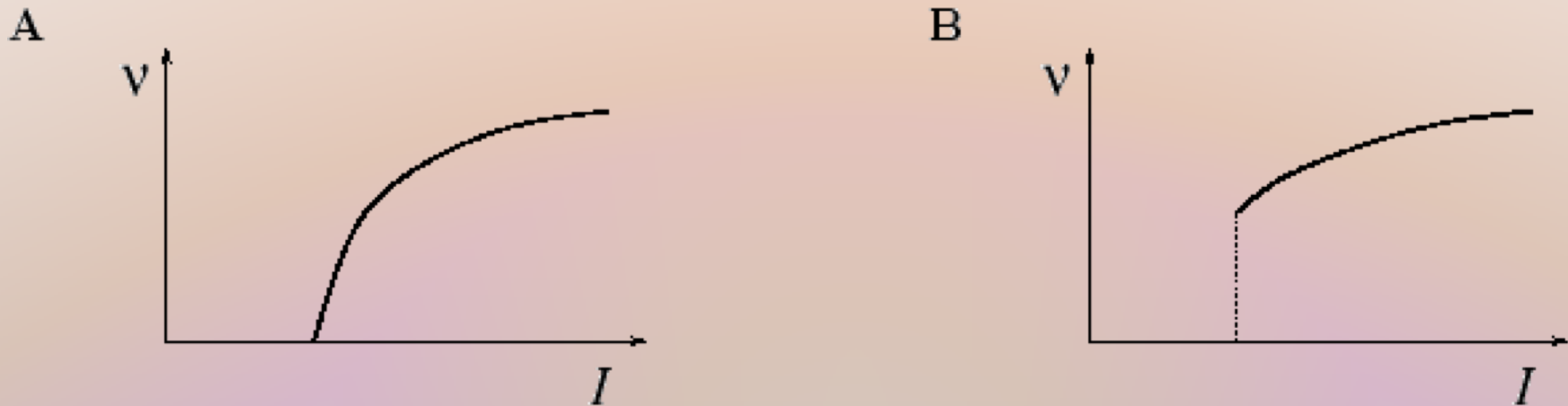
# Type I Model (Elevated Bias)



# Type I Model (Elevated Bias 2)



# Type I vs. Type II



- Continuous vs. threshold behavior of frequency
- Slow-spiking vs. fast-spiking neurons

fig. < Gerstner & Kistler